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## Background and Aim

The **pig** is a very convenient animal for different pre-clinical applications. Their **physiological similarities to humans**, particularly in terms of **cardiovascular systems**, make them the ideal **models for various biomedical research studies** [1].

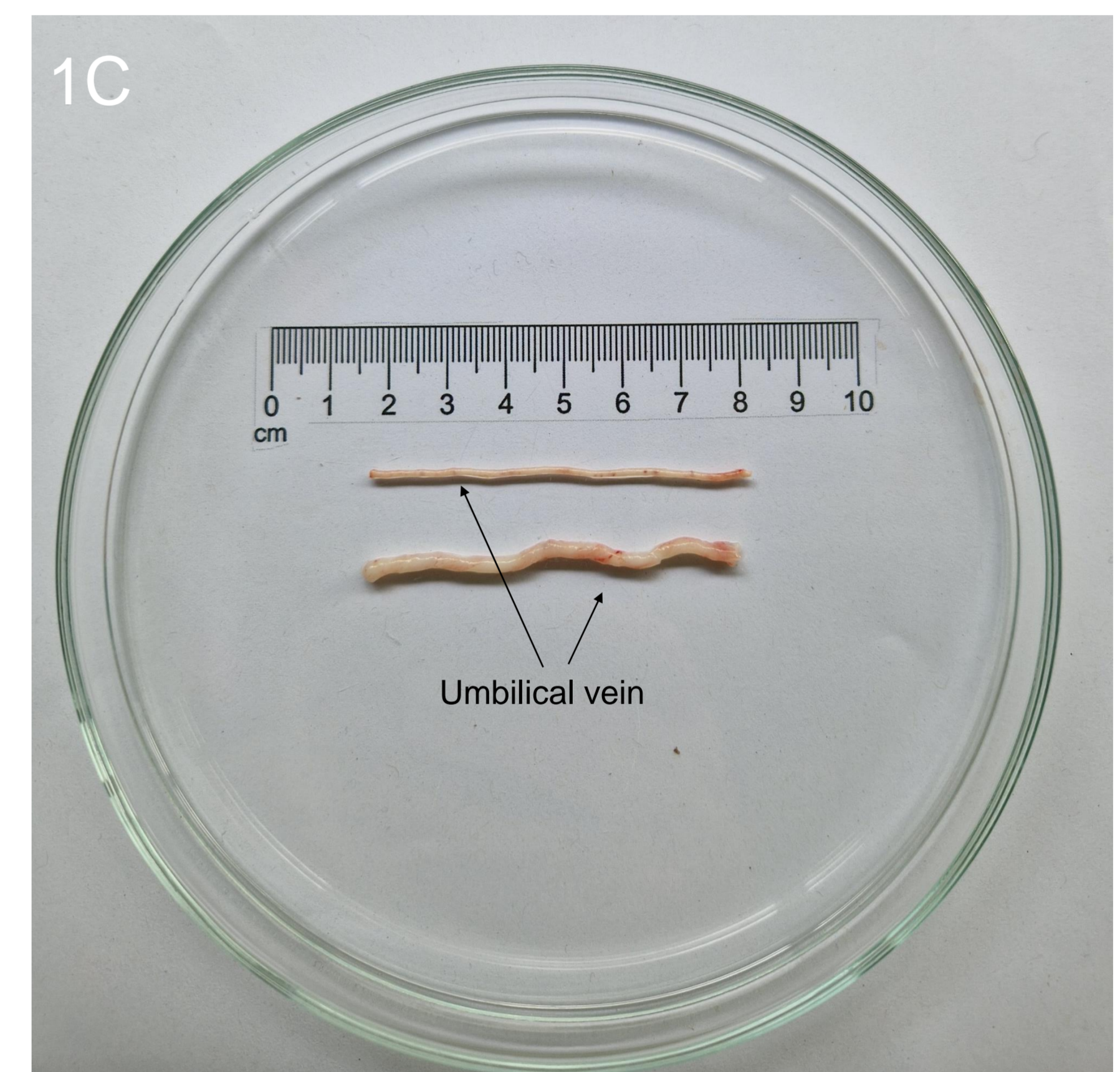
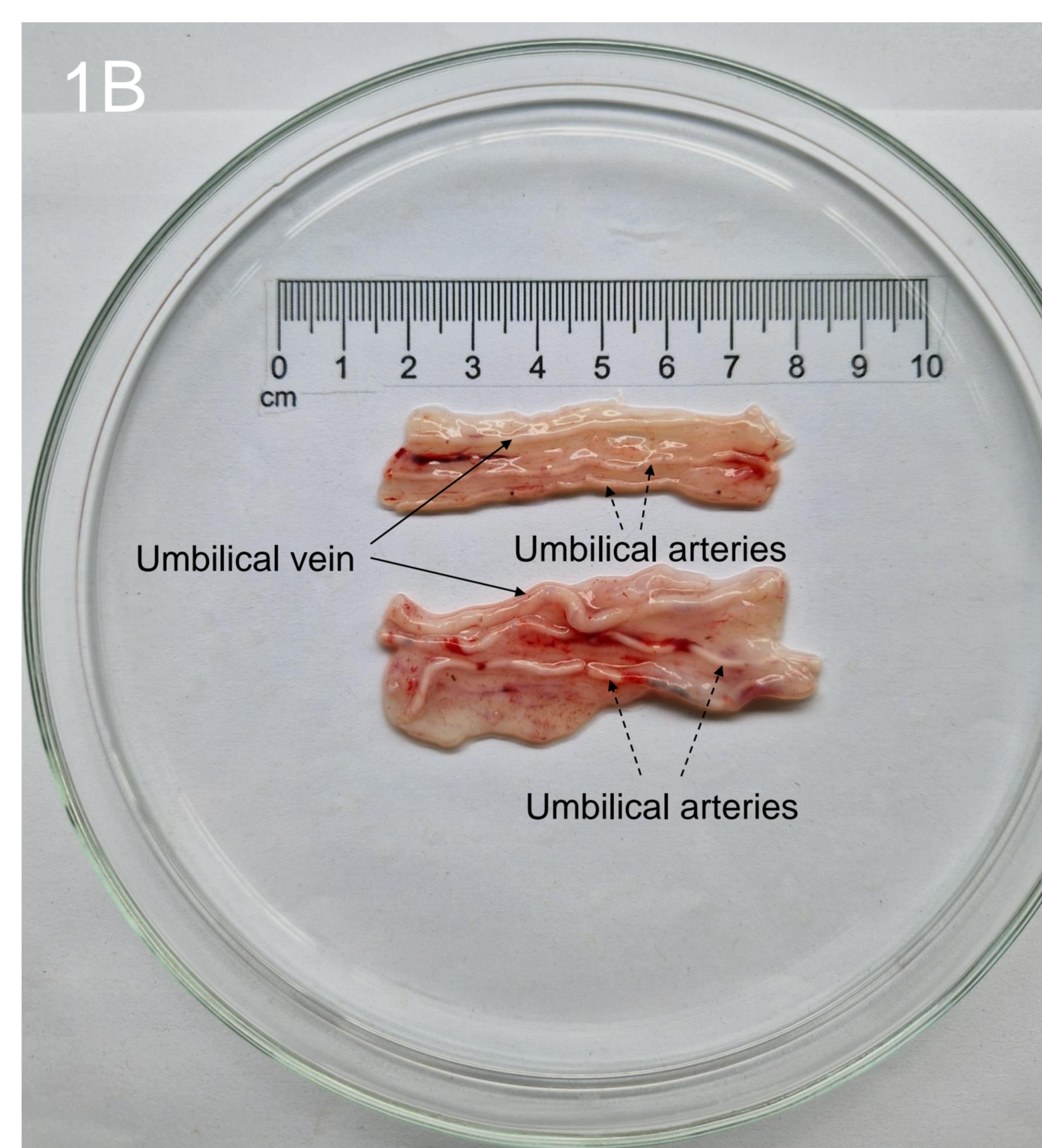
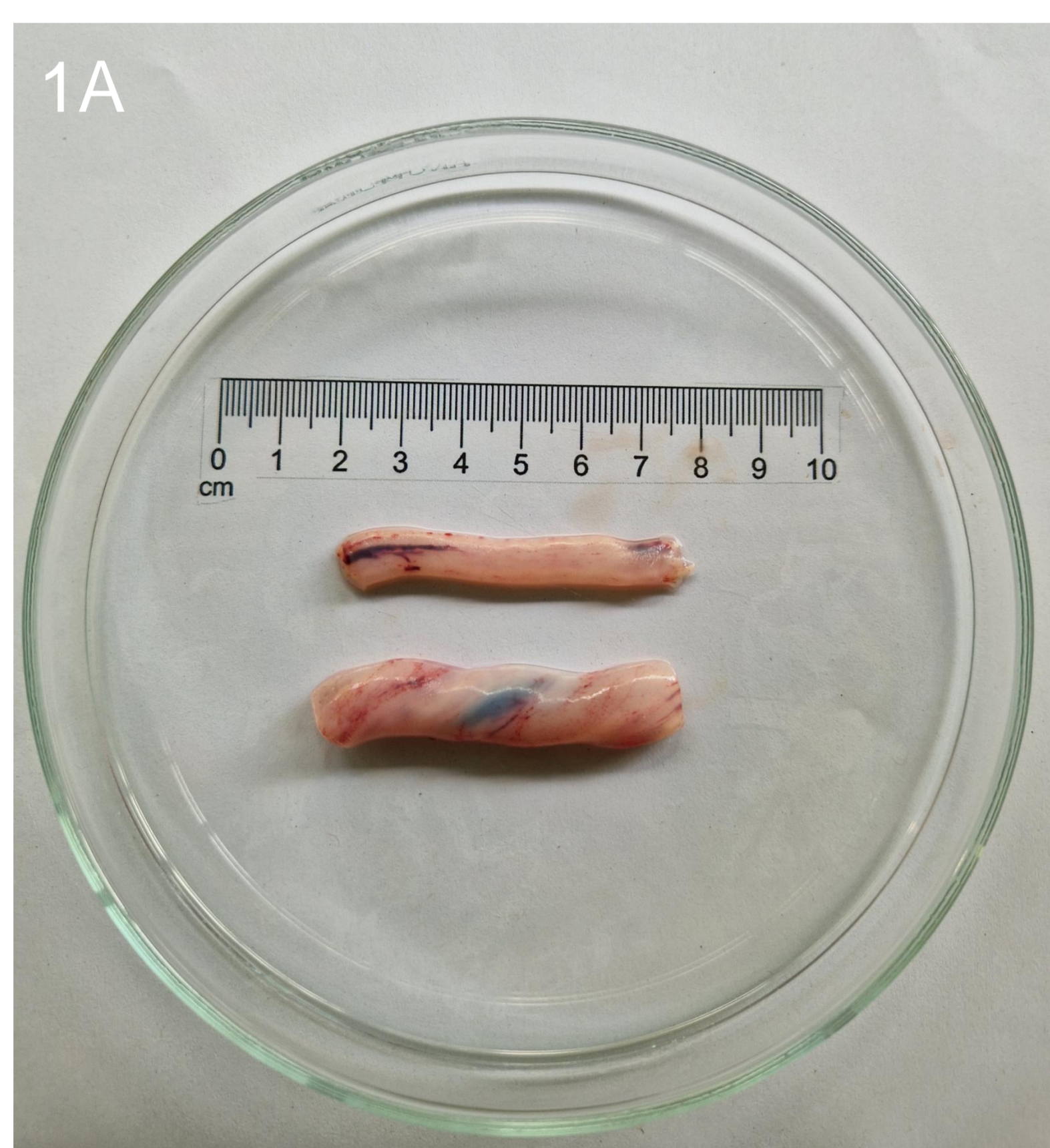
This similarity extends to processes such as **angiogenesis**, where pigs serve as effective models for studying vascular growth and development, critical for understanding and treating cardiovascular diseases [2]. Therefore, establishing **porcine umbilical vein endothelial cell (PUVECs)** lines is not just a need but a pressing requirement [3].

**Sex as a biological variable** has been absent from most *in vitro* work [4]. Reporting the sex of biological material is critical for transparency and reproducibility in science and should be consistently reported. Sex chromosomes in cells **can potentially affect protein expression and molecular signaling pathways** [5].

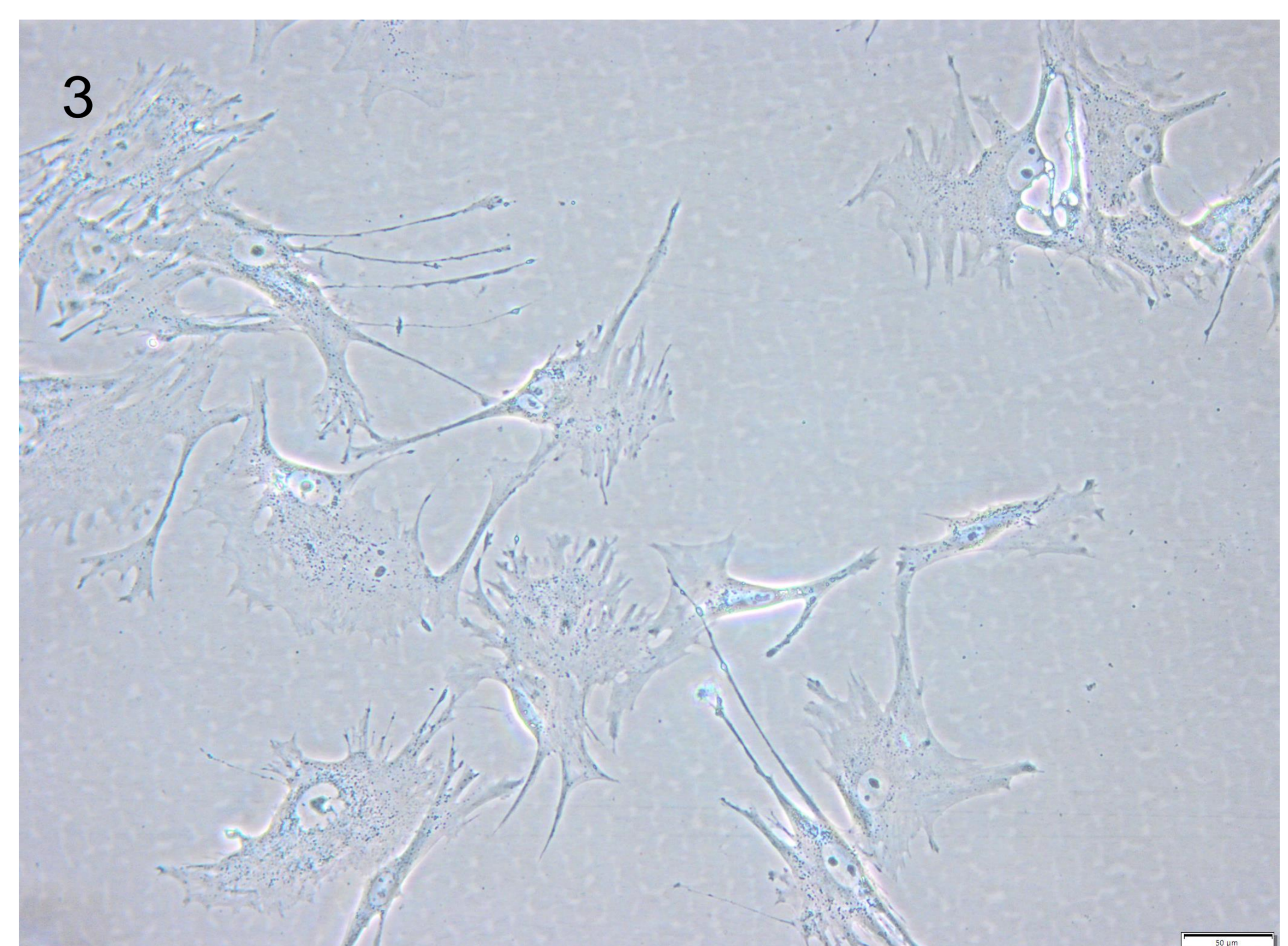
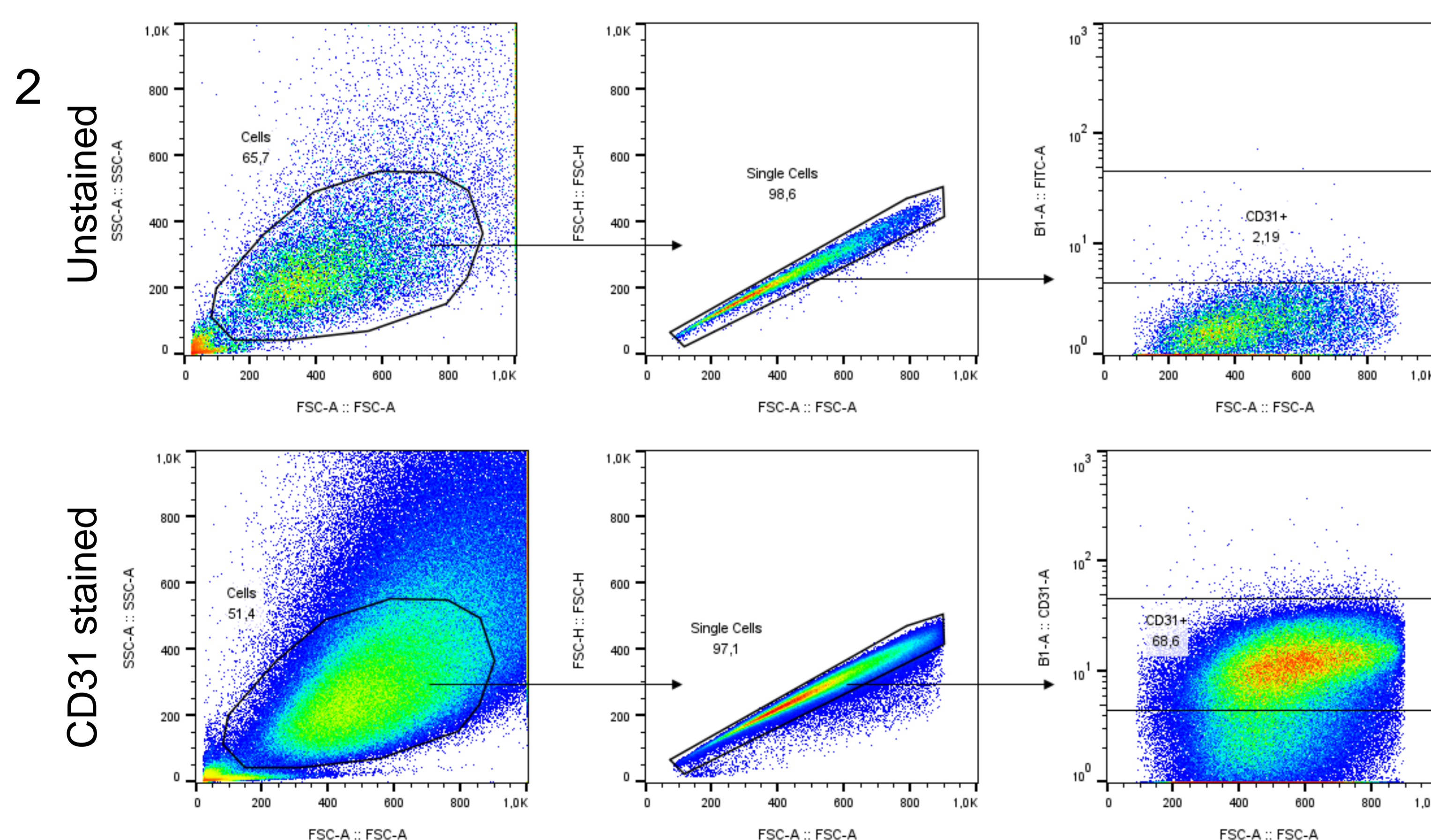
**This study aims to set-up a PUVEC cell line with a discrimination between sexes in order to evaluate the angiogenesis capacity of both sexes.**

## Materials and Methods

1. Umbilical cords (Fig. 1A) from both sexes were collected from pigs immediately after birth.
2. Within 6 hours after harvesting, umbilical cords were dissected (Fig. 1B), the umbilical vein (Fig. 1C) was identified and placed with the lumen towards the bottom of a 0.1% gelatine in DPBS coated 24-well plate.



3. Cells migrated from the harvested tissue towards the 24-well plate while incubating at 38°C at 5% CO<sub>2</sub> in a M199/RPMI1640 (50/50) medium supplemented with 5% FBS, 1% L-glutamine, 1 mM Na-pyruvate, 1% Penicillin-streptomycin, 1% NEAA, 15 mM HEPES, 0.5% endothelial supplement.
4. When a monolayer of cells was grown, we selected PUVECs from this co-culture using CD31, an endothelial cell marker, via fluorescence-activated cell sorting with a FITC-CD31 monoclonal antibody (Fig. 2). After sorting, PUVECs were cultivated and evaluated for their typical morphology *in vitro* (Fig. 3).



## Results and Conclusions

Collection and **cultivation of PUVECs** as a primary cell line from umbilical veins is **challenging** since cells grow **very slowly** ( $\pm$  50 days). Furthermore, when a monolayer of cells is grown, a **co-culture** of different cells is created. Fluorescence-activated cell sorting (using CD31) showed that  $53.7 \pm 10.3\%$  of the cells in culture were CD31<sup>+</sup> cells.

## Future perspectives

The next step would be to perform a **tube formation assay** to assess the **angiogenesis capacity** of PUVECs from different sexes. These results will give us more **insight into possible sex differences in cardiovascular diseases**.

## References

- [1] Jia *et al.* (2024) [4] James *et al.* (2021)  
[2] Nowak-Sliwinska *et al.* (2018) [5] Vallabhajosyula *et al.* (2020)  
[3] Chrusciel *et al.* (2011)