# Low-Temperature Plasma for Biology, Hygiene, and Medicine: Perspective and Roadmap

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Abstract-Plasma, the fourth and most pervasive state of matter in the visible universe, is a fascinating medium that is connected to the beginning of our universe itself. Man-made plasmas are at the core of many technological advances that include the fabrication of semiconductor devices, which enabled the modern computer and communication revolutions. The introduction of low temperature, atmospheric pressure plasmas to the biomedical field has ushered a new revolution in the healthcare arena that promises to introduce plasma-based therapies to combat some thorny and long-standing medical challenges. This article presents an overview of where research is at today and discusses innovative concepts and approaches to overcome present challenges and take the field to the next level. It is written by a team of experts who took an in-depth look at the various applications of plasma in hygiene, decontamination, and medicine, made critical analysis, and proposed ideas and concepts that should help the research community focus their efforts on clear and practical steps necessary to keep the field advancing for decades to come.

*Index Terms*—Bacteria, cancer, cold plasma, decontamination, plasma medicine, radicals, reactive species, wound healing.

## I. INTRODUCTION

THE MULTIDISCIPLINARY field of the biomedical applications of atmospheric pressure low-temperature plasma (AP-LTP) has been 25 years in the making [1]–[12]. Since its introduction in the mid-1990s, the field has expanded tremendously and has reached several critical and very important milestones, which include clinical trials on wound healing and cancer treatment. These advancements and successes have been accomplished, thanks to the outstanding contributions of many investigators and research groups from around the world. A quarter century may seem like a long time for an individual investigator but for a typical field of science, it is a relatively short period of time. Yet, the advances and accumulation of new knowledge about the interaction of plasma with living

Manuscript received November 25, 2021; accepted December 5, 2021. Date of publication December 14, 2021; date of current version February 3, 2022. The work of Annemie Bogaerts and AY was supported by the Research Foundation—Flanders (FWO) under Grant 1200219N. (*Corresponding author: Mounir Laroussi.*)

This work did not involve human subjects or animals in its research.

Please see the Acknowledgment section of this article for the affiliations.

Color versions of one or more figures in this article are available at https://doi.org/10.1109/TRPMS.2021.3135118.

Digital Object Identifier 10.1109/TRPMS.2021.3135118

soft matter have been nothing short of impressive. Today, our scientific knowledge concerning the physical and biochemical pathways involved in the interaction of plasma with biological cells and tissues has reached a good level of maturity. However, more remains to be done both on the fundamental and application levels. So, 25 years seems like a good time marker for the research community to take a good look and evaluate the current state of the field, identify the challenges and obstacles, and propose ideas on how these could be overcome. These are the main objectives of this roadmap paper. A team of experts in the various subtopics of the field has been tasked to each summarize the present status of their subtopic, inform the research community about the remaining challenges, and propose potential approaches and methods that can be employed to overcome these challenges.

Low-temperature plasma (LTP) produces a rich "cocktail" of reactive species, including reactive oxygen species, ROS, and reactive nitrogen species, RNS, which have been shown to play pivotal roles in the observed biomedical outcomes [13]. LTP sources also exhibit high electric fields and produce photons at various wavelengths/energies, which can also play synergistic biological roles. The state of knowledge regarding the effects of these plasma-produced agents is quite advanced as of the present, but many of the effects on the subcellular and molecular levels are either not fully understood or a matter of debate. To help elucidate these effects, advanced modeling and simulation approaches have been brought to bear on many of these issues [14]–[17]. In addition, in the aim of making LTP biological effects reproducible under similar operating conditions, it is important to develop control methods that allow for the minimization of the effects of environmental conditions, such as humidity, temperature, air pressure (e.g., altitude), power fluctuations, etc., while at the same time optimizing the treatment as a function of the properties of the target (shape, surface morphology, thickness/depth, conductivity, etc.). These are some of the outstanding issues that are discussed in this article. Ideas and new concepts to tackle these challenges are proposed, providing an outlook on what still needs to be done and where the field is headed in the near future.

The remainder of this article is organized in seven sections. The first section discusses LTP sources. The application of plasma in biology and medicine would not have been

2469-7311 © 2021 IEEE. Personal use is permitted, but republication/redistribution requires IEEE permission. See https://www.ieee.org/publications/rights/index.html for more information. possible without the development of atmospheric pressure LTP (AP-LTP) sources. However, there are still outstanding issues, such as the control of these sources to achieve optimal performance for any application they are used for. The second section discusses the application of LTP in biological decontamination. This was the first bioapplication of LTP and the initial seed that was at the genesis of the entire field. This application has proved to be of the utmost importance, because of the advent of antibiotic resistance of bacteria and pathogenic proteins (such as prions), and because of viraldriven pandemics, such as the present Coronavirus disease 2019 (COVID-19) pandemic. The third section discusses the use of LTP to treat water and air. Plasma treatment of water is used for nitrogen fixation, reduction of pollutants, and inactivation of pathogens in the food cycle, food safety, and for water purification. For air treatment, traditionally plasma has been used for the inactivation of airborne pathogens. For example, heating, ventilation, and air conditioning (HVAC) systems can be retrofitted with a plasma treatment stage to purify indoor air from biological contaminants. However, with the advent of viral-driven pandemics (such as COVID-19), plasma treatment of air has taken a renewed importance. The fourth section discusses LTP-assisted wound healing. This was the first patient-centered medical application of LTP and maybe one of the most developed and has undergone extensive animal studies followed by clinical trials at various hospitals, especially in Germany. One of the therapeutic challenges is the healing of chronic wounds. Chronic wounds, such as diabetic ulcers, do not heal easily or at all. Thousands of amputations occur every year in the U.S. alone, because of the inability of conventional methods to satisfactorily deal with this problem. LTP has been shown to safely coagulate blood and enhance the proliferation of cells such as fibroblasts, which made gas plasma an attractive technology for wound healing. The fifth section deals with the application of LTP in the field of dentistry. Here, LTP has been used in three areas of dentistry: 1) periodontal; 2) endodontic; and 3) prosthodontic treatments. LTP has also been used "indirectly" for surface modifications of implants, oral surgery to improve interfacial bonding strength of dental objects used in periodontics or prosthodontics, and sterilization of dental instruments. The sixth section discusses cancer treatment by LTP, what is referred to as "plasma oncology." This application has seen great progress in the last decade that recently culminated in preliminary clinical trials in a few hospitals in Germany and the USA. In vitro investigations have shown that LTP can kill cancer cells and *in vivo* experiments have shown that LTP treatment can shrink tumors size and/or delay their growth. However, there is still much work to be done to understand the effects of LTP on cancer cells on the subcellular and molecular levels and the effects on the tissues surrounding tumors. Finally, the seventh section of this article discusses modeling and simulation works that have been conducted so far to further elucidate the plasma-cell and plasma-tissue interactions. The interaction of LTP with entire cells or tissue can up to now only be simulated on a macroscopic scale, obviously providing less detailed information than modeling the interactions with individual cell components, such as the phospholipid

bilayer (PLB), DNA, or proteins, which is possible on the molecular level. Much progress has been made in recent years, but more work has to be done, and major challenges are the compromise between calculation time and level of detail of the simulations, the need for accurate input data and experimental validation, and the translation of the obtained insights toward improving the biomedical applications.

To conclude, the research on the biomedical applications of LTP has, first, produced new, previously missing, fundamental scientific knowledge regarding the interaction of the fourth state of matter with soft biological matter. Second, LTP is proving to be a technology that can help meet many of today's healthcare challenges, including the inactivation of antibiotic-resistant bacteria, the healing of chronic wounds, treatment of various dermatological diseases, and even some types of cancers. More recently, agricultural applications have also emerged, which include treatment/decontamination of fruits and legumes for longer shelf life, the modification of the wettability of the surfaces of seeds, and the enhancement of the germination speed and yield of plant seeds. Finally, LTP may also be a viable technology for space medicine. Here, plasma can replace perishable medication and can be used to decontaminate tools and gear from possible extraterrestrial microorganisms and to disinfect wounds and/or skin infections that astronauts may experience during long-duration space missions. To ensure that LTP technology can meet all the requirements of effectiveness and safety for the abovelisted applications, the research community needs to identify all the remaining scientific and technical challenges and seek creative solutions that can take the field to the next level. We hope that this article plays a role in this endeavor and provides some guidance for future investigations.

## II. LOW-TEMPERATURE PLASMA SOURCES S. REUTER, A. MESBAH, D. LIU, X. LU, M. LAROUSSI, AND K. OSTRIKOV

## A. Current State

Plasma Sources: Two main types of LTP sources have been used in plasma medicine research, dielectric barrier discharges (DBDs), and nonequilibrium atmospheric pressure plasma jets (N-APPJs) [18], of which many varieties exist. N-APPJs quickly became widely applied LTP sources because they can generate a stable and controllable plasma volume outside the confinement area of electrodes within the surrounding environment [19]. This is crucial for a direct plasma treatment of diseased tissue on a patient [20]. Biologic effects of LTP sources in medicine include sterilization/decontamination of biotic and abiotic surfaces, the promotion of growth factors, control of cell migration, induction of apoptosis, angiogenesis, microcirculation, and modulation of the immune response [4], [21], [22]. These effects have been attributed to the reactive oxygen and nitrogen species (RONS), electric field, charged particles, and ultraviolet radiation (UVR) generated by LTP [8], [13], [23]. Because LTP can help in preventing the aerosol transmission of respiratory infectious diseases, such as tuberculosis and the COVID-19 [24], [25]-[27], the recent research focus was set



Fig. 1. Schematics of LTP sources. Direct LTP sources. (a) Volume-air DBDs with a gas gap in the mm range. (b) Typical DBD plasma jets. Indirect LTP sources: (c) Air DBD was generated inside the ceramic tube. (d) Surface air DBDs. LTP sources inactivating bioaerosols: (e) Needle corona discharge array to inactivate Penicillium aerosols. (f) Air DBD system (quartz tube) to kill airborne bacteria or virus.

on plasma sources for the reduction of pathogenic biological aerosols (PBAs).

A second classification of biomedical LTPs is application mode oriented—two key types are distinguished: 1) direct and 2) indirect plasma sources [24]. For the first type, the plasma has direct contact to the treated surface, and the second type does not. In the case of direct LTP sources, the treatment target (skin, wound, tissue, etc.) acts as the counter electrode; therefore, the discharge current (displacement or conduction current) flows through the body. One example of direct LTP sources is shown in Fig. 1(a) [4], [28]. A typical plasma jet is shown in Fig. 1(b). A pin electrode covered by a dielectric tube with one end closed is the power electrode [29] while the ring electrode outside the tube is the ground electrode [30]. A commercial version of this type of N-APPJ is the kINPen [31]. More configurations of plasma jets can be found, e.g., in [19] and [29].

For indirect LTP sources, the plasma is mainly produced between the electrodes and some reactive species of the plasma can reach the surface of the target via the electric field propagation (ionization wave), convection, and diffusion mechanisms [29]. An air DBD-based indirect LTP source is shown in Fig. 1(c) [32]. A commercial version of this type of LTP is the PlasmaDerm device. Convection through the air flow delivers RONS generated by the plasma inside the tube to the treatment target. Another indirect LTP, the surface DBD (SDBD), is shown in Fig. 1(d). A thin plasma layer forms between the grounded electrodes' structures. The reactive species and UV light can interact with the object surface due to the short distance between the grounded electrode and the target's surface [21].

Recently, air plasma sources that can generate stable air discharges under conditions of high air flow rate have been used to sterilize PBAs. Fig. 1(e) shows the structure of bioaerosol disinfector based on an atmospheric-pressure positive corona discharge. Another example of DBD plasmas in air that can inactivate aerosolized *Bacillus subtilis* cells and *Pseudomonas fluorescens* indoor and outdoor is shown in Fig. 1(f) [33], [34]. It is important to note that the large plasma volume covers almost the entire cross section of the quartz tube, largely increasing the interaction intensity between the plasma and PBAs.

*Plasma Processes:* In the past decade, enormous progress has been made in the characterization of atmospheric pressure LTP sources. This can be attributed to the technology development of diagnostics, as well as to a deeper knowledge of plasma chemistry and processes through the combination of modeling and diagnostics. Through reference plasma sources [35] and a continuing development of high-precision diagnostic methods for absorption spectroscopy [36], emission and laser spectroscopy [37], and nonoptical methods such as mass spectrometry [38], a thorough understanding of the plasmas has been achieved [13], [37], [39], [40].

Most recent developments to understand plasma processes focus on electric fields in plasmas, electron properties, and energy dissipation pathways [41]. The emergence of ultrafast diagnostic techniques has enabled researchers to observe the generation of reactive species and space resolved electric field dynamics in plasma breakdown relevant time scales [42]. Finally, especially for the medical field, the interaction and transport processes of the plasma generated reactive species into liquid environments are fundamental [43]. A substantial progress has been made in the ability to control plasma activated solutions and using mixed-phase systems such as plasma-bubble systems to enhance the reactivity and other relevant physical and chemical progress. These advances have been made through the close interaction of physicists, engineers, and chemists.

*Plasma Control:* Control over plasma composition is mandatory for a targeted medical application. Tremendous progress has been made in controlling electron dynamics, e.g., through a multifrequency approach [44]. Ensuring a reproducible composition by using a tailored gas surrounding the active effluent has opened the door to several promising fundamental biomedical studies. Traditionally, control of LTPs has primarily relied on statistical process control strategies, as widely used in semiconductor manufacturing. However, such control strategies are inherently open loop, in which the lack of online sensing and feedback corrective action can compromise the reliability and repeatability of LTPs due to the intrinsic plasma variabilities and external disturbances. The key challenges in feedback control of LTPs stem from: 1) the nonlinear interactions between multiple input variables (i.e., discharge parameters) and multiple output variables of a plasma discharge (i.e., plasma effects); 2) the need to constrain the plasma properties to ensure safe plasma treatment and circumvent adverse plasma-induced surface effects; and 3) the need to realize the proper synergy between the chemical, physical, and electrical effects of LTPs when interacting with complex biological surfaces [45]. Recently, predictive control and learning-based control approaches have shown exceptional promise for safe, effective, and reproducible operation of N-APPJs [46]-[48]. Reduced-order, physics-based models and data-driven models have been used to develop model predictive control (MPC) strategies for constrained, multivariable control of the nonlinear effects of plasma on surfaces [46], [49]. Experimental studies using cell and chemical assays have correlated operational parameters of CAPs such that real-time monitoring of the plasma's chemical effects can be effectively utilized in an adaptive manner for feedback control [50]. These studies have been extended to develop MPC-based strategies for targeted cancer cell treatment using real-time electrochemical impedance spectroscopy measurements [51]. Furthermore, deep neural network-based controllers have been demonstrated to be particularly useful for the control of N-APPJs using resource-limited embedded systems, which are especially suitable for portable and point-of-use biomedical applications [52].

#### B. Challenges and Proposed Solutions

1) Challenges: Despite the fact that many LTP sources suitable for biomedical applications have been developed and successfully applied in vitro and in vivo, the number of clinical trials remains relatively small [53]. One of the crucial challenges relates to the controlled delivery of plasma agents to diseased targets. The challenges are similar to those of plasma processes in the semiconductor industry in the 1990s that arose due to the significant lack of knowledge about the fundamental mechanisms involved in the plasma-materials interactions. A concerted Research and Development effort in academia and industry allowed the identification of the plasma parameters and substrate properties and geometries that led to the desired manufacturing process. Dry etching by plasma thus became a commonplace technology in semiconductor production. The plasma medicine community is currently at a similar development stage with lessons to be learned from the semiconductor processing experience, which necessitated substantial numerical modeling assisted process analysis [54]-[56].

A stark difference between the technological and the medical application of LTPs is the certification procedures of medical plasma sources. While more and more plasmas are used in medical research and some countries have certified plasma sources for medical use, a clear path to bringing a new type of plasma source to clinical application remains to be found.

Although more progress in the field of new LTP sources and innovative biological applications are desirable, there are fundamentals issues that still need to be addressed. These can be subdivided into the three areas: 1) *plasma physics and chemistry*; 2) *biomedicine*; and 3) *LTP technology development*. The following questions require further investigations to be able to reach a knowledge-based plasma medicine approach.

*Plasma Physics and Chemistry:* How do electric field dynamics in N-APPJs lead to the generation of energetic electrons, ions, and RONS? What is the role of rotational-vibrational-translational energy transfer? How can the concentrations and fluxes of RONS be accurately controlled? How can plasma induced reactivity be delivered through different media without undue influence of environmental factors?

*Biomedicine:* What are the effects of treatment target properties and composition (skin, the wound covered by effusion, tumors, aerosols, etc.) on LTP characteristics? How do we standardize the flux and energy of LTP transmitted to cells and tissues to appropriately define "plasma dose"?

LTP Technology Development: How can we use artificial intelligence (AI), new materials, and new power electronics technology to optimize the design of the LTP sources and control the plasma? How do we ensure that LTP biomedical applications are patient compatible? How can LTP technology be designed sustainably to enable end-of-life recycling and reuse of the electrodes, electronic components, and other parts of the plasma devices? How can plasma sources be adapted to the respective treatment requirements and be safely applicable *in-vivo* at the tumor site also in nonsurgical procedures?

2) Proposed Solutions: LTP sources development is an interdisciplinary research area; therefore, collaborations between plasma physics and chemistry, power electronics, fluid dynamics, microbiology, and medical science are required (Fig. 2). In order to set up the roadmap for applied physics here, we concentrate on the challenges in the three areas mentioned above.

It is necessary to carry out experimental and simulation work to thoroughly elucidate and determine all the physical and chemical processes in a cohesive way. For example, in the case of N-APPJs, using fast imagery with high spatial resolution ( $\mu$ m range) and high time resolution (sub-ns range) along with 2-D or 3-D simulation models, the effect of photoionization, seed electrons, and electric field can be obtained.

To understand where and how the active constituents generated by the plasma, RONS, UV, charged particles, and electric fields are generated, their interactions, and interdependence is important. Measurement techniques, such as infrared absorption spectroscopy for radical and molecule detection, optical emission spectroscopy for discharge dynamics, Stark spectroscopy or electric field induced second harmonic generation for electric field, Thomson, Raman, and Rayleigh scattering for electron parameters, laser induced fluorescence (LIF) for radical and molecules detection, and mass spectrometry, can be employed. 2-D or 3-D simulations with hundreds of plasma reactions will facilitate the identification of the main production mechanisms of the plasma reactive species (see chapter on modeling).

When LTP sources are used to treat fluid-covered wounds or bioaerosols, plasma comes in contact with liquids. Therefore, the knowledge of the liquid-based chemistry will improve the



Fig. 2. Three key areas affecting the development of LTP sources—1) plasma physics and chemistry; 2) biomedicine; and 3) LTP technology development. The collaboration of multidisciplinary scientific community will enhance the understanding of fundamental mechanisms of operation of LTP sources. Feedback loops between the key areas include reactive species generation and delivery, protocols and adapted protocols, as well as the ETOP.

understanding of the discharge processes happening at the gas-liquid interface. The evaporation of liquid is an important process, because it not only affects the plasma chemistry but also decreases the lifetime of bioaerosols. On the other hand, when LTP is used to treat skin diseases such as psoriasis, the scurf or accretive tissue may decrease the delivery of plasma, making it necessary to adjust the plasma treatment dose.

Since the reactive components of the plasma consist of a large variety of species, it is difficult to standardize plasma treatments. A proposed metric is the equivalent total oxidation potential (ETOP) (see Fig. 2) that includes all RONS and UV/VUV generated by the plasma, which could be used to evaluate the plasma dose [57]. If the ETOP of a plasma source is known at different values of power, temperature, and humidity, this could facilitate the expansion and application of the LTP technology in medicine.

The plasma source development ranges from small- to largescale devices and the design is driven by the target application. In any case, moving toward mobile and personalized devices will greatly enhance application fields to include ambulant, point of care, or even in a home care environment. Batterypowered devices or low-power devices are promoting this development (e.g., [58]).

The rapid development of AI can be used to transform the mathematical modeling, real-time diagnostics, and optimal operation of LTP sources for applications in plasma medicine [59]. In addition, new oxidation-resistant and highstrength electrode materials, as well as new power semiconductor devices can enhance the efficiency of LTP sources and reduce the cost of their development and manufacturing. Environmental considerations should be taken into consideration to reduce emissions of harmful gases and meet personnel minimum exposure requirements of gases such as ozone.

Finally, the certification issues need to be addressed. The market is steadily growing for plasma medicine, but a standardized certification procedure is still missing, while first attempts have been made for a minimal characterization requirement [60]. Lacking certification procedures could ultimately pose a great risk to the field, due to insufficiently safe plasma sources entering the market.

## III. LTP FOR BIOLOGICAL DECONTAMINATION

## J. WALSH, M. LAROUSSI, M. KEIDAR, AND C. LAUX

#### A. Current State

The use of low-temperature plasma (LTP) for the decontamination of biological agents has been under intensive investigation for several decades. Commercial LTP decontamination systems, employing low-pressure plasma, are nowadays widely used in healthcare settings across the world for the sterilization of temperature-sensitive medical devices. While low pressure LTP systems are undoubtedly efficient for biological decontamination, the added cost and complexity of a vacuumbased process limit their broader appeal. At the end of the 20th century, interest in the use of plasma for biological decontamination was reignited with a demonstration that LTP generated under ambient pressure conditions could be harnessed for the nonthermal inactivation of microorganisms [1]. Much of this early work focused on the use of noble gas plasma jets due to their ease of use and convenience [19], with more efficient and inevitably more complex LTP-based decontamination devices being developed over the subsequent years to meet a growing number of application needs.

As the area of research continues to expand, it is becoming more diverse and varied. While early works focused primarily on the use of LTP to eliminate planktonic bacteria, recent efforts have focused on more representative contamination, such as antibiotic resistant bacteria [61], mixed species biofilms [62], and endospores [63]; conditions that present a formidable challenge to any nonthermal decontamination technology. Beyond bacterial contamination, the global SARS-CoV-2 pandemic has placed a spotlight firmly on the need to develop fast, efficient, and low-cost viral decontamination techniques that can be easily deployed beyond a healthcare setting. Given the minimal equipment costs and the possibility of consumable-free operation of LTP, many members of the plasma science community have turned their attention to viral decontamination. These efforts have resulted in a number of published studies within the past 12 months that comprehensively demonstrate the efficiency of LTP against viral agents [64], [65], a clear indication that the plasma science community is both flexible to adapt and willing to tackle some of the greatest challenges facing humanity.

## B. Challenges and Proposed Solutions

1) Current and Future Challenges: Underpinning the use of LTP for biological decontamination is the multiphase interaction between a nonequilibrium gas plasma and a target organism; given the shear complexity of this interaction, a plethora of scientific challenges remain unresolved. These challenges essentially fall into distinct areas, such as the need to understand and control fundamental LTP processes, and the need to understand the biological effects of LTP exposure at



Fig. 3. Diagram showing the priority decontamination targets for LTP and challenges relating to the understanding and control of LTP for effective, efficient, and repeatable biological decontamination.

the target (Fig. 3). Adding to the complexity is the two-way interaction between the LTP and its target, often giving rise to emergent and synergistic behaviors that are difficult to predict, characterize, and control.

Perhaps, one of the greatest contemporary challenges associated with LTP mediated decontamination relates to unraveling the underpinning mode of action that determines the efficacy of the process; a challenge that is further exasperated by the large number of different plasma systems currently under investigation, with each system presenting different operating characteristics and parameters, combined with the vast number of biological agents being explored [63]. Great progress has been made in this area over the past decade, with increasingly sophisticated techniques being adopted to gain new levels of insight. Despite this progress, more needs to be done, especially at the biomolecular level. Critically, understanding the mode of action is only the first step in developing an effective LTP decontamination system. Being able to manipulate and control the fundamental LTP processes to elicit a desired biological response at the target remains the "holy grail" in the area, yet such control extends beyond our current capabilities. A key advantage of the LTP technology is its ability to be applied directly at the point of need; in many situations, this is likely to be beyond the laboratory in an environment that

is in a constant state of flux. Given that much of the reactivity of an LTP system can be derived from its interactions with the surrounding environment, be it from air entrainment within a plasma jet plume [66], or through the direct generation of LTP in ambient air [67], the question of how to generate a consistent discharge composition in a constantly changing environment remains unresolved.

While understanding and controlling LTP processes is the key to ensure efficiency and repeatability of the decontamination processes, many challenges relating to the underpinning biomolecular processes must also be addressed. Typically, these challenges are highly specific to the biological target.

Bacteria and Biofilm Formation: In the natural environment, microorganisms cluster together and form a protective layer as a means of self-preservation; this community is known as a biofilm. Typically, microorganisms encased within a biofilm are 1000 times more resistant to antimicrobial agents than their planktonic counterparts. When LTP interacts with a biofilm, the antimicrobial agents created must first penetrate this protective layer, known as the extracellular polymeric substances (EPS). The complex composition of the EPS acts to inhibit the transport of reactive agents to the encased microorganisms, significantly diminishing decontamination efficacy. Biofilms also present ideal conditions for the formation of antimicrobial resistance, one of the greatest challenges facing humanity in the 21st century. Indeed, the key questions of if, how and how quickly microorganisms form resistance to LTP remain unanswered.

Viruses: The use of LTP to decontaminate viruses has been under active investigation for several years, yet it was not until the emergence of SARS-CoV-2 and the subsequent pandemic that the approach gained mainstream attention. Recent approaches have demonstrated effective LTP mediated surface and airborne SARS-CoV-2 decontamination [65]. More now needs to be done to understand the underpinning mode of antiviral action and demonstrate efficacy and repeatability. Compounding this challenge is our ability to test LTP systems under real-world conditions; SARS-CoV-2, like many other viruses of concern, are highly infectious and any scientific study must be conducted in a laboratory with an appropriate containment level. Many researchers do not have access to such facilities and have therefore adopted surrogate models, such as the use of bacteriophages and pseudoviruses [68], [69]. Despite extremely promising results, the applicability of these models remains unclear, and more work is needed to understand how the model results translate to real-world efficacy. Another challenge in thus area relates to how best to apply LTP, with it potentially being used to remove airborne viruses that are in an aerosolized form or applied directly to decontaminate surfaces.

Infectious Proteins: Prions are misfolded proteins with the ability to transmit their misfolded shape onto normal variants of the same protein. Prion diseases are a family of rare progressive neurodegenerative disorders that affect both humans and animals. Typically, prion contaminated materials are extremely difficult to decontaminate, with reports indicating that prions remain infective following incineration at 600 °C [70]. Consequently, prions present a formidable challenge for LTP

decontamination yet numerous studies have reported promising results [71]. Several challenges are yet to be resolved in this promising area, perhaps the most important of which relates to the underpinning mode of action and impact of LTP exposure on the protein structure and conformation. Another key issue that is yet to be fully addressed relates to the fate of the exposed prions, are they completely eradicated or are they simply dislodged from the surface by LTP and transported elsewhere?

2) Proposed Solutions: The LTP community has an evergrowing arsenal of sophisticated diagnostics techniques at their disposal; methods, such as laser Thomson scattering (LTS) and femtosecond two-photon absorption laser-induced fluorescence (fs-TALIF), are becoming more widespread and can provide an unprecedented level of insight into fundamental plasma processes [72], [73]. Despite this progress, the shear level of complexity and multiscale nature of plasma-bio interactions dictates that experimental techniques alone will not be sufficient to uncover a full understanding; computational modeling therefore has a major role to play. While computational plasma models are becoming increasingly advanced and provide a deeper level of insight than ever before [74], the introduction of a realistic biological entity into such models extends beyond our current ability. To overcome this formidable challenge, a fusion of different modeling techniques will be essential, with approaches such as those used in molecular dynamics (MD) being integrated within conventional LTP modeling codes to provide a new insight at the biomolecular level.

In addition to our increasing knowledge of LTP processes, more insight into the biomolecular processes induced by LTP exposure is essential to guide process optimization and instil confidence in potential end users. In terms of virus inactivation, understanding the impact of LTP generated components on viral DNA/RNA alongside proteins and membranes is essential. Techniques such as RNA/DNA sequencing should be adopted to characterize the viral genome, and monitor viral gene expression and genome replication in response to LTP exposure. While the efficiency of LTP mediated virus inactivation should be assessed on appropriate cell lines using different standardized methods, such as the plaque assay and the immunohistochemical focus forming assay [75]. To further enhance our knowledge of LTP mediated biofilm denomination genomic, proteomic and metabolomic studies are vital to obtain a quantitative understanding of the spatial and temporal variation in the microenvironment of cells embedded within the biofilm EPS and how LTP-introduced agents affect cell-cell communication. Antimicrobial resistance remains a key challenge for any novel decontamination technology and the impact of LTP exposure on the emergence of resistance and adaptation of microorganisms remains unclear; addressing this will require dedicated genotypic approaches, such as PCR and DNA hybridization methods. Finally, LTP interaction with infectious proteins requires a detailed investigation into possible changes to the protein structure and conformation, using both classical techniques, such as X-ray crystallography and nuclear magnetic resonance (NMR), and coupled with emerging techniques such as cryo-electron microscopy (cryo-EM) [76].

Armed with a clear understanding of the fundamental processes at play, the question of LTP manipulation and control must be addressed. Given that it is desirable to operate LTP under ambient pressure conditions, the resulting plasma is highly collisional; hence, many of the techniques successfully developed to control low-pressure plasma are ineffective. Advanced power sources employing dual frequency excitation or short voltage pulses (nanosecond and subnanosecond duration) may offer the exciting ability to manipulate the electron energy distribution function high-pressure LTP and therefore, provide a means to manipulate RONS generation. While such techniques are promising, knowing how to control LTP processes is only one part of the puzzle, knowing how to control them to achieve the desired biological response in an ever-changing environment is the ultimate key to successes. Addressing this challenge will undoubtedly require a closed-loop control methodology, informed by real-time sensing and diagnostics. Very few LTP systems currently under investigation employ feedback and are thus highly susceptible to fluctuations in the external environment, ultimately impacting their efficiency and repeatability [48], [77]. To support the development of real-time and closed-loop control systems, many lessons can be drawn from the field of data science, machine learning, and AI. Exciting preliminary results have demonstrated that predictive control methodologies can be applied to harness LTP [51], with such strategies facilitating the control of a complex black-box function, such as an LTP interacting with a biological target, with little prior knowledge of the systems characteristics.

To conclude, as the development of LTP technology for biological decontamination spans the field of plasma science, engineering, and life sciences, consequently, a multidisciplinary approach is therefore essential to make progress. While the LTP community has been extremely successful in demonstrating the potential of the technology for biological decontamination in a plethora of exciting application areas, further engagement with scientists from beyond the LTP domain is essential to overcome the formidable research challenges that remain. As with other LTP applications in the biomedical field, a key challenge is understanding the complex underpinning mechanisms of plasma action, an endeavor that can only succeed through a cohesive and multidisciplinary approach.

## IV. PLASMA TREATED AIR AND PLASMA TREATED WATER K. STAPELMANN, S. MEDEDOVIC THAGARD, AND A. FRIDMAN

#### A. Current State

The growing use of atmospheric pressure plasma opened the door for their applications in the treatment of water and air. During the last decade, plasma treatment of water has been investigated for nitrogen fixation, reduction of pollutants, and inactivation of pathogens in the food cycle, food safety, and water purification arenas. The treatment of air, which traditionally has been focused on the inactivation of airborne pathogens in air flows and on their chemical cleaning, has gained significant renewed interest due to the COVID-19 pandemic.



Fig. 4. DBD for nitrogen fixation in water. The plasma is in direct contact with the water surface. To increase the residence time and the surface area to volume ratio, air is flown through the plasma and bubbled into a 500-ml water column (courtesy of Plasma for Life Sciences Laboratory, North Carolina State University).

Plasma Treated Water for the Food Cycle and Food Safety: LTP science and technology has found its place in agriculture for applications ranging from sustainable fertilizer production to food safety and reduction of pollution and pathogens. Sustainable farming is an international priority due to world population growth, global warming, and decreases in arable land. Sustainable farming requires efficient management of water and nitrogen-based fertilizers to conserve scarce water resources, minimize environmental impact, and reduce fossil fuels now used for fertilizer production, mainly with the Haber-Bosch process. To supplement the global nitrogen demand and offer environmental protection, a more flexible process can be developed with plasma-treated water (PTW) (see Fig. 4 for an example PTW reactor). PTW has been used for the irrigation of seeds and seedlings, showing a growth-promoting effect. This opens up the possibility of precisely manipulating plant productivity while enhancing plant resilience to biotic and abiotic stresses [78]. PTW for fertigation (fertigation is the injection of fertilizers and chemicals into an irrigation system) of plants can be used to supplement nitrogen in soil and soilless systems, as well as to control pH and bacterial load in recirculating nutrient solutions in soilless systems. In case of seed irrigation with PTW, the seeds are exposed to hydrogen peroxide (H2O2) and nitric oxides  $(NO_x)$ , which may trigger changes in plant hormones, promoting germination. LTP offers the unique opportunity to tailor and control reactive species produced.

Addressing another grand challenge, the reduction of postharvest loss, PTW has been applied to fresh produce to reduce microorganisms and potential chemical contaminants on food [79]. It is well known that plasma and PTW have antimicrobial effects [3], [80], [81] and expanding the knowl-edge from plasma sterilization and plasma medicine into the plasma agriculture sector is a logical step. PTW can be generated in bulk liquids and applied as a sanitizer [82], [83]. It has been shown that PTW retains or even promotes food quality while effectively inactivating microorganisms with minimal impact on chemical, nutritional, texture, and sensory attributes

of food produce [79], [84]. Significant water conservation in fresh produce washing and protective treatment has been achieved by replacing bulk wash water with plasma-activated micron- and submicron-water mist. In addition to the significant decrease of wash water consumption, the plasma-mist processing of fresh produce demonstrated high efficacy of bacterial inactivation [85].

The efficient generation and transport of reactive species into the liquid is the key to advancing the widespread use of plasma devices for microbial inactivation, nitrogen fixation, and for treating different source water, as discussed next. Regardless of the application, the process owes its strong chemical conversion/oxidation and disinfection capabilities to the generation of RONS in the gas phase and their subsequent solvation into the liquid phase and also has the advantage of synergistic effects of high electric fields, and UV/VUV light emissions. The best performing plasma reactors feature a high surface area to volume ratio, which ensures increased solvation rates [86]. Plasma in contact with a liquid on large surfaces, plasma in bubbles in liquids, and water droplets in the plasma have all been utilized to increase this ratio [87].

Purification of Water Supplies: The first report on utilizing electrical discharge plasmas to directly degrade organic contaminants dates back to the 1980s when Clements et al. reported decolorizing a dye by using an electrical discharge within a liquid [91]. Since then, over a hundred different plasma reactors have been developed to treat a range of contaminants of environmental importance, including pharmaceuticals, herbicides, pesticides, warfare agents, bacteria, yeasts, and viruses using direct-in-liquid discharges with and without bubbles, and discharges in a gas over and contacting the surface of a liquid giving rise to electrohydraulic reactors, bubble reactors, spray reactors, hybrid gas-liquid reactors, falling water film reactors, etc. Different excitation sources, including ac, nanosecond pulsed, and dc voltages, have been utilized to produce pulsed corona, corona-like, spark, arc, and glow discharges, among other discharge types. Plasmabased water treatment (PWT) generates a range of RONS and other species, as explained in the previous section, requires no chemicals, and can be optimized for batch and continuous processing. The best performing plasma reactors effectively utilize plasma-generated species by concentrating the contaminants at the plasma-liquid interface and, as noted earlier, feature high surface area to volume ratios. However, despite the obvious benefits and advantages of PWT, the technology is only now reaching a level of development where it can be commercially used to treat emerging and legacy contaminants, as demonstrated recently on pharmaceutical waste [92] and groundwater contaminated with poly- and perfluoroalkyl substances (PFAS) (Fig. 5 [93]).

PWT is generally considered a highly energy-intensive process and a significantly more complex technology compared to the existing advanced oxidation processes. However, the scientific evidence to support these claims is scarce and often misleading as the plasma-based treatment of PFAS has been shown to outperform all existing destructive technologies in terms of energy efficiency and treatment time on bench and pilot scales by an order of magnitude [94], [95]. To overcome



Fig. 5. Commercial 10-gpm water treatment system based on electrical discharge plasma (courtesy of Plasma Research Laboratory, Clarkson University).

the widespread skepticism toward plasma as an advanced oxidation technology and position the process at the forefront of water purification, the technology cannot be simply as good as the other oxidation processes but significantly better and able to treat a wide range of common water constituents.

Decontamination of Air Supply: Another use of plasma attracting significant interest is the treatment of air focused both on inactivation of airborne pathogens in air flows, as well as on their chemical cleaning. The recent coronavirus related problems, environmental protection challenges, as well as the growing threat of bioterrorism have brought into focus the need for effective methods to decontaminate and sterilize not only different surfaces but also gases and especially air. In the early 2000s, most of the plasma-based air sterilizers have been successful only when coupling the plasma technology with high efficiency particulate air (HEPA) filters to both trap and kill microorganisms [96]. Usually, such coupling of filtration and plasma systems means trapping the microorganisms with the filter followed by sterilization of the filter using nonthermal plasma discharges.

A few studies, some inspired by the COVID-19 pandemic and recently reviewed by Mohamed *et al.* [97], have shown the efficacy of plasmas in inactivating airborne pathogens without the use of a filter, especially to prevent virus transmission. Most commonly, DBDs are used in different configurations (see Fig. 6 for an example). DBDs offer the advantages of operating in ambient air and at room temperature. As a mode of action, the short-lived reactive species OH and singlet oxygen have been suggested. Hence, the most promising results reported in the literature were obtained when aerosols were passed through active plasma volume (rather than effluent). The challenge of air treatment in ventilation systems is the short residence time on the order of subseconds of the air inside the active plasma region.

Plasma air treatment, similar to plasma water treatment, is generally considered a highly energy intensive process and a significantly more complex technology compared to the existing air cleaning technologies. However, the current gold



Fig. 6. Pathogen detection and remediation facility (PDRF): above—general view and below—dielectric barrier grating discharge (DBGD) with air sterilization chamber [99].

standard for the reduction of airborne pathogens is filtering them out with HEPA filters. These filters effectively retain particles, such as bacteria, viruses, and pollen without inactivating them. The filters cause significant pressure losses in HVAC systems giving rise to higher energy and maintenance costs. The HVAC system itself has to be designed for the pressure losses, which does not allow for a later upgrading of the HVAC system with such filters without substantial costs. Especially during the COVID-19 pandemic, air cleaning systems that can be installed in existing HVAC systems would have been of great advantage. As the *Decadal Assessment of Plasma Science* conducted by the National Academies of Science, Engineering, and Medicine stated in May 2020: "… we may have missed an opportunity to have another tool at our disposal to aid in the current health crisis" [98].

## B. Challenges and Proposed Solutions

1) Present and Future Challenges (Plasma Treated Water for the Food Cycle and Food Safety): To date, most of the literature available using PTW for fertigation focuses on plant growth in early growth stages, typically in soilless systems, as recently reviewed by Ranieri *et al.* [78]. The effect of PTW on plants needs to be studied beyond the seedling stage for the process to become a viable alternative for agriculture. Furthermore, knowledge about the effects of PTW on plant growth in actual soil systems and the impact of PTW on soil health and the microbiome structure is limited. The effect of other RONS generated in PTW beyond  $NO_3^-$  is also not well understood. Further research is needed on  $H_2O_2$  and NO, which may act as signaling molecules and were proposed as strategies to enhance crop stress adaptation and to prolong shelf life [88]. To understand the impact of PTW on the whole plant and its ecosystem, multidisciplinary research approaches, including plant biologists, soil scientists, microbiologists, and plasma scientists and engineers, are necessary.

For the translation of laboratory results to field conditions, the impact of organic matter in water on the plasma chemistry also needs to be considered and studied. Tap water or well water is typically used for irrigation in greenhouses and fields. It is well known that the buffering capacity of tap water influences the liquid chemistry [87], [89]. While the use of deionized or ultrapure water in the lab is justified for the reproducibility of results and to gain fundamental understanding in simpler systems, the presence of organic matter that may become part of chemical reaction pathways [90] needs to be studied in more detail to understand, optimize, and eventually commercialize PTW for nitrogen fixation.

The use of PTW or plasma-activated mist in food safety applications is a promising field. However, limited information is available about the impact on produce on a molecular scale beyond pH, color, and texture. Some studies report discoloration or decrease in firmness after exposure to PTW, some reported no reduction in terms of vitamins, while others reported increased vitamin D content, for example. The scientific community is using a variety of different plasma sources, with different excitation mechanisms, gas compositions, different liquid volumes, and inconsistent times between PTW preparation and use. Due to the transient nature of many reactive species believed to be crucial for inactivation of bacteria, the time between PTW preparation and application is of significant importance, yet not always reported in the literature. Given the varieties of produce and plasma systems used, it is difficult to understand the fundamental science of PTW interacting with fresh produce.

*Purification of Water Supplies:* Despite the convoluted effects of a multitude of parameters affecting the performance of plasma reactors, there is increasing evidence that the current plasma reactors are superior for the treatment of surfactant compounds (e.g., PFAS, foams, and dyes) but are unable to competitively treat nonsurfactants that in reality constitute the majority of the compounds of environmental importance. Therefore, increasing the conversion efficiency for the treatment of nonsurfactants to the levels of surfactant-like compounds, and increasing the energy efficiency and throughput for the treatment of all compounds represent and remain the key challenges for PWT.

The slow progress in the development and scaleup of plasma reactors in general is rooted in three fundamental challenges. The first one relates to the lack of understanding of the basic processes in the plasma, in the bulk liquid and at the plasmaliquid interface, including the nature of their interaction. The second challenge surrounds the plasma reactor design and the lack of understanding of the extent to which the classical chemical reactor design theory is applicable to the development of (heterogeneous) plasma reactor systems. There also exists a large gap between the fundamental discoveries and translation of those discoveries into engineering solutions. The plasma reactor scaleup and the applicable scaling laws belong to the third category of challenges.

The plasma reactor development for water purification purposes has been and still is largely trial and error, with research predominantly conducted in simple systems containing a single compound and using a range of different plasma excitation sources. Plasma diagnostics to quantify the plasma physics and chemistry remains a challenge. The bulk liquid composition and transport have been recognized as significant factors affecting the reactor performance and more research are needed using techniques, such as LIF and particle image velocimetry (PIV) coupled with kinetic experiments to determine the relative importance of diffusive and convective mass transport and elucidate their individual roles in compound degradation [100], [101]. For a range of organic compounds, we are still unable to differentiate among different plasma agents responsible for their degradation, which hinders the plasma reactor design and optimization. The roles of individual ROS, RNS, V(UV), ions, metastables, and solvated electrons including that of the electric field are largely unknown due to the numerous issues associated with the existing techniques, protocols, assays, and commercial kits that have been developed. It is largely due to the absence of the adequate probe(s) and/or techniques to measure the concentrations of dissolved short-lived species that the community is still unable to learn how to control the plasma and bulk liquid chemistry with the input operational parameters and ultimately maximize the production of oxidative species or reductive species (plasma is a both an oxidative and reductive process) for a particular water treatment application.

To understand the fundamental science behind interfacial dynamics and develop accurate theories that describe process performance, more data are usually better. However, to upscale a plasma reactor by any degree requires information on only the key processes and design parameters that could be empirical. To date, ensuring a high plasma area to liquid volume ratio has been identified as the critical requirement but also the key technological challenge in the plasma reactor scaleup [95].

*Decontamination of Air Supply:* Three major challenges are related to successful commercial and industrial application of plasma sterilization and cleaning of air flows: 1) energy cost; 2) productivity; and 3) selectivity.

 Energy Cost of Plasma Air Treatment: Plasma air decontamination systems consume electric energy, which determines most of the operational cost of the relevant technologies. The electric energy cost is a crucial challenge of plasma air cleaning since the majority of conventional and novel competing air cleaning technologies are less electric energy intensive (or do not consume electric energy at all). Cost is especially important for decontamination of large air volumes. The analysis of cost effectiveness shows that to be economically viable, the electric energy consumption must be below  $1-3 \times 10^{-3}$  kWh/m<sup>3</sup> of air for decontamination of large air volumes.

- 2) Thruput (Capacity) of the Plasma System: To be effectively commercialized, the air flow capacity of plasma systems should be scaled up to typical industrial/commercial levels of thousands of  $m^3$  per hour. Even at the expected energy effective regimes (electric energy cost below  $1 3 \times 10^{-3}$  kWh/m<sup>3</sup>), the airflows on the level of thousands of  $m^3$  per hour require at least 10 kW of cold plasma power. This is a significant engineering challenge that necessitates the development of powerful cold plasma discharges and building relevant power supplies with high efficiency matching characteristics.
- 3) Selectivity of Air Cleaning Processes: Plasma systems may generate nonacceptable treatment byproducts, such as ozone, nitrogen oxides, etc. These byproducts restrict types and operational regimes of applied nonthermal plasma discharges. Detailed understanding of relevant plasmochemical process mechanisms and analysis of plasma system combination with other air treatment approaches are still needed.

## C. Proposed Solutions

Plasma Treated Water for the Food Cycle and Food Safety: As a common theme for plasma agriculture applications, upscaling is one of the biggest challenges. In order to optimize the process and to design highly efficient plasma-water or plasma-mist reactors, more fundamental understanding is required of: 1) the kinetics and collisional processes in the highly nonequilibrium plasma regime; 2) the (efficient) transport of species into the liquid; and 3) the plasma-generated species required for the optimum outcome. For a more fundamental understanding, advances in diagnostic capabilities in the liquid phase and at the plasma-liquid interface are required, as well as advances in modeling, capturing the properties of nonequilibrium plasma kinetics and the transport through the plasma-liquid interface into the liquid. The effect of charged species and photon-induced effects, as well as reactions involving short-lived species like atomic oxygen have yet to be addressed as many reaction rates, cross sections, and rate constants are unknown. Calculations and measurements of these unknown quantities are critical for advancing the understanding of the plasma-liquid interfacial processes.

Regarding the applications of PTW in the food cycle and food safety arenas, the plasma parameters affecting the enhanced plant growth and resilience, healthy soil, and nutrient management must be better understood to optimize the process. While the effect of plasma-generated nitrate or ammonium as fertilizer is well known, other RONS and their potential benefits for plant growth and the plants' ecosystem have not been studied in detail yet. For food safety applications, the impact of RONS in the liquid on food quality and nutrients is also not well understood. The use of appropriate controls, e.g.. nitrate, nitrite, and hydrogen peroxide controls with a concentration matching the concentrations in PTW, would help to identify mechanisms and to pave the road for upscaling and commercialization efforts.

Purification of Water Supplies: One of the biggest challenges in the design, optimization, and scaleup of highly efficient plasma reactors is finding the way for the community to work together toward this common goal and build on each other's findings. A large volume of literature currently exists on the effectiveness of over a hundred different plasma reactors for treating dozens of different compounds and biological species. While each of these studies is unarguably useful and informative, systematic comparison of the compound(s)/microorganisms studied is needed, including their degradation/inactivation rates and starting concentrations, reactor types, discharge energies, excitation sources, and treatment volumes to be able to deconvolute and extract any information regarding the key design and operational principles that control the system performance. While such studies are rare due to their laborious nature, they are extremely informative and may set empirical guidelines for the plasma reactor design [95].

To remedy this issue and essentially calibrate multiple concurrent reactor design and scaleup research efforts, the community should agree on using and characterizing a reference plasma treatment system similar to the COST Reference Microplasma Jet that is currently being used in the biomedical community to identify the parameters and processes that guide and control the plasma reactor design and the process performance. An example of such a system would be a point gas discharge in a noble gas contacting the surface of the treated liquid with a grounded plate immersed in the liquid. Common electrode material, electrode dimensions, reactor volume, and the compound/microorganism treated would be chosen. Beyond these set parameters, immediate new research would welcome investigations on the excitation source types, process gas types, mixing, etc., and include advanced spectroscopic/laser techniques to quantify the plasma physics and chemistry. In the liquid, complementary analytical measurements would be focused on the quantification and/or identification of the extent of the compound degradation, byproducts and their toxicity, reaction mechanisms, and the species responsible for the chemical transformations. Concurrently, techniques, such as LIF and PIV, would be applied to determine the location of degradation/inactivation, visualize the flow, and measure the velocity fields in the liquid. The outcomes of these fundamental investigations would assist in determining the limitations and the advantages of the technology, its level of performance with respect to the existing technologies and, if applicable, the key technical challenges that must be overcome before the technology can be upscaled. Once the experimental, design, and operational conditions under which the system performs equally well or better compared to the existing technologies are determined, then the actual scaleup can commence.

## D. Decontamination of Air Supply

Meeting the challenges of energy efficiency, air flow capacity, and process selectivity requires the following.

1) *Effective combination of plasma technologies with conventional approaches*, including scrubbing, filtration, etc., allows for a significant decrease of the total process energy cost. In particular, the combination of plasma treatment with filtration and scrubbing limits plasma treatment to only charging and partial oxidation of biological and chemical pollutants, thus decreasing electric energy consumption in orders of magnitude.

- 2) The development of *effective and not-expensive cold plasma power supplies* operating on the power level up to hundreds of kW with an appropriate source that has plasma load matching characteristics.
- Plasma-chemistry optimization to suppress generated nonacceptable treatment byproducts, such as ozone, nitrogen oxides, etc., requires, in particular, analysis and characterization of nonuniformity of plasma-chemical processes in air.

To conclude, there is substantial interest in water treatment using electrical discharge plasmas, especially for the degradation of legacy and emerging contaminants or point-of-use fertilizer production. Similarly, the recent COVID-19 pandemic inspired renewed interest in alternative air treatment methods. The Decadal Assessment of Plasma Science by the National Academies of Science, Engineering, and Medicine released in May 2020 states: "The central challenge in many of the proposed applications of LTP activated processes to energy, water, food, and agriculture is in scaling. Even if plasma can be shown to be effective on small laboratory scales, the process must be scaled sufficiently to make it useful in an industrial setting, whether in the factory or the corporate farm" [98]. The feasibility of current plasma devices for the treatment of water and air has been demonstrated on the bench scale; however, their widespread commercial and industrial applicability necessitate further optimization of energy efficiency and the actual scaleup of the devices and their plasma-generating networks to the required levels. For the wide adoption of the plasma-based devices, the process cannot be just as good as the existing technologies but significantly better.

While there have been significant advances in understanding the chemistry and physics of the single and multiphase (gas–liquid) plasma systems, these have been generally studied separately, either from the plasma or from the bulk liquid side. To understand the processes that control chemical conversion and energy efficiency is crucial for the scaleup of any plasma system and requires a holistic multidisciplinary approach that identifies the key measurements, sets of data, and/or the reactor design element(s). In other words, to translate fundamental findings into engineering solutions, the community has to rethink its approach toward the common goal of plasma air and plasma water treatment commercialization.

## V. LTP FOR WOUND HEALING S. BEKECHUS

#### A. Current State

Nonhealing and chronic wounds are a major burden to both patients and healthcare systems. About two decades ago, the application of LTP for wound healing was among the first medical problems envisioned to be addressed using gas plasma technologies after finding promising antimicrobial effects of LTP in the 1990s [1], [102], [103]. However, taking the

concept into the clinical routine took place only in the last decade. The requirements for bench-to-bedside concepts are multifold. The plasma device must be safe, generating a reproducible plasma, manufactured based on quality management procedures, receiving appropriate accreditation by authorities, fulfill reasonable criteria for basing business models on existing needs in the healthcare market, and, ultimately, medical doctors willing to explore such novel tool in the clinical setting.

The current state of LTP in wound healing can be described from several perspectives, such as: 1) regional practice; 2) preclinical versus clinical; and 3) the plasma devices' specifications. From the regional perspective, LTP-mediated wound healing in the clinical setting is most advanced in central Europe, especially Germany. All plasma devices that were accredited by official medical authorities to treat chronic wounds within dermatological centers were engineered and put into practice in Germany. According to the directive on Medical Device Regulation (MDR, EU 2017/745), these devices can be used across the European Union. Official numbers of devices in the field are not available, but based on personal communications, it is estimated that there are a few hundred devices currently sold to or rented by dermatological centers and practitioners. There are efforts in, e.g., the U.S. and the Republic of Korea to achieve similar local authorization for plasma devices aimed at targeting chronic wounds in humans. It is emphasized that this text only focuses on clinical and human cases of wound healing, not on cosmetic or veterinary applications.

For clinical wound healing, several hypotheses exist to explain the benefits from LTP exposure, suggesting several mechanisms simultaneously at play to promote wound healing. Usually being less than 1 log (90%), the antimicrobial efficacy of LTP-treated wounds is documented but overall modest [105], [106], also when analyzed ex vivo [107]. Accordingly, a recent prospective, randomized, placebocontrolled, patient-blinded clinical trial comparing plasma versus antiseptics in a head-to-head study indicated LTPpromoted wound healing independent to antimicrobial efficacy as patients in all groups received antiseptics. The effect of LTP-derived electric fields in wound healing is unclear, especially since there is a lack of studies on intensity-matched exposure of electric fields alone in appropriate in vivo wound models. The mild heat transferred to the wound from the LTP is rather beneficial than detrimental to wound healing [108]. The dominant effector hypothesized to drive LTP-induced wound healing is RONS (ROS/RNS) [13], [109]. Albeit direct clinical evidence of the roles of ROS/RNS is challenging to obtain, overwhelming data are supporting this idea, as described by Graves [23] in his 2012 review. In addition to this, the recent work in LTP-treated patient wounds suggests plasma to spur on microcirculation and tissue oxygenation in superficial and deeper layers [110], [111].

The preclinical evidence of plasma-treated wounds in mice from Schmidt *et al.* [112] confirmed these findings and suggested an important role of focal adhesions and complexes and matrix remodeling in plasma-treated wounds [104] (see Fig. 7). Most importantly, an array of in vivo data in



Fig. 7. Hyperspectral imaging of wounded ear (left) with orange wound region in control mice (top) and enhanced signal generation in the plasmatreated wound (bottom). Boxplots show quantitative data for several parameters of hyperspectral imaging. Figure is adapted from [104].

noninfected wounds of different origins (e.g., punch biopsies, burn, and laser induced), animal species (e.g., mice, rats, and sheep), and locations (e.g., back and ear) has shown plasma treatment to promote healing in the absence of infection [113]–[120]. This markedly substantiates the idea of redox control of wound healing via plasma-derived ROS/RNS. The discussion on mechanisms is beyond this article's scope, but it has been recently suggested that the redox-regulating transcription factor Nrf2 and altered immune cell infiltration take part in plasma-accelerated healing using the kINPen [121]. Also, the mechanism of action of the accredited microplaster plasma device was delineated to redox action and stimulation of wound-resident cells of the body [122]–[125]. The details on *in vitro* findings were summarized recently [126].

Regarding the perspective on the types of cold plasma sources, only devices in clinical practice are discussed because an overwhelming number of plasma devices have only been covered in relatively few papers and rarely in a clinical setting, while usually lacking accreditation. Of the three devices currently marketed based on thorough clinical experience and medical product guideline compliance or FDA approval specifically for chronic wound healing in dermatology, two are DBDs (PlasmaDerm from Cinogy and SteriPlas from Adtec), and one is a jet (kINPen MED from neoplas med). All have been investigated in clinical trials, as summarized before [127]-[129]. Two more devices from Germany, PlasmaCare from TerraPlasma (clinical trial completed) and a silicon-based plasma-plaster called CPTpatch from the INP spin-off company ColdPlasmaTech (clinical trial in preparation), are within authorization phases. The DBD device PlasmaCare is innovative because of its battery-driven nature, making it an excellent option for ambulant and home wound care. The CPTpatch is a single-use wound dressing that is planned to be left on the wound to ignite a plasma directly in the wound bed multiple times. This status suggests a dominance of DBD devices over jets in present and future medical plasma wound care. Notwithstanding, clinicians repeatedly emphasize the scalpel-like precision treatment option that comes with plasma jet systems. Especially, experienced wound nurses and healthcare practitioners know that some areas in chronic wounds are more problematic than others [130], which can be effectively targeted with plasma jets in a locoregionalized manner and, e.g., with individual treatment times. Regardless of the device type, the plasma therapy complements and does not replace existing wound care measures. Critical

steps, such as wound debridement and dressing change, remain to be vital for therapeutic success.

## B. Challenges and Proposed Solutions

The challenges and opportunities in the transdisciplinary field of plasma wound that are covering knowledge from physics, biochemistry, and medicine are vast. A detailed discussion, especially individual cell type responses to plasma treatment, is therefore out of scope of this work. The most critical aspects will be discussed nevertheless.

The perhaps greatest asset and caveat at the same time in plasma medicine is the large number of devices reported on. While this is beneficial for exploring the science behind the physics and biology in plasma medicine, it is detrimental to clinical practice. An ideal clinical plasma device would have been accredited across all continents to spur global research efforts, clinical trials, and, ultimately, a certain degree of comparability between results. These would then be independent of the device and depend on treatment parameters, such as treatment duration per cycle, frequency of treatments per week, onset indications of treatment, and offset indications for stopping plasma therapy. Currently, the published evidence on such aspects is absent, hampering the motivation of clinicians less experienced with plasma therapy to implement this technology into their treatment portfolio. Hence, the future plasma therapy needs clear guidelines that state the boundary conditions of application, which less plasma-experienced clinicians can also follow. Such guideline is currently set up in Germany within the framework of the so-called AWMF catalog of treatments. Such medical guidelines would allow reimbursement of plasma therapy, a critical step in the German healthcare system that draws the line between reimbursed standard therapies covered by the obligatory healthcare insurance and nonreimbursed experimental therapies covered by the patient or medical doctor. After all, the additional treatment costs have to significantly reduce the long-term costs of existing wound care to be regarded as superior to existing therapies. In general, low-cost accredited plasma devices commercially available globally will be the key for taking plasma wound therapy outside its current niche, with great potential benefits, especially in developing countries.

From a clinician scientist's point of view, several points are unclear in the plasma wound therapy, for example: What is the best treatment schedule in terms of duration and frequency? Should the entire wound be treated or, e.g., predominantly infected parts? Are some plasma devices superior in some types of wounds over others? What is the role of infection? Are there common traits in patients that do not respond to plasma wound therapy? Is optimization possible of existing medical product DBD and plasma jet device, and if so, how specifically? What role plays the wound etiology underlying the disease, e.g., diabetes? Should the treatment regimen be adjusted for wound location and size? Is plasma therapy affecting existing wound care measures positively or negatively, and can it be combined with other therapies such as hyperbaric oxygen or vacuum? This shortlist of questions exemplifies the need for applying and reporting clinical plasma wound therapy

experiences to a much greater extent than currently being the case. Ideally, only a few plasma devices should be used globally in such settings to build knowledge consecutively instead of serially, as is currently the case. Other, although admittedly less complex technologies based on physical principles that had entered medical practice, such as radiotherapy and electrochemotherapy, went similar routes of standardization.

From a preclinical perspective, understanding the interface reactions of plasma with the target tissue is perhaps the least explored but the most needed challenge in plasma wound therapy and medicine in general [126]. First, it needs to be understood that there is no genuine bulk liquid in wounds. In addition to wound cells and microorganisms, amino acids, proteins, lipids, and nucleic acids will always be close by to the plasma streamers, offering plenty of reaction partners for plasma-derived ROS/RNS. It is therefore plausible that the role of short-lived ROS/RNS is much greater than in liquid-rich laboratory assay systems with cells, in which diffusion distances between the ROS/RNS source (plasma) and the target (cells) are too high for the short-lived ROS/RNS to maintain reactivity. The argument that the tissue is composed of water is also void. A majority of water is in the cytosol of cells, a compartment packed with redox-active proteins, adding to the already complex plasma chemistry. Three conclusions follow from this argumentation. First, it is vital to increase the understanding of which types of ROS/RNS reach the target to what extent. Unfortunately, the number of tools available in redox research to distinguish the different types of ROS/RNS on a quantitative scale is limited. For the fluorescent redox indicators proposed to be specific, virtually all of them have cross-reactivity with multiple types of ROS/RNS, especially when introduced into cells [131], [132]. Second, the research on noncellular targets as mentioned above needs to be identified to clarify their role and contribution to plasma-mediated cellular effects. The fact that oxidative modification of such targets can be of biological and immunological relevance has recently been demonstrated for the kINPen [133], [134], apart from a broad array of work in the field of redox biology for single ROS/RNS [135]-[137]. Third, and based on the questions proposed from a clinical perspective, it is critical to understand that only animal models recapitulate the wound environment to a sufficient extent to allow true preclinical conclusions on plasma parameters and biological effectors. The contribution of the immune system, which cannot be accurately mimicked in vitro, is highly critical for the onset and offset of the different wound healing phases to allow complete healing [138]–[140]. Other vital aspects, such as angiogenesis and matrix remodeling, also take place in appropriate test systems only. By directly comparing infected versus noninfected wounds, final conclusions may also be drawn toward the role of antimicrobial plasma effects in supporting healing responses. The role of cells of the immune system in plasma-enhanced wound healing might be analyzed using immune-deficient mice or antibody-mediated depletion.

From the plasma device perspective, the field of plasma jets currently offers the greatest opportunity of improvement. Jets have the caveat of being limited to a small area, while patient wounds can be significantly large, leading to long exposure times and ultimately also costs. Several groups have mitigated this issue using multijet arrays [141], [142] or are currently working on this. Plasma jets (and gas-driven DBDs) also offer the opportunity of tuning the feed gas toward applicationoriented effects, a principle shown multiple times by several groups across in physical, biochemical, antimicrobial, biological, and *in vivo* characterization [143]–[148], including wound healing in rodent models [113], [149], [150]. Ultimately, however, it might be more reasonable to optimize existing plasma devices that have received or are planned to receive accredition as medical products to facilitate the translation of data and knowledge from physics and biology plasma medicine into clinical plasma medicine.

## VI. LTP IN DENTISTRY C. JIANG

## A. Current State

The earliest application of gas discharges in dental practice dates back to 50 years ago; in 1970s, plasma-sprayed titanium (Ti) or Ti plasma-coating for metals and ceramics was used for oral implants [151]. In the late 80s and early 90s, more applications of low-pressure plasmas in dentistry emerged, including dental instrument sterilization [152]-[154] and surface treatment of biomaterials, such as denture, splint ribbon, composite post, and dentin to assist miscellaneous dental procedures [151], [155], [156]. It was not until the early 2000s, however, the direct treatment of teeth and oral surfaces for potential chairside uses became possible due to the development of AP-LTP based technologies [2]. Applications of plasmas in dentistry have since flourished and entwined with other new technologies in different dental specialties for advancement of the relevant techniques. Reviews or updates on similar topics frequent in dental, medical, and engineering journals, especially in the past decade [155], [157]-[161]. The status/update reported here hence serves as an addition but not a replacement to the previous reviews on the applications of LTP in dentistry.

According to the potential clinical use, applications of the plasma-based technology can be categorized as *direct* and *indirect* uses of plasma. Direct uses of plasma refer to applying direct plasma treatment on dentin, gum, implant, or other dental objects of a patient in a dental office for chairside uses. Any other uses of plasma in dental practice, such as the use of plasma for cleaning, preparation, or modification of dental materials and surfaces, which can be achieved in a laboratory and outside of a dental office, belong to the indirect uses of plasma. Due to the different application settings of direct and indirect uses of plasma, the requirements and challenges for technology adaption are hence significantly different, which also impact the state of the technology development for specific applications.

The direct uses of plasma can be classified as three areas in dentistry: 1) periodontal; 2) endodontic; and 3) prosthodontic treatments. While the antimicrobial effects of plasma play the most important roles in the first two areas of applications, surface modification to improve the adhesion properties or change

the coloration of dentin is the main cause for the prosthodontic treatment. Table I lists the specific applications that can be classified in different dental practice areas, the related study models, plasma sources used, and their corresponding status. Periodontal treatments include both removal pathogenic microorganisms or biofilms from cavities (such as caries) or implants that were previously applied in a patient and causing peri-implantitis. For research effort, cavity cleaning may overlap with root canal disinfection, although the choice of microbial strains and treatment substrates may vary. For root canal disinfection, accessibility of the plasma-induced disinfecting effect into the apex and the rest tubular structures of a root canal is an additional challenge for the application, and hence, the outcome from ex vivo and in vivo studies would weigh more to help evaluate the potential success of the technology. The status of both caries removal and endodontic disinfection is hence similar: it is evident that LTP treatment can be as effective as the use of bleach such as 5%–6% NaOCl or 2% CHX, while the combination of LTP with bleach irrigation can achieve even better disinfection outcome [158], [168]–[171]. Studies for implant surface cleaning using LTP are fewer comparing to other areas of applications. Although the efficacy of atmospheric plasmas, powered by pulsed microwave (MW) or radio frequency (RF), on implant surface decontamination has been demonstrated, safety studies of these plasma applications are lacking. Additionally, a recent study showed that the effect of plasma is not significant for implant surface disinfection when the plasma treatment was added to an air-polishing protocol [165]. Prosthodontic treatments are dental procedures for esthetics or cosmetic restorations. They include tooth bleaching and dentin treatment for bonding to resins. The tooth bleaching application of plasma uses the surface modification properties of plasma and enhances the bleaching activity of bleach agents on dentin surfaces. Studies have shown the combination of LTP and bleach agents enhanced the bleaching effectiveness by a factor of 3 or higher [175], [176]. For the dentin treatment, a recent systematic review on the effect of LTP on adhesives resin-dentin micro-tensile bond strength ( $\mu$ TBS) showed that LTP application could enhance resin-dentin  $\mu$ TBS of etchand-rinse (ER) adhesives or universal adhesives applied in the ER mode [158]. Some studies showed that an optimal dosage (e.g., treatment time) was important to achieve better interfacial bonding strength or  $\mu$ TBS, while overtreatment resulted decreased interfacial bonding strength [179].

The indirect uses of plasma in dentistry include surface modifications of implants for implantology or oral surgery, improving the interfacial bonding strength or adhesion of dental objects used in periodontics or prosthodontics, and sterilization of dental instruments. Surface modification using plasmas, both low-pressure and atmospheric pressure, is the most widely used plasma-induced process that impacts various applications, including surface modification/activation of Ti, Zr, or CaP-coated implants, dental alloys, improving interfacial bonding strength or adhesion of resins, resin composites, glass fibers, ceramics, polyetherketoneketone (PEKK), and dentures. For decontamination of dental instruments, inactivation of dental pathogens with ion bombardment and oxidation induced by low-pressure nonequilibrium plasmas is used to dominate the plasma applications for sterilizing endodontic files or other operative instruments [155], [159]. Although most of these indirect applications started with low-pressure plasmas and may still be able to use low-pressure plasmas, the use of atmospheric pressure LTP eliminates the need of a vacuum system, potentially achieves faster cleaning process, and can hence reduce the cost of operation substantially. As a result, LTP has been attracting more and more interests in research and development and may gradually replace the use of low-pressure plasmas in almost every dental field.

The *in vitro* and *in vivo* studies demonstrated that LTP could enhance surface colonization and osteoblast activity, and accelerate mineralization of implant (e.g., Ti and Zr) surfaces, as well as improve osseointegration of implant [155], [159], [180]. The antimicrobial effects of LTP facilitate biofilm removal and prepare a clean surface of an implant [155], [159]. Additionally, the combination of mechanical and LTP treatments may result in surface "rejuvenation" against peri-implantitis by activating the synergistic antimicrobial effects and surface improvement [155], [159].

## B. Challenges and Proposed Solutions

Decades of research activities have led to substantial development of plasma applications in almost all areas of dentistry. For indirect treatments, the challenges of plasma applications are mainly the efficacy of the plasma-based technologies and the effective cost compared with other nonplasma techniques. The increasing uses of LTP are making the plasma-based technology more economically competitive, especially comparing to lasers or UV-light sources required techniques. The highly controlled surface modification by LTP treatment is becoming the future trend in oral surgery and implantology [159], where chemical functionalization, deposition of antibacterial thin films and coatings, and surface structuring to create antifouling surfaces impact significantly on application outcomes. More importantly, adapting LTP into additive manufacturing, such as three-dimensional (3-D) printing and polymerization to assist multiple areas in dentistry, will decisively pin the plasma-based technology in the map of dental technologies. The reports of plasma sterilization of 3-D printed dental objects [181] or removal of chemical gradients inside porous structures to enhance cell viability in 3-D porous scaffolds (Wan et al. [182] Biomaterials) for tissue engineering are promising start for more plasma-processing-assisted dentistry.

The road to chairside uses of direct plasma treatments, on the other hand, remains more challenging. The large variations in treatment outcome, particularly concerning the antimicrobial effects of LTP [157], and the diverse selections of different LTP, which can be powered by pulsed DC, RF or MW, present dilemmas for dentistry researchers and students to decide for the next-step clinical trial. Nevertheless, studies designed in a clinically relevant model, such as using multispecies microbial biofilms from patients grown on a dentinal surface or in a root canal, as well as animal models have been abundant [162], [167], [168], can be the ultimate testing

## TABLE I DIRECT USES OF AP-LTPS IN DENTISTRY

Specific Applications	Related study models	Plasma sources (power type, working gas)	Status	Ref.
Periodontal Treatments				
Removal of dental caries	dental pathogens and pathogenic biofilms grown on well-plates, agar, HA discs, dentins and other substrates, <i>in vitro</i>	LTP plasma jets, pulsed or RF, Ar/He + (O <sub>2</sub> ) or Air	Large variations in CFU reduction factors depending on the substrates and plasma treatment dosages. In general, plasma effect is comparable with the use of 5-6% NaOCl or 2% CHX irrigation on dentin surfaces. Biofilms are harder to remove.	[157, 162- 164]
Implant disinfection	single-strain microorganisms (e.g., <i>S. mitis</i> , <i>C. albicans</i> , <i>P. gingivalis</i> ) or peri- implantitis biofilms on Ti discs, Ti implants, steel wafers, <i>in vitro</i> or <i>ex vivo</i>	LTP plasma jets, MW or RF, Ar/He + (O <sub>2</sub> , H <sub>2</sub> O) or Air	NTAP was shown effective in removing <i>P. gingivalis</i> biofilm from titanium discs, <i>in vitro</i> . Comparing with other techniques such as air-polishing, the effect of NTAP is not significant and needs more study.	[165, 166]
Endodontic Treatments				
Root canal disinfection	<i>E. faecalis</i> , <i>C. albicans</i> or multi- strain endodontic biofilms grown in root canals or tubular structures mimicking root canals, <i>in vitro</i> , <i>ex vivo</i> , and <i>in vivo</i>	LTP plasma jets, pulsed or RF, Ar/He + (O <sub>2</sub> or bleach solutions)	In most studies, plasma effect is comparable with the use of ~3% NaOCl or 2% CHX irrigation. Biofilms are harder to remove.	[157, 167- 170]
Prosthodontic Treatments				
Tooth bleaching	Stained or natural extracted human teeth (sectioned or whole), in vitro/ex vivo, rabbit teeth in vivo, were subject to bleach (H <sub>2</sub> O <sub>2</sub> , carbamide peroxide) in solution or gel, combined treatment of plasma and bleach	LTP plasma jets, low frequency (e.g. 15- 20 kHz) AC or pulsed, He or Ar	Combining LTP and bleach improved the bleaching efficacy (by a factor of 3 or more)	[171-176]
Dentin treatment for ethetics	HA disc or dentin surfaces prepared from extracted human or bovine teeth, after crown removal, followed by etching and rinsing	LTP glow discharge or DBD, MW, RF, or low frequency AC, He or Ar	Plasma treatment of the dentin surface (for an optimal dosage such as the treatment time) increases the interfacial bonding strength or $\mu$ TBS, and facilitate the adhesion process	[158, 177- 179]

model for *ex vivo* or *in vivo* studies. Comparisons between LTP and other nonplasma treatment protocols are important. Comparable antimicrobial effects of LTP against dental

biofilms in *ex vivo* root canals or animal models are great milestones in plasma applications for periodontal or endodontic treatments. Close collaborations between the dentists and plasma engineers are essential to identify the effective treatment protocols and efficiently use resources for technology advancement. The other challenge is the safety concern. The RONS produced by LTP has been considered playing important roles in antimicrobial processes and surface modification, and also known to cause oxidative and nitrosative stress to cells and tissue. Nam et al. recently conducted a histological safety study in a rabbit model using an LTP for tooth bleaching [183]. The LTP was based on a DBD configuration, powered by 15 kHz ac at 10-kV peak voltage. The working gas was Ar. The gap temperature was about 35 °C. The study concluded that the combinational treatment of plasma and 15% carbamide peroxide (CP) resulted in the most color change compared with 15% CP treatment alone, and the combination treatment did not indicate inflammatory responses nor thermal damages [183]. Nam et al. also noted that the plasma did not cause histological damage in oral soft tissues during tooth bleaching [183]. Another recent in vivo study conducted by Yao et al. [167] demonstrated the antimicrobial effectiveness of an NTAP against apical periodontitis in a beagle model, comparable to that of 2% CHX irrigation, with no detected structural damage in dentin or tissue necrosis at the periapical region. LTP was generated from a handheld plasma jet device, using He as the primary working gas, and powered by pulsed dc at 10 kV and 10 kHz [167]. Yao et al. [167] commented that modifying the LTP jet by following He through 2% CHX enhanced the antimicrobial efficacy. However, the safety study on the modified method was not reported. After all, the generation of RONS was associated with many technologies in dentistry, including lasers, photosensitizers, bleaching agents, and even resin cements [184]. Hence, LTP is not the only technology that may cause safety concerns. The critical point here is the dosage control. More safety studies similar to the above two examples would help pave the way carrying the LTPbased technology through clinical trial and toward the dental chair.

## VII. LTP FOR CANCER TREATMENT M. KEIDAR, V. MILLER, S. BEKESCHUS, D. YAN, H. TANAKA, M. LAROUSSI, K. OSTRIKOV, AND M. HORI

## A. Current State

In recent years LTP, LTP, is increasingly being investigated as a possible new modality for cancer treatment [12], [185]. The novelty of LTP lies in its multifactorial effects that include RONS (ROS/RNS) produced in the plasma, and physical factors, such as emitted electromagnetic waves and the electric fields that are formed when plasma impinges on the tissue [13]. Rapid advances in LTP technology have allowed great control on the final composition and properties of plasma for specific biological processes aimed at clinical applications. Specifically in oncology, LTP has enabled scientists to explore the mechanisms of eliminating cancer cells that would be applicable in diverse types of cancers. Focal solid tumors require different treatment approaches than more diffuse or metastatic tumors. Broadly, the experimental approaches encompass two methods of inducing cancer cell death: 1) direct killing of cancer cells by LTP in situ and 2) stimulation of immune responses by inducing controlled oxidative stress in cancer cells. Numerous studies use *in vitro* 2-D cell culture models, and the use of 3-D, *inovo*, and animal models that are very much in alignment with the cutting-edge approaches in oncology therapeutics is increasingly being reported. These studies expand depth and breadth of knowledge about the efficacy of LTP in producing the desired anti-cancer effects and the mechanisms of LTP action further enabling improved methods of plasma delivery. The encouraging aspect of these investigations is that an initial proof-of-concept clinical case study has already been performed [186], and its results can be utilized to design studies that ask practical questions [187].

The widely accepted hypothesis of LTP interaction with cells and tissue is based on the notion that chemical elements of LTP are stimulating or toxic, depending on their concentration [126]. Various unique combinations of these species might provide great potential for activation of specific signaling pathways in cells where no single component is yet identified as the key effector. LTP application to tumor cells can have cytotoxic effects both in vitro and in vivo. Some studies suggest that nonmalignant cells are less sensitive to the same regime of LTP in some studies, though this is not necessarily always the case [188]–[190]. These outcomes are believed to be related to the chemical species in plasma. The sensitivity of cancer cells to either direct treatment or indirect treatment is proposed to be due to redox imbalance and to specific antioxidant systems that are linked to the expression of tumor suppressor genes such as p53 [191].

The basic strategy of indirect LTP treatment is the application of plasma-treated liquids (PTS), such as cell culture medium or aqueous solutions like lactate to kill cancer cells or destroy tumor tissues. The anticancer effects are achieved by the chemical components, including reactive species and other chemically modified substances in these solutions [192]-[194]. Typically, PTS are produced through direct contact of LTP with the solution during treatment. However, PTS can also be produced by the gas discharge within the solution [195], [196]. PTS has been shown to selectively kill many cancer cell lines in vitro or have anti-tumor effects when directly injected in subcutaneous or intraperitoneal tumor models in vivo [197]-[200]. Once produced, PTS is independent of plasma source, which is an advantage over direct plasma treatment. Treatment by PTS eliminates the local effects of electric fields, photons, and temperature. Some cancer cell lines, such as the melanoma cell line B16F10, show only modest cytotoxicity to the reactive species in PTS [201]. PTS also shows promising application in the treatment of metastatic cancers as shown in a recent study that addresses epithelial-mesenchymal transition (EMT). EMT is a process that endows cancer cells with increased stemness, metastatic potential, and resistance to conventional therapies. In this study, PTS was shown to be significantly more effective against cancer cells that have undergone EMT than their epithelial parent cells. This effect is hypothesized to be due to the increased ROS level in the mesenchymally transitioned cells [202]. As a potential pharmacological modality, the stability of PTS is an important consideration needing further investigation [203]–[205].

Unlike the indirect LTP treatment, direct plasma treatment is performed when cells or samples are directly accessible to bulk plasma [206]. A layer of physiological solution typically covers the cells during direct in vitro LTP treatments [207]. Such a layer facilitates the diffusion and formation of reactive species from the gas phase to the liquid phase in the medium to produce desired biological changes in cells [208]. Physical factors in LTP, such as thermal effect, UV, and electric fields, may be largely blocked by such an aqueous layer, particularly when it is adequately deep. Direct and indirect treatment share many similarities [209]: many cellular responses have been widely observed in both treatment strategies, including perturbations in the cell membrane, changes in mitochondrial structure and function (increase in intracellular ROS), and damage to the endoplasmic reticulum and DNA [210]. Cell death is the final goal and fate of most LTP-treated cancer cells [211], [212]. Apoptosis is the primary form of cell death; however, necrosis and autophagy-associated cell death have been also reported [213]–[216].

Direct treatment takes advantage of two unique components: 1) short-lived reactive species and 2) physical factors (such as electromagnetic emission from plasma) otherwise absent in PTS. Under the same experimental conditions, the cytotoxicity of direct treatment is sometimes stronger than indirect treatment [217], [218]. The direct LTP treatment is also suggested to produce additional cellular effects, such as endogenous H2O2 generation by cells [219]-[222] and sensitization of cancer cells to the cytotoxicity of long-lived reactive plasma species, such as  $H_2O_2$  and  $NO_2^-$  [223], contributing to the uniqueness of LTP. Physical plasma factors alone can also sensitize cancer cells to the cytotoxic action of ROS, as seen for nanosecond-pulsed magnetic fields against B16F10 melanoma cells [222] and pulsed electric fields against leukemia cells [223]. A recent study demonstrated that glioblastoma cell lines U87MG and A172 could be sensitized to cytotoxicity of temozolomide (TMZ), a drug used for the treatment of brain tumors, by the electromagnetic emission from a helium-discharged tube [224]. Vice versa, drug-sensitized melanoma cells have been shown to be more susceptible to plasma-induced cell death [225].

Direct LTP treatment has already been widely used in many animal studies. Direct plasma treatment on the skin above a subcutaneous glioblastoma U87MG xenograft tumor site in mice reduced tumor size and prolonged their life [226]. Fractionated treatment was demonstrated to have improved anti-tumor effect compared to a single, longer treatment [227]. Similar anti-tumor effects after direct plasma treatment of subcutaneous tumors of different origins, including melanoma, bladder cancer, pancreatic carcinoma, breast cancer, head and neck cancer, and neuroblastoma in animal models, have been reported [228]–[231].

## B. Challenges and Proposed Solutions

A key challenge in the use of LTP for the treatment of cancers is the identification of its mechanism of action. The contribution of individual plasma components to its antitumor effects remains largely unknown. While most of the research describes the cause-and-effect relationship between chemical species and tumor toxicity, it is still unclear how much the physical factors of LTP contribute to the cellular responses observed. An important, practical consideration to keep in mind is that when cancer cells or tissues are directly exposed to LTP, they experience both physical and chemical effectors simultaneously and as such, the observed biological response could be attributed to the synergistic effect of the physical and chemical factors. In vitro, these effects are difficult to study because liquid layers of dozen of micrometers block physical factors and only allow chemical factors such as reactive species to affect cells. An initial attempt was recently published where cells were treated from the bottom of an inverted multiwell plate or cell culture dish to block all factors except electric fields [232]. This in vitro treatment caused necrotic killing effect on at least six cancer cell lines, including cell lines that are resistant to reactive species [233], [234]. In vivo, necrosis may trigger an immune response or an inflammatory response [235], where physical factors in LTP are very relevant in direct treatment strategies. Furthermore, based on the results of a recent study, the effects of MW emission from a plasma were determined to have decontamination efficacy [236]. This transbarrier capacity of LTP treatment suggests the noninvasive potential in medical application but more extensive studies are needed to understand the mechanism of interaction of physically based modalities with cells and tissues.

The second challenge is the insufficient understanding of the biological impact of LTP on tissues, a prerequisite to explore the underlying mechanism of LTP-based anti-cancer effects. Besides the well-accepted direct cytotoxic effects of LTP on cancer cells, evidence is now accumulating to show that it triggers unique immune responses and immunogenic cell death in vivo, providing clues to help understand LTP effects at the tissue level. Immunogenic cancer cell death (ICD) is elicited in response to various stimuli [237]. Sterile inflammation via damage-associated molecular patterns (DAMPs) overrides the immunosuppressive properties associated with apoptosis, which drives dendritic cell (DC) maturation and anti-tumor T cell responses [238]. The importance of immunity [239], immunogenicity, and antigen presentation [239], ICD [240], and vaccination [241], [242] for plasma cancer treatment has been extensively reviewed recently. Increased immunogenicity has been described in several in vitro studies [148], [243]-[249], and a number of reports suggest an enhancement of the anti-tumor immune function following plasma cancer treatment in vivo [250]-[253]. The mechanism proposed is that plasma-induced oxidative stress elicits ICD, albeit direct in vivo evidence, e.g., via systemically enhanced antioxidants or tumor cell clonotypes with enhanced expression of antioxidant enzymes to scavenge ROS/RNS and thus inhibit (immunogenic) cell death, is lacking. From recent in vitro data, it can be concluded that the role of antioxidant defense molecules such as catalase is less pronounced in terms of plasma-induced cell death induction [254]. In vitro, several studies have suggested the activation of professional antigen-presenting cells (APCs), myeloid cells critical in providing T cell co-stimulation and activation, following plasma treatment or following exposure to plasma-treated tumor cells [133], [255]–[259]. A new research field and potential therapeutic approach has been recently opened up by investigating the immunogenicity of proteins, showing that plasma-treated chicken ovalbumin, a model protein, provided enhanced protein immuno-recognition *in vitro* and tumor growth reduction *in vivo*, [133]. The implications of these findings for tumor vaccination and therapeutics are vast and provide a basis for further investigation [242].

The ultimate goal of plasma oncotherapy is its translation for clinical application and it remains an important topic for continued exploration and discussion among experts in the multiple disciplines that contribute to exciting new developments in plasma medicine. Based on the different ways by which tumor tissue may be exposed to LTP, as described above, it is quite clear that it offers great flexibility in different cancer scenarios. Here, we propose our vision for how LTP may be used for cancer therapy.

For the more accessible tumors, like those that are subcutaneous, a direct, fractionated treatment with few to several treatment cycles over the skin of the tumor site would be easy and practical. Preliminary reports using the kINPen MED plasma jet in patients suffering from metastatic melanoma was however unsuccessful, perhaps because of the thickness of the skin. In contrast, the treatment of ulcerating, locally advanced squamous cell carcinoma of the oropharynx [186] inhibited the growth of tumors in some patients. Incisional biopsies found apoptotic tumor cells and a desmoplastic reaction of connective tissue. In another clinical application, LTP was used to treat oral premalignant lesions (leukoplakia) [260]. Side effects were mild, not life threatening, and transient with no long-term after effects [261]. An improved understanding of the mechanism of LTP action could help improve clinical treatment guidelines for better efficacy. In the case of deep tumors such as intraperitoneal carcinomatosis, while topical LTP treatment is not possible, LTP offers a convenient solution via subcutaneous or intraperitoneal injections. The sensitization of cancer cells to the cytotoxicity of drugs provides the added benefit of a third option of synergy with LTP, as previously suggested in in vitro and translational models using the guideline-based drugs gemcitabine and cisplatin [262]. This LTP-induced selective sensitization of cancer cells is another new direction for further investigations: not aim to kill cancer cells but to sensitize cancer cells to traditional chemotherapy by chemical or physical factors as well as their combinations.

LTP may also be used as an adjunct therapy to standard of care surgery by treating exposed tumor tissues or the remnants of tumor tissue during surgery. In a minimally invasive surgical approach, LTP can be used with endoscopic plasma technology to treat deep tumor tissues [263].

LTP as an adjunct therapy was tested clinically for the first time in a patient with stage 4 colon cancer at Baton Rouge General Medical Center in Baton Rouge, Louisiana, immediately after surgery to remove the tumor [185]. Phase I safety clinical trial was completed in 2021 that involved 20 patients to evaluate the safety of the procedure [264].

An important consideration for translation is the precise and controlled delivery of plasma effectors. The chemical composition, physical emission, and other parameters of LTP can be quickly modulated for specific needs [46], [50], [77], [265], [266]. To this end, an intelligent LTP system should scan the cellular responses to LTP and modify discharge conditions in real time via feedback mechanism based on machine learning [267]. A real-time control of LTP is capable of optimizing the killing effect on cancer cells or tissues while protecting normal cells or tissues [268]. Such an intelligent LTP system is proposed as the self-adaptive LTP source, which may guide the design of the next generation of LTP sources [21]. To build an adaptive LTP system, a multiparametric control system is necessary. The feedback control algorithm may use nonlinear MPC, a strategy used to implement a numerical solution of an open-loop optimization in feedback [48], [269]. The adaptive plasma approach may ultimately realize a personalized LTP-based cancer therapeutic that could be adapted for the treatment of other diseases.

## VIII. MODELING OF LTP-CELL AND LTP-TISSUE INTERACTIONS A. BOGAERTS AND M. YUSUPOV

## A. Current State

For improving plasma medicine applications, a good insight is needed in the interactions of LTP with cells and tissues. This can be obtained by modeling, as it can provide detailed information, even on the molecular level, which might experimentally be less (or not) accessible. Below, we make a distinction between modeling the interaction of LTP with: 1) entire cells or tissue and 2) individual cell components, because they require quite different modeling approaches.

Modeling the Interaction of LTP With Entire Cells or Tissue: The interaction of LTP with entire cells or tissue can up to now only be simulated on a macroscopic scale. As a result, the information is less detailed, but it can give an approximate view on the effect of, e.g., electric fields induced by plasma streamers on tissue, as demonstrated by Babaeva and Kushner [15] for a DBD plasma interacting with wounded skin. The actual skin tissue was mimicked by a cellular structure in the model. In collaboration with Ning and Graves, Babaeva et al. [270] also calculated the ion energy and angular distributions incident on cell membranes, and applied MD simulations of ion sputter probabilities of typical lipidlike material. In some follow-up papers (e.g., [54] and [271]), Babaeva, Tian, Lietz, and Kushner extended their model to wounded skin covered by liquid, containing also blood platelets. The major focus was on the plasma-liquid model, but some description was also given on the electric fields delivered to the blood platelets and cells [54], and on the interaction of the DBD treating liquid-covered tissue [271]. Likewise, Kong and coworkers [272] developed a semi-1-D model for plasmabiofilm interaction and plasma-tissue interaction, based on a reactive penetration model for mass transfer of plasma species across the gas-liquid boundary.

Modeling the Interaction of LTP With Individual Cell Components: Modeling LTP interactions with individual cell components is possible on the molecular level, and much progress has been made in recent years. Depending on the system size and level of detail, different modeling approaches can be used, ranging from quantum mechanics (QM), over density functional theory (DFT) and density functional tight binding (DFTB), to classical MD (see [17] for a detailed description). DFT (and other QM methods with higher accuracy) is typically too time consuming for describing realistic model systems of interest for plasma medicine, but DFTB has been applied, e.g., to study the interaction of ROS with the head group of the PLB [273]. Among the classical MD methods, we distinguish reactive and nonreactive MD simulations. Reactive MD allows the breaking and formation of bonds, and thus, the description of chemical reactions between plasma species and biomolecules, as illustrated, e.g., for the interaction of ROS with peptidoglycan [274] (of interest for bacteria killing), with lipids [270] and DNA [275]. Nonreactive MD, also called "molecular mechanics" (MM), does not consider bond breaking and formation, but it allows to describe large systems and over longer time scales, e.g., for the transport of species through a PLB or the structural deformation and stability of biomolecules. This method has become increasingly attractive in plasma medicine, and some examples will be presented in the next sections.

1) Modeling LTP Interaction With the PLB: When RONS produced by LTP comes in contact with cells, they first "see" the cell membrane, composed of a PLB with proteins embedded. It is of great interest to study the permeability of RONS through the PLB. It can provide information on RONS distribution, mobility, and residence times at the membrane-water interface, as well as their translocation free energy barriers across the PLB, showing whether they can easily reach the cell interior. For instance, Cordeiro [276] studied the transport properties of ROS (i.e., OH, HO<sub>2</sub>, H<sub>2</sub>O<sub>2</sub>, and O<sub>2</sub>) across the PLB, employing nonreactive MD simulations. He found that hydrophobic O<sub>2</sub> is able to accumulate at the PLB interior, whereas hydrophilic OH, HO<sub>2</sub>, and H<sub>2</sub>O<sub>2</sub> mainly stay around the lipid head groups [276].

Furthermore, the PLB can also be oxidized by LTP, and therefore, simulations have been performed to compare the RONS permeability through both native and oxidized PLBs [277]. Fig. 8 illustrates the free energy profiles (FEPs), obtained from so-called US nonreactive MD simulations, of various RONS across both native and 50% oxidized PLBs. The FEP of hydrophilic ROS (i.e., OH, HO<sub>2</sub>, and H<sub>2</sub>O<sub>2</sub>) shows a minimum around the head groups, followed by a steep rise toward the center, thus creating a clear energy barrier to cross the PLB. These results are similar to those mentioned above [276]. Upon oxidation, the hydrophilicity of the PLB rises, resulting in a larger permeability for hydrophilic ROS, as demonstrated by the much lower free energy barrier in Fig. 8(b).

Because the hydrophilic ROS prefers to reside close to the head groups, DFTB simulations were performed to study oxidation of these head groups [273]. HO<sub>2</sub> and H<sub>2</sub>O<sub>2</sub> were found not to chemically react with the head groups, while OH radicals cause detachment of some parts in the lipids, enhancing the membrane fluidity and reducing the lipid order. This may allow RONS to penetrate more easily through the PLB, causing



Fig. 8. FEPs of the hydrophilic (a) and (b) and hydrophobic (c) and (d) RONS, across native (a) and (c) and 50% oxidized (b) and (d) PLBs. The average positions of the head group regions (i.e., phosphate atoms of the PLB) are indicated by light blue color.

further lipid tail (per)oxidation, which might result in pore formation (see further).

The hydrophobic RONS (e.g., O<sub>2</sub>, O<sub>3</sub>, NO, NO<sub>2</sub>, and N<sub>2</sub>O<sub>4</sub>) exhibit very low permeation barriers around the PLB head groups and minima in the center [see Fig. 1(c) and (d)]. Hence, they can cause lipid (per)oxidation in the lipid tail region. Upon oxidation, the PLB can become a bit more fluidic, making the FEPs of the hydrophobic RONS a bit smoother, but otherwise they do not change significantly. These simulations correlate well with experiments, where the permeability of hydrophobic RONS (NO and O<sub>2</sub>) was found to be 3-6 orders of magnitude higher than for H<sub>2</sub>O<sub>2</sub> [278]. Thus, hydrophobic RONS may easily permeate through the cell membrane, while the active transport of hydrophilic ROS in and out of the cell should only be possible in the presence of aquaporin (AQP) channels or pores. US simulations [279] indeed predicted a 100 times higher permeability through AQP than through the PLB, which might explain the selectivity of LTP toward cancer cells, because cancer cells typically have a higher AQP expression in their cell membrane. In addition, pore formation in the cell membrane after lipid (per)oxidation has also been studied by nonreactive MD [280], as well as the combined effect of lipid oxidation and an electric field [281].

Besides the higher expression of AQP, some cancer cells also have a lower cholesterol content in their cell membrane than normal cells. MD simulations [282] revealed higher free energy barriers for hydrophilic ROS at higher cholesterol contents, making it more difficult for them to penetrate through the cell membrane. Furthermore, the FEP of  $O_2$  showed new free energy barriers close to the sterol rings, which will limit the probability of lipid (per)oxidation of the lipid tails, and hence, pore formation. Finally, MD simulations also predicted the absence of pore formation for cholesterol fractions above 15% [280]. Hence, RONS should be able to penetrate more easily through the membrane of cells with lower cholesterol fraction, such as cancer cells, resulting in oxidative stress inside the cell, while this effect might not happen in normal cells, due to their higher cholesterol fraction. This might provide another explanation for the selective anti-cancer effect of LTP.

2) Modeling LTP Interaction With DNA: Reactive MD simulations revealed two main types of reactions of OH radicals with a DNA model system, i.e., H-abstraction and OHaddition [275]. The H-abstractions create a radical, resulting in even more (intermolecular) H-abstractions, possibly leading to single strand breaks (SSBs). Combined with a second SSB at the opposite strand in close vicinity, this may lead to a double strand break (DSB). The latter could however not be observed in these MD simulations, because of their limited time scale (see also Section II). The OH-addition on the purine ring of the nucleic bases, more specifically at the C-8' position of dAMP and dGMP, yielded 8-hydroxypurine adduct radicals (8-OH-Ade or 8-OH-Gua), which is the first step toward 8-oxo-guanine (8-O-Gua) and 2,6-diamino-4-hydroxy-5-formamidopyrimidine (FapydG) formation, i.e., markers for oxidative stress in cells. The latter may affect biochemical pathways within the affected cell, e.g., introduction of DNA mutations or inhibition of gene expressions, possibly leading to apoptosis, but again, the simulated time was not long enough to observe these further reactions, due to the high computational cost of the simulations.

3) Modeling LTP Interaction With Proteins: Nonreactive MD, molecular docking, and US (binding free energy) calculations investigated the structural conformation and binding affinity of the human epidermal growth factor (hEGF; one of the important signaling proteins in both wound healing and cancer treatment) to its receptor (hEGFR) under oxidative stress as induced by LTP [283]. Mild oxidation was found to not significantly affect this binding affinity, and will thus not strongly influence cell signaling pathways, and hence, cell proliferation, which might explain why short plasma treatment times are beneficial for chronic wound healing. On the other hand, the interaction was clearly reduced upon stronger oxidation, which might disturb the cell signaling pathways, ultimately leading to disruption of cell proliferation. This may explain why longer plasma treatment times result in inhibiting cancer cell proliferation and even cancer cell death [283].

In general, the effects of posttranslational modifications (e.g., methylation, phosphorylation, glycosylation, and acetylation) on various protein structures have been widely studied in the literature, but it would also be interesting to study in more detail the effects of oxidation by ROS (and nitrosylation by RNS), on the protein stability, and its binding affinity, specifically in the context of LTP for cancer treatment, which is till now only scarcely investigated. Indeed, such studies would help to further unravel the molecular level mechanisms of cancer cell death induced upon oxidation by LTP.

#### IX. CHALLENGES AND PROPOSED SOLUTIONS

## A. Compromise Between Level of Detail and Calculation Time

The major challenge in modeling LTP interactions with cells or tissues is the compromise between calculation time

Fig. 9. Overview of the various molecular-level modeling approaches for the interaction of reactive species (e.g., RONS) as produced by LTP, with biomolecules, or for the (longer term) effects of these interactions, illustrating the length and time scales that can be reached. QM = quantum mechanics, DFTB = density functional-tight binding, QM/MM = quantum mechanics/molecular mechanics, rMD = reactive molecular dynamics, nrMD = nonreactive molecular dynamics, uaMD = united-atom molecular dynamics, and cgMD = coarse-grained molecular dynamics. The length and time ranges are approximate and improve over time with the increase of supercomputer capacities. Adopted from [17] with permission.

and level of detail of the simulations. Macroscopic modeling is fast, but can only provide a general, approximate picture. Molecular-level modeling yields detailed information, but the small time step needed makes these simulations limited to small system sizes and time scales. In addition, the complexity of cells or tissues makes it difficult to study the processes at the molecular level. Thus, important and specific parts of cells (e.g., the PLB or transmembrane proteins) are often chosen as model systems in molecular-level simulations in order to understand the underlying mechanisms of the LTP effect on cancer cells. Fig. 9 presents the time and length scales of the various molecular-level simulations, mentioned in the previous section.

The computational QM methods, including DFT, are the most accurate, but for plasma medicine, only approximate DFT methods (e.g., DFTB) or classical reactive MD methods would be feasible, keeping in mind the required system sizes of relevant biomolecules. While DFT is more approximate than other QM methods, it is also still very time consuming, and thus able to handle system sizes only in the order of 100 atoms for a simulation time ranging from a few days up to a few weeks (on, e.g., a supercomputer node with dual 14-core Intel E5-2680v4 Broadwell generation CPUs connected through an EDR InfiniBand network). Note that the time range depends on which type of software (e.g., VASP or Gaussian) is employed and whether a simple geometry optimization or deeper characterization is carried out, as well as on the system size, i.e., finite (e.g., cluster) or infinite (i.e., system in a periodic box). DFT-based MD calculations, also called "ab initio MD" (AIMD) can handle time scales in the order of picoseconds. DFTB, on the other hand, can typically handle a few



1000 atoms on time scales of 10 s of picoseconds, making it indeed more suitable for biomedical studies on relevant model systems. Classical reactive MD, in which the forces are not based on QM data but on classical fitting parameters, can handle much longer time scales (order of 1 ps to 100 ns) and larger systems  $(10^3 - 10^5 \text{ atoms})$ , depending on the complexity of the force field (interaction potential), but of course, at the expense of the level of detail. Moreover, the accuracy critically depends on the chosen force fields. Because of the larger system sizes, it has been used more often in the literature in the context of plasma medicine (see Section II-B). However, the simulation time scale is still very limited, and thus, many relevant biochemical processes are still beyond reach, as illustrated in Section I-B2, for the interaction of LTP with DNA. Indeed, DSBs in DNA, and the reaction of 8-OH-Ade and 8-OH-Gua into 8-O-Gua and FapydG (i.e., markers for oxidative stress in cells), could not be observed within the time scale of the MD simulations.

Nonreactive MD can handle much larger system sizes and time scales (typically two orders of magnitude) than reactive MD. The so-called "all-atom force fields" (i.e., when all atoms in the system are treated separately) can describe in the order of  $10^5 - 10^7$  atoms, at time scales of 0.1 ns-10  $\mu$ s. The so-called "united-atom force fields" treat all heavy atoms separately, while the H atoms bound to a C atom (e.g., in the apolar tails of phospholipids) are treated as one (methyl or methylene) group. Hence, this yields less separate particles in the system, allowing to simulate larger systems (typically up to one order of magnitude larger), for the same time scales. Finally, "coarse-grained methods," in which the atoms comprising entire functional groups (i.e., typically, three to five heavy atoms with their H atoms) are represented by socalled coarse-grained particles, further reduce the number of particles in the system, and thus allow even larger system sizes (or to speed up the calculations). This opens possibilities for describing larger biomedical systems, albeit at the expense of the level of detail.

In general, there will always be a tradeoff between calculation time and level of detail/accuracy. Thus, it can be interesting to combine the above methods, for a more comprehensive picture. For instance, Yusupov *et al.* [273] combined reactive (DFTB) MD (to study the interaction of ROS with the head groups of the PLB) with nonreactive (classical) MD (for the subsequent effect of head group and lipid tail oxidation on the structural and dynamic properties of the cell membrane).

Furthermore, while the above example illustrates the use of two separate methods, it is also possible to combine different methods in an integrated simulation, in QM/MM (QM/MM) methods. A small (chemically most relevant) part of the system (e.g., the active site of the biological system) can be described at the quantum chemical (electronic) level, while the surrounding embedding atoms and molecules may be treated at a classical (atomic) level. This method can thus combine the strengths of different modeling approaches, and it would be very interesting to explore its potential in plasma medicine. Indeed, QM/MM is successfully used for many years in biochemistry, e.g., to describe the enzymatic reaction mechanisms (or catalytic properties) of various enzymes [284].

Finally, it would be interesting to apply other enhanced sampling methods in the context of plasma medicine. These methods include US (see Section I-B1), metadynamics (MetaD), steered MD (SMD), replica exchange MD (REMD), and accelerated MD (aMD), which are successfully applied to a wide range of biological systems, to study conformational changes of proteins, protein-protein, and protein-ligand interactions, enzyme catalysis, and so on [285]. Each of these approaches has its advantages and disadvantages from the computational and application perspective. For instance, US is very popular for free energy barrier calculations, because of its quick convergence and ability of simultaneously running multiple independent simulations (windows). However, the harmonic potentials used for US windows need to be tuned manually, to obtain reasonable positional overlaps between the windows along the collective variable (CV, e.g., bilayer normal). Moreover, adding further US windows to the system may be necessary to improve the convergence of the sampling. Finally, US simulations are time consuming for large and complex systems. An advantage of MetaD is that *a priori* knowledge of the end states is not required, and several CVs can be used to explore possible pathways. However, the efficiency of MetaD does not scale well with the number of used CVs, and the accuracy and convergence depends on the choice of the necessary parameters [285]. When little is known about the events to be simulated (e.g., protein folding), REMD or aMD are probably the best methods. More information on these and other enhanced sampling and free energy calculation methods are given in [285].

#### B. Need for Accurate Input Data

Another challenge, related to the first one, is the need for accurate input data in both macroscopic and molecular-level simulations. This certainly applies to the macroscopic simulations, which are based on many assumptions, but even for the most accurate molecular-level methods, the accuracy of the calculation results critically depends on the input data, e.g., the chosen functionals in DFT or the force fields in classical MD. Hence, there is a constant need for improved functionals and force fields, to further improve the predictive power of the calculations. Even more, the lack of some force fields is limiting the possibilities of MD to obtain an overall picture. For instance, the lack of accurate force field parameters of RNS (e.g., HNO<sub>2</sub>, HNO<sub>3</sub>, and ONOOH) for reactive MD simulations makes it up to now impossible to study their interaction with cell components.

#### C. Experimental Validation

Experimental validation is another big challenge, certainly for the molecular-level simulations. Indeed, due to small sizes of biomolecular systems, sophisticated experimental methods, such as neutron scattering, small-angle X-ray scattering (SAXS), NMR, cryogenic electron microscopy (cryo-EM), and electrospray ionization mass spectrometry (ESI-MS), would be needed. Furthermore, dedicated experiments should be designed, under very controlled conditions, generating for instance only a beam of OH radicals, instead of the cocktail of RONS produced by LTP. Moreover, experiments should be performed for well-defined model systems, as considered in the simulations, gradually mimicking more complex tissues. Such experiments are set up in various labs, for instance, for studying the separate and synergistic effects of plasmagenerated radicals and UV/VUV photons at the cellular and molecular level for various kinds of biomolecules, or experiments with simple model systems for the cell membrane, based on synthetic phospholipid membrane vesicles or liposomal model membranes (e.g., [286]–[288]).

For instance, in [273], the MD simulations were validated experimentally, using liposomes as model systems for the biological membranes. The so-called Laurdan Assay was used to determine the phase states of the membranes (gel or liquid phase) by measuring the penetration of water molecules into the bilayer, which strongly correlates with the packing of the phospholipids. In addition, high resolution mass spectrometry was applied for studying modifications of the liposomes after LTP treatment. The experiments revealed a slight initial rise in membrane rigidity, followed by a strong and persistent increase in fluidity (indicating a drop in lipid order) upon LTP treatment, in good agreement with the simulations. Besides, also indirect validation, by means of biological assays, can be very useful, and a few examples are presented in the next section.

# D. Translating the Insights for Improving Biomedical Applications

Finally, while the molecular-level simulations provide detailed insights in, e.g., how RONS can penetrate through the cell membrane, either by passive transport (for hydrophobic RONS) or through pores or AQP channels (for hydrophilic RONS), the long computation time dictates the use of simple model systems (e.g., a simple PLB and/or AQP), while the cell membrane contains many other lipids and proteins as well, such as antiporters, catalase, etc. Indeed, the reality in plasma medicine is much more complicated, and it is not straightforward to translate the insights obtained from these simple model systems to the real world, and thus to actually improve the biomedical applications. For instance, taking the same example of the cell membrane, the various RONS might create a myriad of different lipid oxidation (and nitration) products, which are not yet accounted for in these simulations. Hence, more knowledge is crucial on the different products formed, and how they affect the biophysical properties and function of the cell membrane. Furthermore, lipid peroxidation might lead to the liquid ordered-liquid disordered phase separation in membranes [289], and this might favor pore formation. Thus, it would be interesting to account for phase-separated membranes (or lipid rafts) as well, as they can play a crucial role in cell permeability. Specifically, the interface region between these rafts (e.g., gel and fluid domains), the thickness mismatch between both phases, the membrane elasticity and the lipid packing, all together can affect the membrane permeability [290].

Likewise, studies on LTP-induced oxidation of certain proteins [e.g., hEGF (see Section I-B3)] typically account for only some effects, but in reality, many more proteins play a role in plasma medicine. Specifically, for plasma cancer immunotherapy, it would be important to investigate the interaction between immune cell proteins and cancer cell proteins, under native conditions and oxidative stress (as induced by LTP). Protein–protein interactions have indeed been computationally studied in many different fields (e.g., [291]). While the effect of mutations in proteins and drugs that block the binding of immune checkpoint proteins to receptors has been simulated (e.g., [292]), the interaction between immune cell and cancer cell proteins under oxidative stress, as induced by LTP, has only been the subject of one recent study [293] (see also next paragraph), showing the need for such type of modeling, for a better insight in plasma cancer immunotherapy.

In general, however, molecular-level simulations certainly contribute to a better understanding in plasma medicine, and there are some recent examples in the literature on the combined in silico, in vitro, and even in vivo studies for the effect of LTP on specific biomolecules, where simulations did provide a better understanding of the experiments. For instance, Lin et al. [293] performed a combined in silico, in vitro (with 3-D tumor models), and in vivo study on the effect of LTP on CD47 (a key innate immune checkpoint, which was found to be modulated after LTP treatment). Simulations revealed that the potential oxidized salt-bridges are responsible for conformational changes. US simulations of CD47 demonstrated that oxidation-induced conformational changes reduce its binding affinity with its receptor (signalregulatory protein alpha; SIRP $\alpha$ ), thus providing new insight into the potential of LTP for cancer immunotherapy, i.e., for reducing immunosuppressive signals on the surface of cancer cells. Also, Shaw et al. [294] used MD simulations to support in vitro and in ovo experiments on the synergistic cytotoxic effect of PTL with melittin (a small peptide component of bee venom, with reported anti-cancer effects in vitro and in vivo). Indeed, the simulations revealed that LTP-induced oxidation of the PLB results in a lower energy barrier for translocation of melittin compared to the nonoxidized PLB. Likewise, in [295], the molecular effect of LTP-induced oxidative stress on the interaction between CD44 and hyaluronan (HA) was investigated. Multilevel atomistic simulations revealed a drop in the binding free energy of HA to CD44 upon oxidation, supporting the experimental results, and the hypothesis that LTP-induced oxidation disturbs the CD44-HA interaction, which can inhibit proliferative signaling pathways inside tumor cells, and induce cancer cell death.

Finally, besides the molecular-level simulations, it would be of great interest to study cell signaling pathways, which might also explain the selectivity of plasma for cancer therapy [296]. Indeed, experiments indicated that two apoptosis-inducing signaling pathways, which are both inhibited by catalase in the extracellular compartment, could be reactivated upon inactivation of catalase in the cell membrane of cancer cells. If LTP could inactivate catalase, it may induce cancer cell death upon reactivating these signaling pathways. It is clear that a better understanding of this apoptosis-inducing mechanism and the role of catalase inactivation would be of great value. Such mechanisms can obviously not be simulated by molecularlevel simulations, and macroscopic reaction kinetics modeling would be needed to study the mechanisms of the cell's antioxidant defense system and redox signaling, as extensively used in the field of redox biology (e.g., [297]). Developing such models is very challenging, and would have to be done stepby-step, starting with simple model systems, and gradually adding complexity. This has recently been demonstrated by Bengtson and Bogaerts, who developed a mathematical model to study the role of catalase inactivation for reactivating the two above-mentioned apoptotic signaling pathways, trying to understand the underlying cause of the anti-cancer effect of LTP [298]. In addition, they also developed another model to describe the key species and features of the cellular response, in an attempt to quantify the (possible) selective and synergistic anti-cancer effects of PTL. The latter model investigated the cell susceptibility toward exogenous H<sub>2</sub>O<sub>2</sub> (alone and in combination with  $NO_2^{-}$ ), in terms of two variables, i.e., the H<sub>2</sub>O<sub>2</sub> membrane diffusion rate constant and the intracellular catalase concentration, and predicted that the maximum intracellular H<sub>2</sub>O<sub>2</sub> concentration could be used to quantify the cell susceptibility toward exogenous H<sub>2</sub>O<sub>2</sub> [299]. However, clearly more modeling efforts in this field are needed, to gain further insight in the underlying mechanisms of the (selective) anti-cancer action of LTP.

In general, it is clear that much more research will be needed to elucidate all the underlying mechanisms of LTP for biomedical applications. However, this section showed that computer simulations can definitely contribute to elucidate some pieces of the puzzle.

#### X. CONCLUDING REMARKS

On the fundamental scientific level, the advent of the application of LTP in biology and medicine has allowed for the creation of new scientific knowledge regarding the interaction of the 4th state of matter with soft matter, such as biological cells and tissues. Twenty-five years ago, such knowledge was simply missing. For decades, plasma technology has been playing a major role in several industries that serve our modern society, but its introduction to the biomedical field is comparatively recent. Nonetheless, the potential socioeconomic impact of using AP-LTP to overcome outstanding hygiene, safety, and medical challenges cannot be overstated. As shown in this article, LTP is today a well-established and accepted technology that is helping solve many serious issues related to public health, food and water safety, and others. However, to push the field to a higher level of maturity, more fundamental scientific work and novel advanced application concepts are still needed. To help facilitate this process, the team of experts who contributed to this roadmap paper has proposed viable ideas and potential approaches that should carry the field forward into the next decade.

## ACKNOWLEDGMENT

All authors declare that they have no known conflicts of interest in terms of competing financial interests or personal

relationships that could have an influence or are relevant to the work reported in this article.

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#### REFERENCES

- M. Laroussi, "Sterilization of contaminated matter with an atmospheric pressure plasma," *IEEE Trans. Plasma Sci.*, vol. 24, no. 3, pp. 1188–1191, Jun. 1996.
- [2] E. Stoffels, A. J. Flikweert, W. W. Stoffels, and G. M. W. Kroesen, "Plasma needle: A non-destructive atmospheric plasma source for fine surface treatment of (bio)materials," *Plasma Sources. Sci. Technol.*, vol. 11, no. 4, pp. 383–388, 2002.
- [3] M. Laroussi, "Nonthermal decontamination of biological media by atmospheric-pressure plasmas: Review, analysis, and prospects," *IEEE Trans. Plasma Sci.*, vol. 30, no. 4, pp. 1409–1415, Aug. 2002.
- [4] G. Fridman, G. Friedman, A. Gutsol, A. B. Shekhter, V. N. Vasilets, and A. Fridman, "Applied plasma medicine," *Plasma Process. Polym.*, vol. 5, pp. 503–533, Aug. 2008.
- [5] M. Laroussi, "Low temperature plasmas for medicine?" *IEEE Trans. Plasma Sci.*, vol. 37, no. 6, pp. 714–725, Jun. 2009.
- [6] T. von Woedtke, S. Reuter, K. Masur, K.-D. Weltmann, "Plasmas for medicine," *Phys. Rep.*, vol. 530, no. 4, pp. 291–320, 2013.
- [7] M. Laroussi, "From killing bacteria to destroying cancer cells: 20 years of plasma medicine," *Plasma Process. Polym.*, vol. 11, pp. 1138–1141, Dec. 2014.
- [8] M. Laroussi, M. Kong, G. Morfill, and W. Stolz, *Plasma Medicine:* Applications of Low-Temperature Gas Plasmas in Medicine and Biology. Cambridge, U.K.: Cambridge Univ. Press, 2012.
- [9] A. Fridman and G. Friedman, *Plasma Medicine*. New York, NY, USA: Wiley, 2013.
- [10] S. Toyokuni, Y. Ikehara, F. Kikkawa, and M. Hori, *Plasma Medical Science*. London, U.K.: Academic, 2018.
- [11] H.-R. Metelmann, T. von Woedtke, and K.-D. Weltmann, Comprehensive Clinical Plasma Medicine. Berlin, Germany: Springer. 2018.

- [12] M. Keidar, *Plasma Cancer Therapy*. Cham, Switzerland: Springer Nat., 2020.
- [13] X. Lu, G. V. Naidis, M. Laroussi, S. Reuter, D. B. Graves, and K. Ostrikov, "Reactive species in non-equilibrium atmospheric-pressure plasmas: Generation, transport, and biological effects," *Phys. Rep.*, vol. 630, pp. 1–84, May 2016.
- [14] N. Y. Babaeva and M. J. Kushner, "Dynamics of dielectric barrier discharges over wounded skin," *IEEE Trans. Plasma Sci.*, vol. 39, no. 11, pp. 2964–2965, Nov. 2011.
- [15] N. Y. Babaeva and M. J. Kushner, "Reactive fluxes delivered by dielectric barrier discharge filaments to slightly wounded skin," J. Phys. D, Appl. Phys., vol. 46, no. 2, 2013, Art. no. 025401.
- [16] E. C. Neyts, M. Yusupov, C. C. Verlackt, and A. Bogaerts, "Computer simulations of plasma–biomolecule and plasma–tissue interactions for a better insight in plasma medicine," *J. Phys D, Appl. Phys.*, vol. 47, no. 29, 2014, Art. no. 293001.
- [17] A. Bogaerts *et al.*, "Multi-level molecular modelling for plasma medicine," J. Phys. D, Appl. Phys., vol. 49, no. 5, 2016, 054002.
- [18] M. Laroussi, "Cold plasma in medicine and healthcare: The new frontier in low temperature plasma applications," *Front. Phys.*, vol. 8, p. 74, Mar. 2020.
- [19] M. Laroussi and T. Akan, "Arc-free atmospheric pressure cold plasma jets: A review," *Plasma Process. Polym.*, vol. 4, no. 9, pp. 777–788, 2007.
- [20] M. Laroussi, "Low-temperature plasma jet for biomedical applications: A review," *IEEE Trans. Plasma Sci.*, vol. 43, no. 3, pp. 703–712, Mar. 2015.
- [21] M. G. Kong et al., "Plasma medicine: An introductory review," New J. Phys., vol. 11, Nov. 2009, Art. no. 115012.
- [22] R. Tiede, J. Hirschberg, G. Daeschlein, T. von Woedtke, W. Vioel, and S. Emmert, "Plasma applications: A dermatological view," *Contrib. Plasma Phys.*, vol. 54, pp. 118–130, Feb. 2014.
- [23] D. B. Graves, "The emerging role of reactive oxygen and nitrogen species in redox biology and some implications for plasma applications to medicine and biology," *J. Phys. D, Appl. Phys.*, vol. 45, Jun. 2012, Art. no. 263001.
- [24] T. Xia, A. Kleinheksel, E. M. Lee, Z. Qiao, K. R. Wigginton, and H. L. Clack, "Inactivation of airborne viruses using a packed bed nonthermal plasma reactor," *J. Phys. D, Appl. Phys.*, vol. 52, Apr. 2019, Art. no. 255201.
- [25] E. L. Anderson, P. Turnham, J. R. Griffin, and C. C. Clarke, "Consideration of the aerosol transmission for COVID-19 and public health," *Risk Anal.*, vol. 40, pp. 902–907, May 2020.
- [26] S. E. Hwang, J. H. Chang, B. Oh, and J. Heo, "Possible aerosol transmission of COVID-19 associated with an outbreak in an apartment in Seoul, South Korea, 2020," *Int. J. Infectious Dis.*, vol. 104, pp. 73–76, Mar. 2021.
- [27] H. Gao et al., "Atmospheric-pressure non-equilibrium plasmas for effective abatement of pathogenic biological aerosols," *Plasma Sources Sci. Technol.*, vol. 30, May 2021, Art. no. 053001.
- [28] U. Kogelschatz, "Filamentary, patterned, and diffuse barrier discharges," *IEEE Trans. Plasma Sci.*, vol. 30, no. 4, pp. 1400–1408, Aug. 2002.
- [29] X. Lu, M. Laroussi, and V. Puech, "On atmospheric-pressure nonequilibrium plasma jets and plasma bullets," *Plasma Sources Sci. Technol.*, vol. 21, Apr. 2012, Art. no. 034005.
- [30] X. Lu, Z. Jiang, Q. Xiong, Z. Tang, X. Hu, and Y. Pan, "An 11 cm long atmospheric pressure cold plasma plume for applications of plasma medicine," *Appl. Phys. Lett.*, vol. 92, Feb. 2008, Art. no. 081502.
- [31] S. Reuter, T. Von Woedtke, and K.-D. Weltmann, "The kINPen—A review on physics and chemistry of the atmospheric pressure plasma jet and its applications," *J. Phys. D, Appl. Phys.*, vol. 51, May 2018, Art. no. 233001.
- [32] A. Sakudo and T. Misawa, "Antibiotic-resistant and non-resistant bacteria display similar susceptibility to dielectric barrier discharge plasma," *Int. J. Mol. Sci.*, vol. 21, p. 6326, Aug. 2020.
- [33] Y. Liang *et al.*, "Rapid inactivation of biological species in the air using atmospheric pressure nonthermal plasma," *Environ. Sci. Technol.*, vol. 46, pp. 3360–3368, Mar. 2012.
- [34] N. Nuchtavorn, F. Bek, M. Macka, W. Suntornsuk, and L. Suntornsuk, "Rapid separations of nile blue stained microorganisms as cationic charged species by chip-CE with LIF," *Electrophoresis*, vol. 33, pp. 1421–1426, May 2012.
- [35] J. Golda *et al.*, "Concepts and characteristics of the 'COST reference microplasma jet" *J. Phys. D, Appl. Phys.*, vol. 49, no. 8, 2016, Art. no. 084003.

- [36] S. Reuter, J. S. Sousa, G. D. Stancu, and J.-P. H. van Helden, "Review on VUV to MIR absorption spectroscopy of atmospheric pressure plasma jets," *Plasma Sources Sci. Technol.*, vol. 24, Aug. 2015, Art. no. 054001.
- [37] R. Engeln, B. Klarenaar, and O. Guaitella, "Foundations of optical diagnostics in low-temperature plasmas," *Plasma Sources Sci. Technol.*, vol. 29, Jun. 2020, Art. no. 063001.
- [38] G. Willems, J. Benedikt, and A. Von Keudell, "Absolutely calibrated mass spectrometry measurement of reactive and stable plasma chemistry products in the effluent of a He/H<sub>2</sub>O atmospheric plasma," *J. Phys. D*, *Appl. Phys.*, vol. 50, Jul. 2017, Art. no. 335204.
- [39] R. Ono, "Optical diagnostics of reactive species in atmosphericpressure nonthermal plasma," J. Phys. D, Appl. Phys., vol. 49, Jan. 2016, Art. no. 083001.
- [40] G. Dilecce, "Optical spectroscopy diagnostics of discharges at atmospheric pressure," *Plasma Sources Sci. Technol.*, vol. 23, Feb. 2014, Art. no. 015011.
- [41] H.-P. Li, K. Ostrikov, and W. Sun, "The energy tree: Nonequilibrium energy transfer in collision-dominated plasmas," *Phys. Rep.*, vols. 770–772, pp. 1–45, Nov. 2018.
- [42] B. M. Goldberg, S. Reuter, A. Dogariu, and R. B. Miles, "1D time evolving electric field profile measurements with sub-ns resolution using the E-FISH method," *Opt. Lett.*, vol. 44, no. 15, pp. 3853–3856, 2019.
- [43] P. J. Bruggeman et al., "Plasma-liquid interactions: A review and roadmap," Plasma Sources Sci. Technol., vol. 25, Sep. 2016, Art. no. 053002.
- [44] I. Korolov *et al.*, "Atomic oxygen generation in atmospheric pressure RF plasma jets driven by tailored voltage waveforms in mixtures of He and O<sub>2</sub>" J. Phys. D, Appl. Phys., vol. 54, Jan. 2021, Art. no. 125203.
- [45] A. Mesbah and D. B. Graves, "Machine learning for modeling, diagnostics, and control of non-equilibrium plasmas," J. Phys. D, Appl. Phys., vol. 52, p. 30LT02, May 2019.
- [46] D. Gidon, D. B. Graves, and A. Mesbah, "Effective dose delivery in atmospheric pressure plasma jets for plasma medicine: A model predictive control approach," *Plasma Sources Sci. Technol.*, vol. 26, Jul. 2017, Art. no. 085005.
- [47] O. Baranov et al., "Plasma under control: Advanced solutions and perspectives for plasma flux management in material treatment and nanosynthesis," Appl. Phys. Rev., vol. 4, Nov. 2017, Art. no. 041302.
- [48] D. Gidon, D. B. Graves, and A. Mesbah, "Predictive control of 2D spatial thermal dose delivery in atmospheric pressure plasma jets," *Plasma Sources Sci. Technol.*, vol. 28, Aug. 2019, Art. no. 085001.
- [49] A. D. Bonzanini, D. B. Graves, and A. Mesbah, "Learning-based SMPC for reference tracking under state-dependent uncertainty: An application to atmospheric pressure plasma jets for plasma medicine," *IEEE Trans. Control Syst. Technol.*, early access, Apr. 9, 2021, doi: 10.1109/TCST.2021.3069825.
- [50] E. Gjika *et al.*, "Adaptation of operational parameters of cold atmospheric plasma for in vitro treatment of cancer cells," *ACS Appl. Mater. Interfaces*, vol. 10, pp. 9269–9279, May 2018.
- [51] L. Lin *et al.*, "Introducing adaptive cold atmospheric plasma: The perspective of adaptive cold plasma cancer treatments based on realtime electrochemical impedance spectroscopy," *Phys. Plasmas*, vol. 27, Jun. 2020, Art. no. 063501.
- [52] A. D. Bonzanini, J. A. Paulson, D. B. Graves, and A. Mesbah, "Toward safe dose delivery in plasma medicine using projected neural networkbased fast approximate NMPC," *IFAC-PapersOnLine*, vol. 53, no. 2, pp. 5279–5285, 2020.
- [53] X. Dai, K. Bazaka, D. J. Richard, E. R. W. Thompson, and K. Ostrikov, "The emerging role of gas plasma in oncotherapy," *Trends Biotechnol.*, vol. 36, pp. 1183–1198, Nov. 2018.
- [54] N. Y. Babaeva, W. Tian, and M. J. Kushner, "The interaction between plasma filaments in dielectric barrier discharges and liquid covered wounds: Electric fields delivered to model platelets and cells," *J. Phys. D*, *Appl. Phys.*, vol. 47, May 2014, Art. no. 235201.
- [55] A. Privat-Maldonado, C. Bengtson, J. Razzokov, E. Smits, and A. Bogaerts, "Modifying the tumour microenvironment: Challenges and future perspectives for anticancer plasma treatments," *Cancers*, vol. 11, p. 1920, Dec. 2019.
- [56] J. G. Wang, X. Y. Liu, D. W. Liu, X. P. Lu, and Y. T. Zhang, "Mathematical model of gas plasma applied to chronic wounds," *Phys. Plasmas*, vol. 20, Nov. 2013, Art. no. 113507.
- [57] H. Cheng, J. Xu, X. Li, D. Liu, and X. Lu, "On the dose of plasma medicine: Equivalent total oxidation potential (ETOP)," *Phys. Plasmas*, vol. 27, Jun. 2020, Art. no. 063514.

- [58] X. Pei *et al.*, "Inactivation of a 25.5 μm Enterococcus faecalis biofilm by a room-temperature, battery-operated, handheld air plasma jet," *J. Phys. D, Appl. Phys.*, vol. 45, Apr. 2012, Art. no. 165205.
- [59] A. D. Bonzanini, K. Shao, A. Stancampiano, D. B. Graves, and A. Mesbah, "Perspectives on machine learning-assisted plasma medicine: Towards automated plasma treatment," *IEEE Trans. Radiat. Plasma Med. Sci.*, early access, Feb. 1, 2021, doi: 10.1109/TRPMS.2021.3055727.
- [60] M. Mann et al., DIN SPEC 91315: General Requirements for Plasma Sources in Medicine. Berlin, Germany: Beuth-Verlag, 2014.
- [61] M. Laroussi, "Plasma medicine: A brief introduction," *Plasma*, vol. 1, no. 1, pp. 47–60, 2018, doi: 10.3390/plasma1010005.
- [62] M. Modic, N. P. McLeod, J. M. Sutton, and J. L. Walsh, "Cold atmospheric pressure plasma elimination of clinically important single- and mixed-species biofilms," *Int. J. Antimicrob. Agents*, vol. 49, no. 3, pp. 375–378, 2017, doi: 10.1016/j.ijantimicag.2016.11.022.
- [63] A. Salmon, G. D. Stancu, and C. O. Laux, "Decontamination of endospores by plasma sources on dried surfaces: A review of key parameters and inactivation results," *Front. Phys.*, vol. 9, pp. 1–9, May 2021, doi: 10.3389/fphy.2021.677971.
- [64] S. Bekeschus, A. Kramer, E. Suffredini, T. von Woedtke, and V. Colombo, "Gas plasma technology—An asset to healthcare during viral pandemics such as the COVID-19 crisis?" *IEEE Trans. Radiat. Plasma Med. Sci.*, vol. 4, no. 4, pp. 391–399, Jul. 2020, doi: 10.1109/trpms.2020.3002658.
- [65] Z. Chen, G. Garcia, Jr., V. Arumugaswami, and R. E. Wirz, "Cold atmospheric plasma for SARS-CoV-2 inactivation," *Phys. Fluids*, vol. 32, no. 11, pp. 1–6, 2020, doi: 10.1063/5.0031332.
- [66] Y. Morabit, R. D. Whalley, E. Robert, M. I. Hasan, and J. L. Walsh, "Turbulence and entrainment in an atmospheric pressure dielectric barrier plasma jet," *Plasma Process. Polym.*, vol. 17, no. 6, pp. 1–12, 2020, doi: 10.1002/ppap.201900217.
- [67] M. J. Pavlovich, D. S. Clark, and D. B. Graves, "Quantification of air plasma chemistry for surface disinfection," *Plasma Sources Sci. Technol.*, vol. 23, no. 6, 2014, Art. no. 065036, doi: 10.1088/0963-0252/23/6/065036.
- [68] L. Guo *et al.*, "Mechanism of virus inactivation by cold atmospheric-pressure plasma and plasma-activated water," *Appl. Environ. Microbiol.*, vol. 84, no. 17, Sep. 2018, Art. no. e00726-18, doi: 10.1128/AEM.00726-18.
- [69] L. Guo et al., "Plasma-activated water: An alternative disinfectant for S protein inactivation to prevent SARS-CoV-2 infection," Chem. Eng. J., vol. 421, Oct. 2021, Art. no. 127742, doi: 10.1016/j.cej.2020.127742.
- [70] P. Brown, E. H. Rau, P. Lemieux, B. K. Johnson, A. E. Bacote, and D. C. Gajdusek, "Infectivity studies of both ash and air emissions from simulated incineration of scrapie- contaminated tissues," *Environ. Sci. Technol.*, vol. 38, no. 22, pp. 6155–6160, 2004, doi: 10.1021/es040301z.
- [71] J. Julák, O. Janoušková, V. Scholtz, and K. Holada, "Inactivation of prions using electrical dc discharges at atmospheric pressure and ambient temperature," *Plasma Process. Polym.*, vol. 8, no. 4, pp. 316–323, 2011, doi: 10.1002/ppap.201000100.
- [72] S. Hubner, J. S. Sousa, J. Van Der Mullen, and W. G. Graham, "Thomson scattering on non-thermal atmospheric pressure plasma jets," *Plasma Sources Sci. Technol.*, vol. 24, no. 5, 2015, Art. no. 054005, doi: 10.1088/0963-0252/24/5/054005.
- [73] J. B. Schmidt *et al.*, "Femtosecond, two-photon-absorption, laserinduced-fluorescence (fs-TALIF) imaging of atomic hydrogen and oxygen in non-equilibrium plasmas," *J. Phys. D, Appl. Phys.*, vol. 50, no. 1, 2017, Art. no. 015204, doi: 10.1088/1361-6463/50/1/015204.
- [74] S. A. Norberg, W. Tian, E. Johnsen, and M. J. Kushner, "Atmospheric pressure plasma jets interacting with liquid covered tissue: Touching and not-touching the liquid," *J. Phys. D, Appl. Phys.*, vol. 47, no. 47, 2014, Art. no. 475203, doi: 10.1088/0022-3727/47/47/475203.
- [75] C. Ye *et al.*, "An improved enzyme-linked focus formation assay revealed baloxavir acid as a potential antiviral therapeutic against hantavirus infection," *Front. Pharmacol.*, vol. 10, pp. 1–8, Oct. 2019, doi: 10.3389/fphar.2019.01203.
- [76] C. L. Lawson *et al.*, "Cryo-EM model validation recommendations based on outcomes of the 2019 EMDataResource challenge," *Nat. Methods*, vol. 18, no. 2, pp. 156–164, 2021, doi: 10.1038/s41592-020-01051-w.
- [77] L. Lin and M. Keidar, "A map of control for cold atmospheric plasma jets: From physical mechanisms to optimizations," *Appl. Phys. Rev.*, vol. 8, no. 1, 2021, Art. no. 011306, doi: 10.1063/5.0022534.

- [78] P. Ranieri *et al.*, "Plasma agriculture: Review from the perspective of the plant and its ecosystem," *Plasma Process. Polym.*, vol. 18, Jan. 2021, Art. no. e2000162, doi: 10.1002/ppap.202000162.
- [79] S. Herianto, C.-Y. Hou, C.-M. Lin, and H.-L. Chen, "Nonthermal plasma-activated water: A comprehensive review of this new tool for enhanced food safety and quality," *Comprehensive Rev. Food Sci. Food Safety*, vol. 20, pp. 583–626, Jan. 2021, doi: 10.1111/1541-4337.12667.
- [80] M. Laroussi, "A brief note on the history of the dielectric barrier discharge and its application for biological decontamination," *IEEE Trans. Radiat. Plasma Med. Sci.*, early access, Jan. 14, 2021, doi: 10.1109/TRPMS.2021.3051805.
- [81] F. Kogelheide *et al.*, "The role of humidity and UV-C emission in the inactivation of *B. subtilis* spores during atmospheric-pressure dielectric barrier discharge treatment," *J. Phys. D, Appl. Phys.*, vol. 53 no. 29, 2020, Art. no. 295201, doi: 10.1088/1361-6463/ab77cc.
- [82] I. Joshi, D. Salvi, D. W. Schaffner, and M. V. Karwe, "Characterization of microbial inactivation using plasma-activated water and plasmaactivated acidified buffer," *J. Food Prot.*, vol. 81, pp. 1472–1480, Aug. 2018, doi: 10.4315/0362-028X.JFP-17-487.
- [83] Y.-M. Zhao, A. Patange, D.-W. Sun, and B. Tiwari, "Plasma-activated water: Physicochemical properties, microbial inactivation mechanisms, factors influencing antimicrobial effectiveness, and applications in the food industry," *Comprehensive Rev. Food Sci Food Safety*, vol. 19, pp. 3951–3979, Nov. 2020, doi: 10.1111/1541-4337.12644.
- [84] Q. Xiang, L. Fan, Y. Li, S. Dong, K. Li, and Y. Bai, "A review on recent advances in plasma-activated water for food safety: Current applications and future trends," *Crit. Rev. Food Sci. Nutr.*, to be published, doi: 10.1080/10408398.2020.1852173.
- [85] P. Ranieri *et al.*, "Microsecond-pulsed dielectric barrier discharge plasma-treated mist for inactivation of escherichia coli *In Vitro*," *IEEE Trans. Plasma Sci.*, vol. 47, no. 1, pp. 395–402, Jan. 2019, doi: 10.1109/TPS.2018.2878971.
- [86] J. Kruszelnicki, A. M. Lietz, and M. J. Kushner, "Atmospheric pressure plasma activation of water droplets," *J. Phys. D, Appl. Phys.* vol. 52, Jul. 2019, Art. no. 355207, doi: 10.1088/1361-6463/ab25dc.
- [87] K. Tachibana and T. Nakamura, "Comparative study of discharge schemes for production rates and ratios of reactive oxygen and nitrogen species in plasma activated water," *J. Phys. D, Appl. Phys.* vol. 52, no. 38, 2019, Art. no. 385202, doi: 10.1088/1361-6463/ab2529.
- [88] M. Rodríguez-Ruiz, R. Zuccarelli, J. M. Palma, F. J. Corpas, and L. Freschi, "Biotechnological application of nitric oxide and hydrogen peroxide in plants," in *Nitric Oxide and Hydrogen Peroxide Signaling in Higher Plants.* Cham, Switzerland: Springer, 2019, pp. 245–270, doi: 10.1007/978-3-030-11129-8\_12
- [89] P. Lukes, E. Dolezalova, I. Sisrova, and M. Clupek, "Aqueous-phase chemistry and bactericidal effects from an air discharge plasma in contact with water: Evidence for the formation of peroxynitrite through a pseudo-second-order post-discharge reaction of H<sub>2</sub>O<sub>2</sub> and HNO<sub>2</sub>" *Plasma Sources Sci. Technol.*, vol. 23, Feb. 2014, Art. no. 015019, doi: 10.1088/0963-0252/23/1/015019
- [90] B. Myers et al., "Measuring plasma-generated ·OH and O atoms in liquid using EPR spectroscopy and the non-selectivity of the HTA assay," J. Phys. D, Appl. Phys., vol. 54, Jan. 2021, Art. no. 145202, doi: 10.1088/1361-6463/abd9a6
- [91] J. S. Clements, M. Sato, and R. H. Davis, "Preliminary investigation of prebreakdown phenomena and chemical reactions using a pulsed high-voltage discharge in water," *IEEE Trans. Ind. Appl.*, vol. IA-23, no. 2, pp. 224–235, Mar. 1987, doi: 10.1109/TIA.1987.4504897.
- [92] P. Ajo, S. Preis, T. Vornamo, M. Mänttäri, M. Kallioinen, and M. Louhi-Kultanen, "Hospital wastewater treatment with pilot-scale pulsed corona discharge for removal of pharmaceutical residues," *J. Environ. Chem. Eng.*, vol. 6, pp. 1569–1577, Apr. 2018, doi: 10.1016/j.jece.2018.02.007.
- [93] C. Nau-Hix *et al.*, "Field demonstration of a pilot-scale plasma reactor for the rapid removal of poly- and perfluoroalkyl substances in groundwater," *ACS EST Water*, vol. 1, pp. 680–687, Jan. 2021, doi: 10.1021/acsestwater.0c00170.
- [94] B. N. Nzeribe, M. Crimi, S. M. Thagard, and T. M. Holsen, "Physicochemical processes for the treatment of per- and polyfluoroalkyl substances (PFAS): A review," *Crit. Rev. Environ. Sci. Technol.*, vol. 49, no. 10, pp. 866–915, 2019, doi: 10.1080/10643389.2018.1542916.
- [95] M. A. Malik, "Water purification by plasmas: Which reactors are most energy efficient?" *Plasma Chem. Plasma Process.*, vol. 30, pp. 21–31, Feb. 2010, doi: 10.1007/s11090-009-9202-2.

- [96] K. Kelly-Wintenberg *et al.*, "Air filter sterilization using a one atmosphere uniform glow discharge plasma (the volfilter)," *IEEE Trans. Plasma Sci.*, vol. 28, no. 1, pp. 64–71, Feb. 2000, doi: 10.1109/27.842866.
- [97] H. Mohamed *et al.*, "Non-thermal plasma as a novel strategy for treating or preventing viral infection and associated disease," *Front. Phys.*, vol. 9, Jun. 2021, Art. no. 683118, doi: 10.3389/fphy.2021.683118.
- [98] National Academies of Sciences, Engineering, and Medicine, Plasma Science: Enabling Technology, Sustainability, Security, and Exploration. Washington, DC, USA: Nat. Acad. Press, 2021.
- [99] A. Fridman and G. Friedman, *Plasma Medicine*. Chichester, U.K.: Wiley, 2013, pp. 340–343, doi: 10.1002/9781118437704.
- [100] H. Taghvaei, V. S. S. K. Kondeti, and P. J. Bruggeman, "Decomposition of crystal violet by an atmospheric pressure RF plasma jet: The role of radicals, ozone, near-interfacial reactions and convective transport," *Plasma Chem. Plasma Process.*, vol. 39, no. 4, pp. 729–749, 2019, doi: 10.1007/s11090-019-09965-w
- [101] X. Feng, B. Yan, W. Lu, Y. Jin, and Y. Cheng, "Visualization of coupled mass transfer and reaction in a gas-liquid dielectric barrier discharge reactor," *Chem. Eng. J.*, vol. 245, pp. 47–55, Jun. 2014, doi: 10.1016/j.cej.2014.01.054.
- [102] M. Laroussi, "Low temperature plasma-based sterilization: Overview and state-of-the-art," *Plasma Process. Poly.*, vol. 2, no. 5, pp. 391–400, 2005, doi: 10.1002/ppap.200400078.
- [103] V. I. Khrupkin, L. V. Pisarenko, S. M. Slostin, A. V. Pekshev, L. N. Bakunova, and V. A. Piatenko, "Use of physical plasma in surgery of wounds and wound complications," *Vestn Khir Im I I Grek*, vol. 157, no. 2, pp. 43–47, 1998.
- [104] A. Schmidt, G. Liebelt, F. Nießner, T. von Woedtke, and S. Bekeschus, "Gas plasma-spurred wound healing is accompanied by regulation of focal adhesion, matrix remodeling, and tissue oxygenation," *Redox Biol.*, vol. 38, Jan. 2021, Art. no. 101809, doi: 10.1016/j.redox.2020.101809.
- [105] G. Isbary *et al.*, "A first prospective randomized controlled trial to decrease bacterial load using cold atmospheric argon plasma on chronic wounds in patients," *Brit. J. Dermatol.*, vol. 163, no. 1, pp. 78–82, 2010, doi: 10.1111/j.1365-2133.2010.09744.x.
- [106] G. Isbary *et al.*, "Successful and safe use of 2 min cold atmospheric argon plasma in chronic wounds: Results of a randomized controlled trial," *Brit. J. Dermatol.*, vol. 167, no. 2, pp. 404–410, 2012, doi: 10.1111/j.1365-2133.2012.10923.x.
- [107] R. Laurita *et al.*, "Cold atmospheric plasma treatment of infected skin tissue: Evaluation of sterility, viability, and integrity," *IEEE Trans. Radiat. Plasma Med. Sci.*, vol. 1, no. 3, pp. 275–279, May 2017, doi: 10.1109/trpms.2017.2679010.
- [108] A. Kramer *et al.*, "Polypragmasia in the therapy of infected wounds— Conclusions drawn from the perspectives of low temperature plasma technology for plasma wound therapy," *GMS Krankenhhyg Interdiszip*, vol. 3, no. 1, p. Doc13, 2008.
- [109] T. von Woedtke, A. Schmidt, S. Bekeschus, K. Wende, and K.-D. Weltmann, "Plasma medicine: A field of applied redox biology," *In Vivo*, vol. 33, no. 4, pp. 1011–1026, 2019, doi: 10.21873/invivo.11570.
- [110] R. Rutkowski *et al.*, "Hyperspectral imaging for in vivo monitoring of cold atmospheric plasma effects on microcirculation in treatment of head and neck cancer and wound healing," *Clin. Plasma Med.*, vols. 7–8, pp. 52–57, Dec. 2017, doi: 10.1016/j.cpme.2017.09.002.
- [111] G. Daeschlein *et al.*, "Hyperspectral imaging: Innovative diagnostics to visualize hemodynamic effects of cold plasma in wound therapy," *Biomed. Eng.*, vol. 63, no. 5, pp. 603–608, 2018, doi: 10.1515/bmt-2017-0085.
- [112] A. Schmidt, F. Nießner, T. von Woedtke, and S. Bekeschus, "Hyperspectral imaging of wounds reveals augmented tissue oxygenation following cold physical plasma treatment in vivo," *IEEE Trans. Radiat. Plasma Med. Sci.*, vol. 5, no. 3, pp. 412–419, May 2021, doi: 10.1109/trpms.2020.3009913.
- [113] H. Y. Kim *et al.*, "Characterization and effects of Ar/air microwave plasma on wound healing," *Plasma Process. Polym.*, vol. 12, no. 12, pp. 1423–1434, 2015, doi: 10.1002/ppap.201500017.
- [114] Nasruddin *et al.*, "A simple technique to improve contractile effect of cold plasma jet on acute mouse wound by dropping water," *Plasma Process. Polym.*, vol. 12, no. 10, pp. 1128–1138, 2015, doi: 10.1002/ppap.201400236.

- [115] M.-H. N. Thi, P.-L. Shao, J.-D. Liao, C.-C. K. Lin, and H.-K. Yip, "Enhancement of angiogenesis and epithelialization processes in mice with burn wounds through ROS/RNS signals generated by nonthermal N2/Ar micro-plasma," *Plasma Process. Polym.*, vol. 11, no. 11, pp. 1076–1088, 2014, doi: 10.1002/ppap.201400072.
- [116] A. Schmidt, S. Bekeschus, K. Wende, B. Vollmar, and T. von Woedtke, "A cold plasma jet accelerates wound healing in a murine model of fullthickness skin wounds," *Exp. Dermatol.*, vol. 26, no. 2, pp. 156–162, 2017, doi: 10.1111/exd.13156.
- [117] P.-L. Shao, J.-D. Liao, T.-W. Wong, Y.-C. Wang, S. Leu, and H.-K. Yip, "Enhancement of wound healing by non-thermal N2/Ar micro-plasma exposure in mice with fractional-CO2-laser-induced wounds," *PLoS ONE*, vol. 11, no. 6, 2016, Art. no. e0156699, doi: 10.1371/journal.pone.0156699.
- [118] T. Hirata, T. Kishimoto, C. Tsutsui, T. Kanai, and A. Mori, "Healing burns using atmospheric pressure plasma irradiation," *Jpn. J. Appl. Phys.*, vol. 53, no. 1, 2014, Art. no. 010302, doi: 10.7567/Jjap.53.010302.
- [119] Y.-W. Hung, L.-T. Lee, Y.-C. Peng, C.-T. Chang, Y.-K. Wong, and K.-C. Tung, "Effect of a nonthermal-atmospheric pressure plasma jet on wound healing: An animal study," *J. Chin. Med. Assoc.*, vol. 79, no. 6, pp. 320–328, 2016, doi: 10.1016/j.jcma.2015.06.024.
- [120] E. Martines *et al.*, "Wound healing improvement in large animals using an indirect helium plasma treatment," *Clin. Plasma Med.*, vols. 17–18, May/Jun. 2020, Art. no. 100095, doi: 10.1016/j.cpme.2020.100095.
- [121] A. Schmidt, T. von Woedtke, B. Vollmar, S. Hasse, and S. Bekeschus, "Nrf2 signaling and inflammation are key events in physical plasmaspurred wound healing," *Theranostics*, vol. 9, no. 4, pp. 1066–1084, 2019, doi: 10.7150/thno.29754.
- [122] S. Arndt *et al.*, "Effects of cold atmospheric plasma (CAP) on β-defensins, inflammatory cytokines, and apoptosis-related molecules in keratinocytes in vitro and in vivo," *PLoS ONE*, vol. 10, no. 3, 2015, Art. no. e0120041, doi: 10.1371/journal.pone.0120041.
- [123] S. Arndt, A. Schmidt, S. Karrer, and T. von Woedtke, "Comparing two different plasma devices kINPen and Adtec SteriPlas regarding their molecular and cellular effects on wound healing," *Clin. Plasma Med.*, vol. 9, pp. 24–33, Mar. 2018, doi: 10.1016/j.cpme.2018.01.002.
- [124] S. Arndt, P. Unger, M. Berneburg, A.-K. Bosserhoff, and S. Karrer, "Cold atmospheric plasma (CAP) activates angiogenesis-related molecules in skin keratinocytes, fibroblasts and endothelial cells and improves wound angiogenesis in an autocrine and paracrine mode," *J. Dermatol. Sci.*, vol. 89, no. 2, pp. 181–190, 2018, doi: 10.1016/j.jdermsci.2017.11.008.
- [125] S. Arndt *et al.*, "Cold atmospheric plasma (CAP) changes gene expression of key molecules of the wound healing machinery and improves wound healing in vitro and in vivo," *PLoS One*, vol. 8, no. 11, 2013, Art. no. e79325, doi: 10.1371/journal.pone.0079325.
- [126] A. Privat-Maldonado et al., "ROS from physical plasmas: Redox chemistry for biomedical therapy," Oxid Med. Cell. Longevity, vol. 2019, Oct. 2019, Art. no. 9062098, doi: 10.1155/2019/9062098.
- [127] S. Bekeschus, A. Schmidt, K.-D. Weltmann, and T. von Woedtke, "The plasma jet kINPen—A powerful tool for wound healing," *Clin. Plasma Med.*, vol. 4, no. 1, pp. 19–28, 2016, doi: 10.1016/j.cpme.2016.01.001.
- [128] L. Boeckmann *et al.*, "Current indications for plasma therapy in dermatology," *Der Hautarzt*, vol. 71, no. 2, pp. 109–113, 2020, doi: 10.1007/s00105-019-04530-0.
- [129] L. Boeckmann *et al.*, "Cold atmospheric pressure plasma in wound healing and cancer treatment," *Appl. Sci.*, vol. 10, no. 19, p. 6898, 2020, doi: 10.3390/app10196898.
- [130] H. Weavers, J. Liepe, A. Sim, W. Wood, P. Martin, and M. P. H. Stumpf, "Systems analysis of the dynamic inflammatory response to tissue damage reveals spatiotemporal properties of the wound attractant gradient," *Current Biol.*, vol. 26, no. 15, pp. 1975–1989, 2016, doi: 10.1016/j.cub.2016.06.012.
- [131] C. C. Winterbourn, "The challenges of using fluorescent probes to detect and quantify specific reactive oxygen species in living cells," *Biochimica et Biophysica Acta*, vol. 1840, no. 2, pp. 730–738, 2014, doi: 10.1016/j.bbagen.2013.05.004.
- [132] B. Kalyanaraman *et al.*, "Measuring reactive oxygen and nitrogen species with fluorescent probes: Challenges and limitations," *Free Radical Biol. Med.*, vol. 52, no. 1, pp. 1–6, 2012, doi: 10.1016/j.freeradbiomed.2011.09.030.
- [133] R. Clemen *et al.*, "Gas plasma technology augments ovalbumin immunogenicity and OT-II T cell activation conferring tumor protection in mice," *Adv. Sci.*, vol. 8, no. 10, 2021, Art. no. 2003395, doi: 10.1002/advs.202003395.

- [134] T. Heusler, G. Bruno, S. Bekeschus, J.-W. Lackmann, T. von Woedtke, and K. Wende, "Can the effect of cold physical plasma-derived oxidants be transported via thiol group oxidation?" *Clin. Plasma Med.*, vol. 14, Jun. 2019, Art. no. 100086, doi: 10.1016/j.cpme.2019.100086.
- [135] R. Strollo *et al.*, "Antibodies to oxidized insulin improve prediction of type 1 diabetes in children with positive standard islet autoantibodies," *Diabetes Metab. Res. Rev.*, vol. 35, no. 4, p. e3132, 2019, doi: 10.1002/dmrr.3132.
- [136] I. Verrastro, S. Pasha, K. T. Jensen, A. R. Pitt, and C. M. Spickett, "Mass spectrometry-based methods for identifying oxidized proteins in disease: Advances and challenges," *Biomolecules*, vol. 5, no. 2, pp. 378–411, 2015, doi: 10.3390/biom5020378.
- [137] R. Clemen, and S. Bekeschus, "Oxidatively modified proteins: Cause and control of diseases," *Appl. Sci.*, vol. 10, no. 18, p. 6419, 2020, doi: 10.3390/app10186419.
- [138] J. V. Dovi, A. M. Szpaderska, and L. A. DiPietro, "Neutrophil function in the healing wound: Adding insult to injury?" *Thromb. Haemost.*, vol. 92, no. 2, pp. 275–280, 2004, doi: 10.1160/TH03-11-0720.
- [139] J. N. Jacobsen, A. S. Andersen, M. K. Sonnested, I. Laursen, B. Jorgensen, and K. A. Krogfelt, "Investigating the humoral immune response in chronic venous leg ulcer patients colonised with Pseudomonas aeruginosa," *Int. Wound J.*, vol. 8, no. 1, pp. 33–43, 2011, doi: 10.1111/j.1742-481X.2010.00741.x.
- [140] J. E. Park and A. Barbul, "Understanding the role of immune regulation in wound healing," *Amer. J. Surg.*, vol. 187, no. 5A, pp. 11S–16S, 2004, doi: 10.1016/S0002-9610(03)00296-4.
- [141] T. Maho, X. Damany, S. Dozias, J.-M. Pouvesle, and E. Robert, "Atmospheric pressure multijet plasma sources for cancer treatments," *Clin. Plasma Med.*, vol. 9, pp. 3–4, Feb. 2018, doi: 10.1016/j.cpme.2017.12.005.
- [142] O. J. Lee *et al.*, "An experimental burn wound-healing study of non-thermal atmospheric pressure microplasma jet arrays," *J. Tissue Eng. Regen. Med.*, vol. 10, no. 4, pp. 348–357, 2016, doi: 10.1002/term.2074.
- [143] H. A. Aboubakr *et al.*, "Virucidal effect of cold atmospheric gaseous plasma on feline calicivirus, a surrogate for human norovirus," *Appl. Environ. Microbiol.*, vol. 81, no. 11, pp. 3612–3622, 2015, doi: 10.1128/AEM.00054-15.
- [144] S. Bekeschus *et al.*, "The feed gas composition determines the degree of physical plasma-induced platelet activation for blood coagulation," *Plasma Sources Sci. Technol.*, vol. 27, no. 3, 2018, Art. no. 034001, doi: 10.1088/1361-6595/aaaf0e.
- [145] J. Kim, B. Ghimire, S. Lim, E. H. Choi, H.-K. Park, and N. K. Kaushik, "Coagulation, deformability, and aggregation of RBCs and platelets following exposure to dielectric barrier discharge plasma with the use of different feeding gases," *J. Phys. D, Appl. Phys.*, vol. 52, no. 15, 2019, Art. no. 155202, doi: 10.1088/1361-6463/ab0198.
- [146] D. Purevdorj, N. Igura, O. Ariyada, and I. Hayakawa, "Effect of feed gas composition of gas discharge plasmas on Bacillus pumilus spore mortality," *Lett. Appl. Microbiol.*, vol. 37, no. 1, pp. 31–34, 2003, doi: 10.1046/j 1472-765X.2003.01341.x.
- [147] H. Jablonowski, A. Schmidt-Bleker, K. D. Weltmann, T. von Woedtke, and K. Wende, "Non-touching plasma–liquid interaction—Where is aqueous nitric oxide generated?" *Phys. Chem. Chem. Phys.*, vol. 20, no. 39, pp. 25387–25398, 2018, doi: 10.1039/c8cp02412j.
- [148] S. Bekeschus, R. Clemen, F. Nießner, S. K. Sagwal, E. Freund, and A. Schmidt, "Medical gas plasma jet technology targets murine melanoma in an immunogenic fashion," *Adv. Sci.*, vol. 7, no. 10, 2020, Art. no. 1903438, doi: 10.1002/advs.201903438.
- [149] Y. Lee *et al.*, "Wound healing effect of nonthermal atmospheric pressure plasma jet on a rat burn wound model: A preliminary study," *J. Burn Care Res.*, vol. 40, no. 6, pp. 923–929, 2019, doi: 10.1093/jbcr/irz120.
- [150] B. S. Lou *et al.*, "Helium/argon-generated cold atmospheric plasma facilitates cutaneous wound healing," *Front. Bioeng. Biotechnol.*, vol. 8, p. 683, Jun. 2020, doi: 10.3389/fbioe.2020.00683.
- [151] M. Scacchi, "The development of the ITI dental implant system. Part 1: A review of the literature," *Clin. Oral Implants Res.*, vol. 11, no. S1, pp. 8–21, 2000.
- [152] R. E. Baier, J. M. Carter, S. E. Sorensen, A. E. Meyer, B. D. McGowan, and S. A. Kasprzak, "Radiofrequency gas plasma (glow discharge) disinfection of dental operative instruments, including handpieces," *J. Oral Implantol.*, vol. 18, no. 3, pp. 236–242, 1992.
- [153] J. Ehrman, "Gas-plasma 'microwave' sterilizes instruments," *Dentist*, vol. 67, no. 8, pp. 23–27, 1989.
- [154] M. H. Anderson, "Gas plasma sterilization: Innovation in practice," Navy Med., vol. 80, no. 5, pp. 9–10, 1989.

- [155] J.-H. Kim, M.-A. Lee, G.-J. Han, and B.-H. Cho, "Plasma in dentistry: A review of basic concepts and applications in dentistry," *Acta Odontologica Scandinavica*, vol. 72, no. 1, pp. 1–12, 2014.
- [156] T.E. Miller, S. Margalit, and T. J. Creamer, "Emergency direct/indirect polyethylene-ribbon-reinforced composite resin, fixed partial denture: A case report," *Compendium Continu. Educ. Dent.*, vol. 17, no. 2, p. 182, 1996.
- [157] G. Jungbauer, D. Moser, S. Muller, W. Pfister, A. Sculean, S. Eick, "The antimicrobial effect of cold atmospheric plasma against dental pathogens-a systematic review of in-vitro studies," *Antibiotics*, vol. 10, no. 2, p. 211, 2021.
- [158] M. M. Awad *et al.*, "Effect of non-thermal atmospheric plasma on micro-tensile bond strength at adhesive/dentin interface: A systematic review," *Materials*, vol. 14, no. 4, p. 1026, 2021.
- [159] W. L. Hui, V. Perrotti, F. Iaculli, A. Piattelli, and A. Quaranta, "The emerging role of cold atmospheric plasma in implantology: A review of the literature," *Nanomaterials*, vol. 10, no. 8, p. 1505, 2020.
- [160] S. Wu, Y. Cao, and X. Lu, "The state of the art of applications of atmospheric-pressure nonequilibrium plasma jets in dentistry," *IEEE Trans. Plasma Sci.*, vol. 44, no. 2, pp. 134–151, Feb. 2016.
- [161] G. McCombs, M. Darby, and M. Laroussi, "Dental applications," in *Plasma Medicine*, M. Laroussi, M. Kong, G. Morfill, and W. Stolz, Eds. Cambridge, U.K.: Cambridge Univ. Press, 2012.
- [162] L. W. Figueira, B. H. D. Panariello, C. Y. Koga-Ito, and S. Duarte, "Low-temperature plasma as an approach for inhibiting a multi-species cariogenic biofilm," *Appl. Sci.*, vol. 11, no. 2, p. 570, 2021.
- [163] F. Theinkom *et al.*, "Antibacterial efficacy of cold atmospheric plasma against Enterococcus faecalis planktonic cultures and biofilms in vitro," *Plos ONE*, vol. 14, no. 11, 2019, Art. no. e0223925.
- [164] C. Jiang, C. Schaudinn, D. E. Jaramillo, P. Webster, and J. W. Costerton, "In vitro antimicrobial effect of a cold plasma jet against enterococcus faecalis biofilms," *ISRN Dent.*, vol. 2012, no. 5, 2012, Art. no. 295736.
- [165] W. L. Hui *et al.*, "Novel technique using cold atmospheric plasma coupled with air-polishing for the treatment of titanium discs grown with biofilm: An in-vitro study," *Dent. Mater.*, vol. 37, no. 2, pp. 359–369, 2021.
- [166] J.-Y. Lee *et al.*, "The bactericidal effect of an atmospheric-pressure plasma jet on Porphyromonas gingivalis biofilms on sandblasted and acid-etched titanium discs," *J. Periodontal Implant Sci.*, vol. 49, pp. 319–329, Oct. 2019.
- [167] Y. Yao *et al.*, "In vitro and in vivo research of atmosphere pressure nonequilibrium plasmas on root canal disinfection: Implication for alternative strategy for irrigation," *Clin. Oral Investig*, vol. 25, pp. 5833–5842, Mar. 2021, doi: 10.1007/s00784-021-03888-7.
- [168] C. Schaudinn *et al.*, "Evaluation of a nonthermal plasma needle to eliminate ex vivo biofilms in root canals of extracted human teeth," *Int. Endodont. J.*, vol. 46, no. 10, pp. 930–937, 2013.
- [169] J. Pan *et al.*, "Cold plasma therapy of a tooth root canal infected with enterococcus faecalis biofilms in vitro," *J. Endodont.*, vol. 39, pp. 105–110, Jan. 2013.
- [170] T. Du *et al.*, "Effect of modified nonequilibrium plasma with chlorhexidine digluconate against endodontic biofilms in vitro," *J. Endodont.*, vol. 39, pp. 1438–1443, Nov. 2013.
- [171] M. Kury *et al.*, "Effects of violet radiation and nonthermal atmospheric plasma on the mineral contents of enamel during inoffice dental bleaching," *Photodiagnosis Photodyn. Ther.*, vol. 31, Sep. 2020, Art. no. 101848.
- [172] B. Celik, I. D. Capar, F. Ibis, N. Erdilek, and U. K. Ercan, "Deionized water can substitute common bleaching agents for nonvital tooth bleaching when treated with non-thermal atmospheric plasma," *J. Oral Sci.*, vol. 61, no. 1, pp. 103–110, 2019.
- [173] S. H. Nam, H. J. Lee, J. W. Hong, and G. C. Kim, "Efficacy of nonthermal atmospheric pressure plasma for tooth bleaching," *Sci. World J.*, vol. 2015, Jan. 2015, Art. no. 581731.
- [174] D. Claiborne, G. McCombs, M. Lemaster, M. A. Akman, and M. Laroussi, "Low-temperature atmospheric pressure plasma enhanced tooth whitening: The next-generation technology," *Int. J. Dental Hygiene*, vol. 12, pp. 108–114, May 2014.
- [175] H. W. Lee, S. H. Nam, A. A. H. Mohamed, G. C. Kim, and J. K. Lee, "Atmospheric pressure plasma jet composed of three electrodes: Application to tooth bleaching," *Plasma Processes Polym.*, vol. 7, nos. 3–4, pp. 274–280, 2010.
- [176] H. W. Lee, G. J. Kim, J. M. Kim, J. K. Park, J. K. Lee, and G. C. Kim, "Tooth bleaching with nonthermal atmospheric pressure plasma," *J. Endodont.*, vol. 35, no. 4, pp. 587–591, 2009.

- [177] J. N. Stasic *et al.*, "Effects of non-thermal atmospheric plasma treatment on dentin wetting and surface free energy for application of universal adhesives," *Clin. Oral Investig.*, vol. 23, no. 3, pp. 1383–1396, 2019.
- [178] A. Stancampiano *et al.*, "The effect of cold atmospheric plasma (CAP) treatment at the adhesive-root dentin interface," *J. Adhesive Dent.*, vol. 21, no. 3, pp. 229–237, 2019.
- [179] A.C. Ritts *et al.*, "Dentin surface treatment using a non-thermal argon plasma brush for interfacial bonding improvement in composite restoration," *Eur. J Oral Sci.*, vol. 118, no. 5, pp. 510–516, 2010.
- [180] Z. Zheng *et al.*, "Effects of novel non-thermal atmospheric plasma treatment of titanium on physical and biological improvements and in vivo osseointegration in rats," *Sci. Rep.*, vol. 10, Jun. 2020, Art. no. 10637.
- [181] E. Shaheen, A. Alhelwani, E. Van De Casteele, C. Politis, and R. Jacobs, "Evaluation of dimensional changes of 3D printed models after sterilization: A pilot study," *Open Dent. J.*, vol. 12, pp. 72–79, Jan. 2018.
- [182] Y. Wan, C. Tu, J. Yang, J. Bei, and S. Wang, "Influences of ammonia plasma treatment on modifying depth and degradation of poly (L-lactide) scaffolds," *Biomaterials*, vol. 27, no. 13, pp. 2699–2704, 2006.
- [183] S. H. Nam, B. B. R. Choi, and G. C. Kim, "The whitening effect and histological safety of nonthermal atmospheric plasma inducing tooth bleaching," *Int. J. Environ. Res. Public Health*, vol. 18, no. 9, p. 4714, 2021.
- [184] N. Jha, J. J. Ryu, E. H. Choi, and N. K. Kaushik, "Generation and role of reactive oxygen and nitrogen species induced by plasma, lasers, chemical agents, and other systems in dentistry," *Oxidative Med. Cell. Longevity*, vol. 2017, Oct. 2017, Art. no. 7542540.
- [185] M. Keidar, D. Yan, and J. H. Sherman, *Cold Plasma Cancer Therapy*. San Rafael, CA, USA: Morgan Calypool Publ., 2019.
- [186] H.-R. Metelmann *et al.*, "Clinical experience with cold plasma in the treatment of locally advanced dead and neck cancer," *Clin. Plasma Med.*, vol. 9, pp. 6–13, Mar. 2018.
  [187] M. Schuster *et al.*, "Visible tumor surface response to physical plasma
- [187] M. Schuster *et al.*, "Visible tumor surface response to physical plasma and apoptotic cell kill in head and neck cancer," *J. Cranio-Maxillofacial Surg.*, vol. 44, no. 9, pp. 1445–1452, 2016.
- [188] L. Bundscherer *et al.*, "Viability of human blood leukocytes compared with their respective cell lines after plasma treatment," *Plasma Med.*, vol. 3, nos. 1–2, pp. 71–80, 2013.
- [189] P.-M. Girard *et al.*, "Synergistic effect of H2O2 and NO2 in cell death induced by cold atmospheric He plasma," *Sci. Rep.*, vol. 6, no. 2, 2016, Art. no. 29098.
- [190] S. Bekeschus *et al.*, "Tumor cell metabolism correlates with resistance to gas plasma treatment: The evaluation of three dogmas," *Free Radical Biol. Med.*, vol. 167, no. 5, pp. 12–28, May 2021.
- [191] Y. Ma *et al.*, "Non-thermal atmospheric pressure plasma preferentially induces apoptosis in p53-mutated cancer cells by activating ROS stressresponse pathways," *PLoS ONE*, vol. 9, no. 4, 2014, Art. no. e91947.
- [192] H. Tanaka S. Bekeschus, D. Yan, M. Hori, M. Keidar, and M. Laroussi, "Plasma-treated solutions (PTS) in cancer therapy," *Cancers*, vol. 13, no. 7, p. 1737, 2021.
- [193] D. Yan, J. H. Sherman, and M. Keidar, "The application of the cold atmospheric plasma-activated solutions in cancer treatment," *Anticancer Agents Med. Chem.*, vol. 18, no. 6, p. 769, 2018.
- [194] H. Tanaka, K. Nakamura, M. Mizuno, K. Ishikawa, and K. Takeda, "Non-thermal atmospheric pressure plasma activates lactate in Ringer' s solution for anti-tumor effects," *Sci. Rep.*, vol. 6, no. 11, 2016, Art. no. 36282.
- [195] N. Kumar, J. H. Park, S. N. Jeon, B. S. Park, E. H. Choi, and P. Attri, "The action of microsecond-pulsed plasma-activated media on the inactivation of human lung cancer cells," *J. Phys. D. Appl. Phys.*, vol. 49, no. 11, 2016, Art. no. 115401.
- [196] B. R. Locke and K. Y. Shih, "Review of the methods to form hydrogen peroxide in electrical discharge plasma with liquid water," *Plasma Sources Sci. Technol.*, vol. 20, no. 3, 2011, Art. no. 034006.
- [197] S. Mohades, M. Laroussi, J. Sears, N. Barekzi, and H. Razavi, "Evaluation of the effects of a plasma activated medium on cancer cells," *Phys. Plasmas*, vol. 22, no. 12, 2015, Art. no. 122001.
- [198] E. Freund *et al.*, "Physical plasma-treated saline promotes an immunogenic phenotype in CT26 colon cancer cells in vitro and in vivo," *Sci. Rep.*, vol. 9, no. 1, p. 634, 2019.
- [199] H. Tanaka *et al.*, "Plasma-activated medium selectively kills glioblastoma brain tumor cells by down-regulating a survival signaling molecule, AKT kinase," *Plasma Med.*, vol. 1, nos. 3–4, pp. 265–277, 2011.

- [200] K. R. Liedtke *et al.*, "Non-thermal plasma-treated solution demonstrates antitumor activity against pancreatic cancer cells in vitro and in vivo," *Sci. Rep.*, vol. 7, no. 1, p. 8319, 2017.
- [201] D. Yan, L. Lin, J. H. Sherman, J. Canady, B. Trink, and M. Keidar, "The correlation between the cytotoxicity of cold atmospheric plasma and the extracellular H<sub>2</sub> O<sub>2</sub> -scavenging rate," *IEEE Trans. Radiat. Plasma Med. Sci.*, vol. 2, no. 6, pp. 618–623, Nov. 2018.
- [202] P. Wang et al., "Epithelial-to-mesenchymal transition enhances cancer cell sensitivity to cytotoxic effects of zcold atmospheric plasmas in breast and bladder cancer systems," *Cancers*, vol. 13, no. 12, p. 2889, 2021.
- [203] T. Adachi, H. Tanaka, S. Nonomura, H. Hara, S. I. Kondo, and M. Hori, "Plasma-activated medium induces A549 cell injury via a spiral apoptotic cascade involving the mitochondrial-nuclear network," *Free Radical Biol. Med.*, vol. 79, pp. 28–44, Feb. 2015.
- [204] D. Yan, N. Nourmohammadi, K. Bian, F. Murad, J. H. Sherman, and M. Keidar, "Stabilizing the cold plasma-stimulated medium by regulating medium's composition," *Sci. Rep.*, vol. 6, no. 4, 2016, Art. no. 26016.
- [205] D. Yan, N. Nourmohammadi, J. Milberg, and J. H. Sherman, "Guidelines for using 3-nitro-L-tyrosine as an antidegradation reagent of H2 O2 in the cold atmospheric plasma-stimulated solutions," *Plasma. Med.*, vol. 8, no. 2, pp. 121–130, 2018.
- [206] S. Mohades, N. Barekzi, H. Barekzi, V. Maruthamuthu, and M. Laroussi, "Temporal evaluation of the anti-tumor ef fi ciency of plasma-activated media," *Plasma Process. Polym.*, vol. 13, no. 2, p. 1206, 2016.
- [207] G. Fridman *et al.*, "Floating electrode dielectric barrier discharge plasma in air promoting apoptotic behavior in Melanoma skin cancer cell lines," *Plasma Chem. Plasma Process.*, vol. 27, no. 2, pp. 163–176, 2007.
- [208] C. H. Kim *et al.*, "Effects of atmospheric nonthermal plasma on invasion of colorectal cancer cells," *Appl. Phys. Lett.*, vol. 96, no. 24, pp. 2008–2011, 2010.
- [209] Y. Gorbanev, D. O'Connell, and V. Chechik, "Non-thermal plasma in contact with water: the origin of species," *Chem. Eur. J.*, vol. 22, no. 10, pp. 3496–3505, 2016.
- [210] A. Malyavko *et al.*, "Cold atmospheric plasma cancer treatment, direct versus indirect approaches," *Mater. Adv.*, vol. 1, no. 6, pp. 1494–1505, 2020.
- [211] F. Utsumi *et al.*, "Effect of indirect nonequilibrium atmospheric pressure plasma on anti-proliferative activity against chronic chemoresistant ovarian cancer cells in vitro and in vivo," *PLoS ONE*, vol. 8, no. 12, 2013, Art. no. e81576.
- [212] T. Adachi et al., "Iron stimulates plasma-activated medium-induced A549 cell injury," Sci. Rep., vol. 6, no. 2, 2016, Art. no. 20928.
- [213] J. Y. Kim, S. O. Kim, Y. Wei, and J. Li, "A flexible cold microplasma jet using biocompatible dielectric tubes for cancer therapy," *Appl. Phys. Lett.*, vol. 96, no. 20, pp. 19–22, 2010.
- [214] M. Ishaq, K. Bazaka, and K. Ostrikov, "Pro-apoptotic NOXA is implicated in atmospheric-pressure plasma-induced melanoma cell death," *J. Phys. D. Appl. Phys.*, vol. 48, no. 46, 2015, Art. no. 464002.
- [215] L. Shi et al., "Non-thermal plasma induces a stress response in mesothelioma cells resulting in increased endocytosis, lysosome biogenesis and autophagy," *Free Radic. Biol. Med.*, vol. 108, no. 4, pp. 904–917, 2017.
- [216] M. Adhikari, B. Adhikari, B. Ghimire, S. Baboota, and E. H. Choi, "Cold atmospheric plasma and silymarin nanoemulsion activate autophagy in human melanoma cells," *Int. J. Mol. Sci.*, vol. 21, no. 6, p. 1939, 2020.
- [217] D. Yan *et al.*, "The specific vulnerabilities of cancer cells to the cold atmospheric plasma-stimulated solutions," *Sci. Rep.*, vol. 7, no. 1, p. 4479, 2017.
- [218] F. Saadati, H. Mahdikia, H. Abbaszadeh, and M. Abdollahifar, "Comparison of direct and indirect cold atmospheric-pressure plasma methods in the B16 F10 melanoma cancer cells treatment," *Sci. Rep.*, vol. 8, no. 1, p. 7689, 2018.
- [219] D. Yan *et al.*, "The strong cell-based hydrogen peroxide generation triggered by cold atmospheric plasma," *Sci. Rep.*, vol. 7, no. 1, 2017, Art. no. 10831.
- [220] D. Yan, L. Lin, W. Xu, N. Nourmohammadi, and H. Jonathan, "Universality of micromolar-level cell-based hydrogen peroxide generation during direct cold atmospheric plasma treatment," *Plasma Med.*, vol. 8, no. 4, pp. 335–343, 2018.
- [221] D. Yan, W. Xu, X. Yao, L. Lin, J. H. Sherman, and M. Keidar, "The cell activation phenomena in the cold atmospheric plasma cancer treatment," *Sci. Rep.*, vol. 8, no. 1, 2018, Art. no. 15418.

- [222] W. Xu *et al.*, "The activation of cancer cells by a nanosecond-pulsed magnetic field generator," *J. Phys. D. Appl. Phys.*, vol. 53, no. 12, 2020, Art. no. 125401.
- [223] C. M. Wol, J. F. Kolb, K. Weltmann, T. Von Woedtke, and S. Bekeschus, "Combination treatment with cold physical plasma and pulsed electric fields augments ROS production and cytotoxicity in lymphoma," *Cancers*, vol. 12, no. 4, p. 845, 2020.
- [224] X. Yao *et al.*, "Sensitization of glioblastoma cells to temozolomide by a helium gas discharge tube," *Phys. Plasmas*, vol. 27, no. 11, 2020, Art. no. 114502.
- [225] S. K. Sagwal, G. Pasqual-Melo, Y. Bodnar, R. K. Gandhirajan, and S. Bekeschus, "Combination of chemotherapy and physical plasma elicits melanoma cell death via upregulation of SLC22A16," *Cell Death Dis.*, vol. 9, no. 12, p. 1179, 2018.
- [226] M. Vandamme *et al.*, "Antitumor effect of plasma treatment on u87 glioma xenografts: Preliminary results," *Plasma Process. Polym.*, vol. 7, nos. 3–4, pp. 264–273, 2010.
- [227] M. Vandamme *et al.*, "Response of human glioma U87 xenografted on mice to non thermal plasma treatment," *Plasma Med.*, vol. 1, no. 1, pp. 27–43, 2011.
- [228] M. Keidar *et al.*, "Cold plasma selectivity and the possibility of a paradigm shift in cancer therapy," *Brit. J. Cancer*, vol. 105, no. 9, pp. 1295–1301, 2011.
- [229] J. I. Ikeda, Y. Tsuruta, S. Nojima, H. Sakakita, M. Hori, and Y. Ikehara, "Anti-cancer effects of nonequilibrium atmospheric pressure plasma on cancer-initiating cells in human endometrioid adenocarcinoma cells," *Plasma Process. Polym.*, vol. 12, no. 12, pp. 1370–1376, 2015.
- [230] L. Brullé *et al.*, "Effects of a non thermal plasma treatment alone or in combination with gemcitabine in a MIA PaCa2-luc orthotopic pancreatic carcinoma model," *PLoS ONE*, vol. 7, no. 12, 2012, Art. no. e52653.
- [231] M. Vandamme *et al.*, "ROS implication in a new antitumor strategy based on non-thermal plasma," *Int. J. Cancer*, vol. 130, no. 9, pp. 2185–2194, 2012.
- [232] D. Yan *et al.*, "A physically triggered cell death via transbarrier cold atmospheric plasma cancer treatment," ACS Appl. Mater. Interfaces, vol. 12, no. 31, pp. 34548–34563, 2020.
- [233] Q. Wang et al., "A comparative study of cold atmospheric plasma treatment, chemical versus physical strategy," J. Phys. D. Appl. Phys., vol. 54, no. 9, 2020, Art. no. 095207.
- [234] D. Yan *et al.*, "The anti-glioblastoma effect of cold atmospheric plasma treatment: Physical pathway vs. chemical pathway," *Sci. Rep.*, vol. 10, no. 1, 2020, Art. no. 11788.
- [235] K. L. Rock and H. Kono, "The inflammatory response to cell death," Annu. Rev. Pathol. Dis., vol. 3, pp. 99–126, Feb. 2008.
- [236] P. Shaw *et al.*, "Evaluation of non-thermal effect of microwave radiation and its mode of action in bacterial cell inactivation," *Sci. Rep.*, vol. 11, no. 6, 2021, Art. no. 14003.
- [237] L. Galluzzi *et al.*, "Consensus guidelines for the definition, detection and interpretation of immunogenic cell death," *J. Immunother. Cancer*, vol. 8, no. 1, 2020, Art. no. e000337.
- [238] L. Galluzzi, A. Buqué, O. Kepp, L. Zitvogel, and G. Kroemer, "Immunogenic cell death in cancer and infectious disease," *Nat. Rev. Immunol.*, vol. 17, no. 2, pp. 97–111, 2021.
- [239] V. Miller, A. Lin, and A. Fridman, "Why target immune cells for plasma treatment of cancer," *Plasma Chem. Plasma Process.*, vol. 36, no. 1, pp. 259–268, 2016.
- [240] M. Khalili et al., "Non-thermal plasma-induced immunogenic cell death in cancer," J. Phys. D. Appl. Phys., vol. 52, no. 42, 2019, Art. no. 423001.
- [241] H. Mohamed, R. A. Esposito, M. A. Kutzler, B. Wigdahl, F. C. Krebs, and V. Miller, "Nonthermal plasma as part of a novel strategy for vaccination," *Plasma Process. Polym.*, vol. 17, no. 10, 2020, Art. no. e2000051.
- [242] R. Clemen and S. Bekeschus, "ROS cocktails as an adjuvant for personalized antitumor vaccination?" *Vaccines*, vol. 9, no. 5, p. 527, 2021.
- [243] A. Lin *et al.*, "Non-thermal plasma as a unique delivery system of shortlived reactive oxygen and nitrogen species for immunogenic cell death in melanoma cells," *Adv. Sci.*, vol. 6, no. 6, 2019, Art. no. 1802062.
- [244] A. Lin *et al.*, "Uniform nanosecond pulsed dielectric barrier discharge plasma enhances anti-tumor effects by induction of immunogenic cell death in tumors and stimulation of macrophages," *Plasma Process. Polym.*, vol. 12, no. 12, pp. 1392–1399, 2015.
- [245] J. Van Loenhout *et al.*, "Cold atmospheric plasma-treated PBS eliminates immunosuppressive pancreatic stellate cells and induces immunogenic cell death of pancreatic cancer cells," *Cancers*, vol. 11, no. 10, p. 1597, 2019.

- [246] L. Miebach *et al.*, "Tumor cytotoxicity and immunogenicity of a novel V-jet neon plasma source compared to the kINPen," *Sci. Rep.*, vol. 11, no. 1, p. 136, 2021.
- [247] S. Tomić *et al.*, "Plasma-activated medium potentiates the immunogenicity of tumor cell lysates for dendritic cell-based cancer vaccines," *Cancers*, vol. 13, no. 7, p. 1626, 2021.
- [248] K. Rödder *et al.*, "Activation of murine immune cells upon co-culture with plasma-treated B16F10 melanoma cells," *Appl. Sci.*, vol. 9, no. 4, p. 660, 2019.
- [249] H. Mahdikia *et al.*, "Gas plasma irradiation of breast cancers promotes immunogenicity, tumor reduction, and an abscopal effect in vivo," *Oncoimmunology*, vol. 10, no. 1, 2021, Art. no. 1859731.
- [250] A. G. Lin *et al.*, "Non-thermal plasma induces immunogenic cell death in vivo in murine CT26 colorectal tumors," *Oncoimmunology*, vol. 7, no. 9, 2018, Art. no. e1484978.
- [251] K. Mizuno, Y. Shirakawa, T. Sakamoto, H. Ishizaki, Y. Nishijima, and R. Ono, "Plasma-induced suppression of recurrent and reinoculated melanoma tumors in mice," *IEEE Trans. Radiat. Plasma Med. Sci.*, vol. 2, no. 4, pp. 353–359, Jul. 2018.
- [252] K. Mizuno, K. Yonetamari, Y. Shirakawa, T. Akiyama, and R. Ono, "Anti-tumor immune response induced by nanosecond pulsed streamer discharge in mice," *J. Phys. D. Appl. Phys.*, vol. 50, no. 12, 2017, Art. no. 12LT01.
- [253] F. Cheng, D. Yan, J. Chen, M. Keidar, and E. Sotomayor, "Cold plasma with immunomodulatory properties has significant anti-lymphoma activities in vitro and in vivo," *Blood*, vol. 134, p. 5307, Nov. 2019.
- [254] S. Bekeschus *et al.*, "The plasma-induced leukemia cell death is dictated by the ROS chemistry and the HO-1/CXCL8 axis," *IEEE Trans. Radiat. Plasma Med. Sci.*, vol. 5, no. 3, pp. 398–411, May 2021.
- [255] S. Bekeschus *et al.*, "Plasma treatment of ovarian cancer cells mitigates their immuno-modulatory products active on THP-1 monocytes," *Plasma*, vol. 1, no. 1, pp. 201–217, 2018.
- [256] S. Bekeschus *et al.*, "Argon plasma exposure augments costimulatory ligands and cytokine release in human monocyte-derived dendritic cells," *Int. J. Mol. Sci.*, vol. 22, no. 7, p. 3790, 2021.
- [257] H. Mohamed *et al.*, "Non-thermal plasma modulates cellular markers associated with immunogenicity in a model of latent HIV-1 infection," *PLoS One*, vol. 16, no. 3, 2021, Art. no. e0247125.
- [258] V. Miller, A. Lin, G. Fridman, D. Dobrynin, and A. Fridman, "Plasma stimulation of migration of macrophages," *Plasma Process. Polym.*, vol. 11, no. 12, pp. 1193–1197, 2014.
- [259] N. K. Kaushik *et al.*, "Cytotoxic macrophage-released tumour necrosis factor-alpha (TNF-α) as a killing mechanism for cancer cell death after cold plasma activation," *J. Phys. D. Appl. Phys.*, vol. 49, no. 8, 2016, Art. no. 084001.
- [260] C. Seebauer *et al.*, "Effects of cold physical plasma on oral lichen planus: An in vitro study (effects of CAP on OLP)," *Oral Dis.*, vol. 27, no. 7, pp. 1728–1737, 2021.
- [261] M. Schuster *et al.*, "Side effects in cold plasma treatment of advanced oral cancer—Clinical data and biological interpretation," *Clin. Plasma Med.*, vol. 10, pp. 9–15, Jun. 2018.
- [262] K. R. Liedtke *et al.*, "Gas plasma-conditioned Ringer's lactate enhances the cytotoxic activity of cisplatin and genetiabine in pancreatic cancer in vitro and in ovo," *Cancers*, vol. 12, no. 1, p. 123, 2020.
- [263] J. Winter, T. M. C. Nishime, S. Glitsch, H. Lühder, and K. D. Weltmann, "On the development of a deployable cold plasma endoscope," *Contrib. Plasma Phys.*, vol. 58, no. 5, pp. 404–414, 2018.
- [264] JCRI-ABTS and USMI Successfully Complete Phase 1 Multi-Center Clinical Trial Using Canady Helios<sup>TM</sup> Cold Plasma for the Treatment of Cancer. [Online]. Available: https://www.businesswire.com/news/ home/20210420005842/en (Accessed: Apr. 20, 2021).
- [265] S. Bekeschus, A. Schmidt, F. Niessner, T. Gerling, K. D. Weltmann, and K. Wende, "Basic research in plasma medicine—A throughput approach from liquids to cells," *J. Vis. Experiments*, vol. 2017, no. 129, pp. 1–9, 2017.
- [266] M. Keidar, "A prospectus on innovations in the plasma treatment of cancer," *Phys. Plasmas*, vol. 25, no. 8, 2018, Art. no. 083504.
- [267] M. Keidar, D. Yan, I. I. Beilis, B. Trink, and J. H. Sherman, "Plasmas for treating cancer: Opportunities for adaptive and selfadaptive approaches," *Trends Biotechnol.*, vol. 36, no. 6, pp. 586–593, 2018.
- [268] L. Lin, D. Yan, E. Gjika, J. H. Sherman, and M. Keidar, "Atmospheric plasma meets cell: Plasma tailoring by living cells," ACS Appl. Mater. Interfaces, vol. 11, no. 34, pp. 30621–30630, 2019.

- [269] Y. Lyu, L. Lin, E. Gjika, T. Lee, and M. Keidar, "Mathematical modeling and control for cancer treatment with cold atmospheric plasma jet," *J. Phys. D. Appl. Phys.*, vol. 52, no. 18, 2019, Art. no. 185202.
- [270] N. Y. Babaeva, N. Ning, D. B. Graves, and M. J. Kushner, "Ion activation energy delivered to wounds by atmospheric pressure dielectric-barrier discharges: Sputtering of lipid-like surfaces," *J. Phys. D Appl. Phys.*, vol. 45, no. 11, 2012, Art. no. 115203, doi: 10.1088/0022-3727/45/11/115203.
- [271] A. M. Lietz, and M. J. Kushner, "Air plasma treatment of liquid covered tissue: Long timescale chemistry," J. Phys. D Appl. Phys., vol. 49, no. 42, 2016, Art. no. 425204, doi: 10.1088/0022-3727/49/42/425204.
- [272] D. Liu *et al.*, "Aqueous reactive species induced by a surface air discharge: Heterogeneous mass transfer and liquid chemistry pathways," *Sci. Rep.*, vol. 6, no. 1, 2016, Art. no. 23737, doi: 10.1038/srep23737.
- [273] M. Yusupov, K. Wende, S. Kupsch, E. C. Neyts, S. Reuter, and A. Bogaerts, "Effect of head group and lipid tail oxidation in the cell membrane revealed through integrated simulations and experiments," *Sci. Rep.*, vol. 7, no. 1, p. 5761, 2017, doi: 10.1038/s41598-017-06412-8.
- [274] M. Yusupov, E. C. Neyts, U. Khalilov, R. Snoeckx, A. C. T. van Duin, and A. Bogaerts, "Atomic-scale simulations of reactive oxygen plasma species interacting with bacterial cell walls," *New J. Phys.*, vol. 14, no. 9, 2012, Art. no. 093043, doi: 10.1088/1367-2630/14/9/093043.
- [275] C. C. W. Verlackt *et al.*, "Atomic-scale insight into the interactions between hydroxyl radicals and DNA in solution using the ReaxFF reactive force field," *New J. Phys.*, vol. 17, no. 10, 2015, Art. no. 103005, doi: 10.1088/1367-2630/17/10/103005.
- [276] R. M. Cordeiro, "Reactive oxygen species at phospholipid bilayers: Distribution, mobility and permeation," *Biochimica Biophysica Acta Biomembranes*, vol. 1838, no. 1, pp. 438–444, 2014, doi: 10.1016/j.bbamem.2013.09.016.
- [277] J. Razzokov, M. Yusupov, R. M. Cordeiro, and A. Bogaerts, "Atomic scale understanding of the permeation of plasma species across native and oxidized membranes," *J. Phys. D Appl. Phys.*, vol. 51, no. 36, 2018, Art. no. 365203, doi: 10.1088/1361-6463/aad524.
- [278] M. N. Möller, J. R. Lancaster, and A. Denicola, "Chapter 2 the interaction of reactive oxygen and nitrogen species with membranes," in *Current Topics in Membranes*, S. Matalon, Ed. Amsterdam, The Netherlands: Academic, 2008, pp. 23–42, doi: 10.1016/S1063-5823(08)00202-0.
- [279] M. Yusupov, D. Yan, R. M. Cordeiro, and A. Bogaerts, "Atomic scale simulation of H2O2 permeation through aquaporin: Toward the understanding of plasma cancer treatment," *J. Phys. D Appl. Phys.*, vol. 51, no. 12, 2018, Art. no. 125401, doi: 10.1088/1361-6463/aaae7a.
- [280] J. Van der Paal, E. C. Neyts, C. C. W. Verlackt, and A. Bogaerts, "Effect of lipid peroxidation on membrane permeability of cancer and normal cells subjected to oxidative stress," *Chem. Sci.*, vol. 7, no. 1, pp. 489–498, 2016, doi: 10.1039/C5SC02311D.
- [281] M. Yusupov, J. Van der Paal, E. C. Neyts, and A. Bogaerts, "Synergistic effect of electric field and lipid oxidation on the permeability of cell membranes," *Biochimica Biophysica Acta Gen. Subjects*, vol. 1861, no. 4, pp. 839–847, 2017, doi: 10.1016/j.bbagen.2017.01.030.
- [282] J. Van der Paal, C. Verheyen, E. C. Neyts, and A. Bogaerts, "Hampering effect of cholesterol on the permeation of reactive oxygen species through phospholipids bilayer: Possible explanation for plasma cancer selectivity," *Sci. Rep.*, vol. 7, no. 1, 2017, Art. no. 39526, doi: 10.1038/srep39526.
- [283] M. Yusupov, J.-W. Lackmann, J. Razzokov, S. Kumar, K. Stapelmann, and A. Bogaerts, "Impact of plasma oxidation on structural features of human epidermal growth factor," *Plasma Processes Polym.*, vol. 15, no. 8, 2018, Art. no. 1800022, doi: 10.1002/ppap.201800022.

- [284] R. P. Magalhães, H. S. Fernandes, and S. F. Sousa, "Modelling enzymatic mechanisms with QM/MM approaches: Current status and future challenges," *Israel J. Chem.*, vol. 60, no. 7, pp. 655–666, 2020, doi: 10.1002/ijch.202000014.
- [285] Q. Liao, "Enhanced sampling and free energy calculations for protein simulations," *Progr. Mol. Biol. Transl. Sci.*, vol. 170, pp. 177–213, 2020, doi: 10.1016/bs.pmbts.2020.01.006.
- [286] J.-W. Lackmann *et al.*, "Photons and particles emitted from cold atmospheric-pressure plasma inactivate bacteria and biomolecules independently and synergistically," *J. Royal Soc. Interface*, vol. 10, no. 89, 2013, Art. no. 20130591, doi: 10.1098/rsif.2013.0591.
- [287] E. A. J. Bartis *et al.*, "Deactivation of lipopolysaccharide by Ar and H2 inductively coupled low-pressure plasma," *J. Phys. D Appl. Phys.*, vol. 47, no. 4, 2014, Art. no. 045202, doi: 10.1088/0022-3727/47/4/045202.
- [288] J. Van der Paal *et al.*, "How membrane lipids influence plasma delivery of reactive oxygen species into cells and subsequent DNA damage: An experimental and computational study," *Phys. Chem. Chem. Phys.*, vol. 21, no. 35, pp. 19327–19341, 2019, doi: 10.1039/C9CP03520F.
- [289] C. K. Haluska, M. S. Baptista, A. U. Fernandes, A. P. Schroder, C. M. Marques, and R. Itri, "Photo-activated phase separation in giant vesicles made from different lipid mixtures," *Biochimica Biophysica Acta Biomembranes*, vol. 1818, no. 3, pp. 666–672, 2012, doi: 10.1016/j.bbamem.2011.11.025.
- [290] R. M. Cordeiro, "Molecular structure and permeability at the interface between phase-separated membrane domains," J. Phys. Chem. B, vol. 122, no. 27, pp. 6954–6965, 2018, doi: 10.1021/acs.jpcb.8b03406.
- [291] D. E. Scott, A. R. Bayly, C. Abell, and J. Skidmore, "Small molecules, big targets: Drug discovery faces the protein–protein interaction challenge," *Nat. Rev. Drug Disc.*, vol. 15, no. 8, pp. 533–550, 2016, doi: 10.1038/nrd.2016.29.
- [292] J. Y. Lee *et al.*, "Structural basis of checkpoint blockade by monoclonal antibodies in cancer immunotherapy," *Nat. Commun.*, vol. 7, no. 1, 2016, Art. no. 13354, doi: 10.1038/ncomms13354.
- [293] A. Lin *et al.*, "Oxidation of innate immune checkpoint CD47 on cancer cells with non-thermal plasma," *Cancers*, vol. 13, no. 3, p. 579, 2021, doi: 10.3390/cancers13030579.
- [294] P. Shaw, N. Kumar, D. Hammerschmid, A. Privat-Maldonado, S. Dewilde, and A. Bogaerts, "Synergistic effects of melittin and plasma treatment: A promising approach for cancer therapy," *Cancers*, vol. 11, no. 8, p. 1109, 2019, doi: 10.3390/cancers11081109.
- [295] M. Yusupov et al., "Oxidative damage to hyaluronan–CD44 interactions as an underlying mechanism of action of oxidative stress-inducing cancer therapy," *Redox Biol.*, vol. 43, Jul. 2021, Art. no. 101968, doi: 10.1016/j.redox.2021.101968.
- [296] G. Bauer and D. B. Graves, "Mechanisms of selective antitumor action of cold atmospheric plasma-derived reactive oxygen and nitrogen species," *Plasma Processes Polym.*, vol. 13, no. 12, pp. 1157–1178, 2016, doi: 10.1002/ppap.201600089.
- [297] D. T. Erudaitius, G. R. Buettner, and V. G. J. Rodgers, "The latency of peroxisomal catalase in terms of effectiveness factor for pancreatic and glioblastoma cancer cell lines in the presence of high concentrations of H2O2: Implications for the use of pharmacological ascorbate in cancer therapy," *Free Radical Biol. Med.*, vol. 156, pp. 20–25, Aug. 2020, doi: 10.1016/j.freeradbiomed.2020.05.023.
- [298] C. Bengtson, and A. Bogaerts, "On the anti-cancer effect of cold atmospheric plasma and the possible role of catalase-dependent apoptotic pathways," *Cells*, vol. 9, no. 10, p. 33, 2020, doi: 10.3390/cells9102330.
- [299] C. Bengtson and A. Bogaerts, "The quest to quantify selective and synergistic effects of plasma for cancer treatment: Insights from mathematical modeling," *Int. J. Mol. Sci.*, vol. 22, no. 9, p. 5033, 2021, doi: 10.3390/ijms22095033.