



Capturing the heterogeneity of the PDAC tumor microenvironment: novel triple co-culture spheroids for drug screening and angiogenic evaluation

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Dye: Blue - 9 peaks - DG_Ruben_BxPC_3-009-015.fsa

Supplementary Figure S1. Short tandem repeats (STR) profile of BxPC-3.



Dye: Blue - 9 peaks - DG_Ruben_HMEC_1-F09-005.fsa

Supplementary Figure S2. STR profile of HMEC-1.



Dye: Blue - 9 peaks - DG_Ruben_hPSC21-D09-003.fsa

Supplementary Figure S3. STR profile of hPSC21.





Supplementary Figure S4. STR profile of MiaPaCa-2.



Dye: Blue - 8 peaks - DG_Ruben_RTL_PSC-B09-001.fsa

Supplementary Figure S5. STR profile of RLT-PSC.



Supplementary Figure S6. Initial optimization process of triple co-culture spheroids. Representative images of (a) MiaPaCa-2:hPSC21 (1:1) spheroids which are not forming a single spheroid, and (b) MiaPaCa-2:hPSC21 (1:2) as a single spheroid. (c) Example of the masking of cancer (red), stellate (green) and endothelial (blue) cells using the Orbits image analysis software to quantify each cell type proportion on MiaPaCa-2:hPSC21:HMEC-1 (5:12:8) and (d) the original processed image of the Spark® Cyto (Tecan).



Supplementary Figure S7. MCDB131 is the most suitable medium for spheroid culture. Proliferation rate comparison of MiaPaCa-2, BxPC-3, RLT-PSC, hPSC21, and HMEC-1 in DMEM and MCDB131 is showed based on the confluence fold ratio normalized to day 1. Data are represented as mean \pm SD (n \ge 10 from three independent experiments). Statistics were performed using two-way ANOVA with Sidak's multiple comparison test using Prism v10.1.0 (GraphPad Software, San Diego, CA, USA). * = p \le 0.005; **** = p \le 0.0001; ns = not significant.



Supplementary Figure S8. DMEM with VEGF/FGF supplements is not able to provide sufficient growth factors for HMEC-1 growth. Proliferation rate comparison of HMEC-1 growth in MCDB131 vs DMEM vs DMEM with supplements, based on green fluorescence intensity fold ratio as a measure of confluency normalized to day 1. Data are represented as mean \pm SD (n \geq 9 from three independent experiments).



Supplementary Figure S9. Flow cytometry gating strategy. (a) Singlets, (b) cells, (c) double negative population for GFP and CD31, (d) positive population of the mKate2 (Cancer). (e) FMO control for CD31 on BxPC-3:RLT-PSC:HMEC-1 spheroids. (f) FMO control for CD31 on MiaPaCa-2:hPSC21:HMEC-1.



Supplementary Figure S10. Quantitative flow cytometric data of triple co-culture spheroid ratios that were not selected as optimal. Data shows the relative contribution of each cell population (each bar represents one flow cytometry measurement of a pool of 48 spheroids). Examples of TCC spheroids that were not selected due to an inaccurate representation of PDAC tumors.



Supplementary Figure S11. H&E and immunohistochemical staining of representative PDAC samples illustrating typical characteristics of the PDAC. (a) H&E staining of a PDAC sample at the invasive front of the tumor (black line). Note the abundant amount of desmoplastic stroma surrounding neoplastic ductular structures around the invasive carcinoma (left, black arrows). (b) Immunohistochemical staining for alpha smooth muscle actin (α -SMA) of the same tumor as shown in (a), illustrating myofibroblastic differentiation of the tumor stroma. (c) Different tumor composed of perineural invasion (orange horizontal arrow), isolated tumor cells (white vertical arrows) and poorly formed ducts (black horizontal arrows) embedded in abundant desmoplastic stroma consisting of activated fibroblasts and myofibroblasts. (d) Immunohistochemical staining for ERG, a nuclear marker of endothelial cells (brown), of the same tumor as shown in (c), illustrating sparse vasculature consisting of thin-walled capillary-size vessels with scanty luminal diameter (black arrows).